



Proposed Regulatory Decision Document PRDD2003-05

Kresoxim-methyl Technical and Sovran[®] Fungicide

The active ingredient kresoxim-methyl and the formulated end-use product Sovran[®] Fungicide, for the control of apple scab and powdery mildew in apple orchards in Canada, are proposed for full registration under Section 13 of the Pest Control Products (PCP) Regulations.

This Proposed Regulatory Decision Document (PRDD) provides a summary of data reviewed and the rationale for the proposed full registration of these products. The Pest Management Regulatory Agency (PMRA) will accept written comments on this proposal up to 45 days from the date of publication of this document. Please forward all comments to the Publications Coordinator at the address below.

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Foreword

The submission for full registration of the active ingredient kresoxim-methyl and the end-use product Sovran[®] Fungicide, a fungicide developed by BASF Canada Inc. for control of apple scab and powdery mildew in apple orchards, has been reviewed by the Pest Management Regulatory Agency (PMRA) of Health Canada.

The PMRA had previously issued a temporary registration (Regulatory Note REG2000-14) for these products with the requirement that BASF Canada Inc. carry out additional environmental and occupational exposure studies. These studies have now been completed.

The PMRA has carried out an assessment of available information in accordance with Section 9 of the Pest Control Products (PCP) Regulations and has found it sufficient pursuant to Section 18(*b*), to allow a determination of the safety, merit and value of the active ingredient kresoxim-methyl and the end-use product Sovran[®] Fungicide. The Agency has concluded that the use of the active ingredient kresoxim-methyl and the end-use product Sovran[®] Fungicide in accordance with the label has merit and value consistent with Section 18(*c*) of the PCP Regulations and does not entail an unacceptable risk of harm pursuant to Section 18(*d*). Based on the considerations outlined above, therefore, the use of the active ingredient kresoxim-methyl and the end-use product Sovran[®] Fungicide, for the control of apple scab and powdery mildew in apple orchards, is proposed for full registration, pursuant to Section 13 of the PCP Regulations.

Methods for analyzing kresoxim-methyl residues in various environmental media can be provided to monitoring agencies and research institutions upon request to the PMRA.

The PMRA will accept written comments on this proposal up to 45 days from the date of publication of this document to allow interested parties an opportunity to provide input into the proposed registration decision for this product.

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1.0 The active substance, its properties, and uses

Kresoxim-methyl belongs to the Quinone Outside Inhibitor (QoI) class of fungicide. It inhibits mitochondrial respiration by blocking electron transfer at the bc_1 complex in fungi. The protective effect is due to the inhibition of both spore germination and host infection. The curative and eradicated effects are the result of kresoxim-methyl's inhibition of mycelial growth and sporulation.

The commercial formulation, Sovran[®] Fungicide, is a foliar fungicide for use on apples to control apple scab and powdery mildew. It provides commercially acceptable control of scab and powdery mildew when applied at 90–180 g active ingredient (a.i.)/hectare (ha) and 120–225 g a.i./ha, respectively. A maximum of four applications per season is recommended.

1.1 Identity of the active substance and impurities

Active substance: Kresoxim-methyl

Function: Fungicide

Chemical name:
(International Union of Pure
and Applied Chemistry): Methyl (E)-2-methoxyimino-2-[2-(*o*-tolylloxymethyl)
phenyl]acetate

Chemical name (Chemical
Abstracts Service [CAS]): α -(methoxyimino)-2-[(2-
methylphenoxy)methyl]benzeneacetic acid methyl ester

CAS number: 143390-89-0

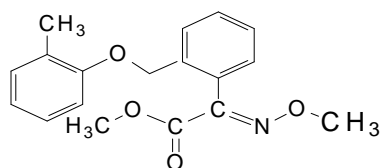
Nominal purity of active: 94.0%

Identity of relevant
impurities of toxicological,
environmental and other
significance: The technical material is not known to contain any toxic
microcontaminants identified as Track-1 substances in the
Toxic Substances Management Policy (TSMP)

Molecular formula: $C_{18}H_{19}NO_4$

Molecular mass: 313.36

Structural formula:



1.2 Physical and chemical properties of active substance and end-use product

Table 1.2.1 Technical product: Kresoxim-methyl

Property	Result	Comment																		
Colour and physical state	Pure active ingredient (PAI): white crystals Technical grade active ingredient (TGAI): light-brown powder																			
Odour	Odourless																			
Melting point and range	PAI: 97.2–101.7°C TGAI: 98–100°C																			
Density	PAI: 1.258 g/cm ³ at 20°C																			
Vapour pressure	TGAI: 2.3×10^{-6} Pa at 20°C (by extrapolation)	The active ingredient will be non-volatile under field conditions.																		
Henry's Law constant	3.6×10^{-4} Pa·m ³ ·mol ⁻¹	Will not volatilize from moist soil and water surfaces																		
UV and visible spectrum at 26°C	λ_{max} at 204 nanometres (nm), no absorption at $\lambda > 350$ nm	Minimal phototransformation is expected																		
Solubility in water at 20°C	PAI: 2.00 ± 0.08 mg/L	Low solubility																		
Solubility (g/100 mL) in organic solvents at 20°C	<table border="1"> <thead> <tr> <th>Solvent</th> <th>Solubility</th> </tr> </thead> <tbody> <tr> <td><i>n</i>-heptane</td> <td>0.172</td> </tr> <tr> <td>toluene</td> <td>11.1</td> </tr> <tr> <td>CH₂Cl₂</td> <td>93.9</td> </tr> <tr> <td>methanol</td> <td>1.46</td> </tr> <tr> <td>acetone</td> <td>2.17</td> </tr> <tr> <td>ethyl acetate</td> <td>12.3</td> </tr> <tr> <td>acetonitrile</td> <td>16.6</td> </tr> <tr> <td><i>i</i>-propanol</td> <td>0.480</td> </tr> </tbody> </table>	Solvent	Solubility	<i>n</i> -heptane	0.172	toluene	11.1	CH ₂ Cl ₂	93.9	methanol	1.46	acetone	2.17	ethyl acetate	12.3	acetonitrile	16.6	<i>i</i> -propanol	0.480	
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acetonitrile	16.6																			
<i>i</i> -propanol	0.480																			
<i>n</i> -Octanol–water partition coefficient (K_{ow})	$\log K_{\text{ow}} = 3.4 \pm 0.02$	Potential for bioaccumulation																		
Dissociation constant (pK_{a})	No pK_{a} value at pH 2–12	Does not dissociate																		
Oxidizing properties	Stable at 54°C for 14 days Compound contains no moiety that could exert oxidizing properties																			
Storage stability	Not applicable to the technical product																			

Table 1.2.2 End-use product: Sovran® Fungicide

Property	Result
Physical state	Granular powder
Formulation type	Wettable granules
Guarantee	50%

1.3 Details of Uses

This technical active ingredient and end-use product are fungicides submitted under Use Site Category # 14 (Terrestrial Food Crops). The proposed use pattern for Sovran™ is as a control of powdery mildew (*Podosphaera leucontricha*) and apple scab (*Venturia inaequalis*) in apple orchards in Ontario, Quebec, British Columbia, and the Maritime Provinces. There are two sources of the TGAI, which are guaranteed at 94% and 97% kresoxim-methyl. The water-dispersible granular formulation, Sovran, contains 50% kresoxim-methyl plus several formulants, which fall under the U.S. EPA Inert List 4B (compounds in List 4B may be acutely toxic but under the current use pattern (quantity and use) they have been determined to pose no threat; new use patterns may increase the risk; the environmental fate and ecological effect may need to be determined).

Sovran should be applied with ground equipment (generally via airblast) at an application rate of 180–360 g EP/ha for apple scab and a rate of 240–450 g EP/ha for powdery mildew. To control apple scab, applications should be made when the trees are at the 12 mm (½ inch) green stage of flower bud development or when environmental conditions become favourable for primary scab. For powdery mildew control, applications are initiated when the flower buds are at the 12-mm green stage. For both diseases, applications should be redone every 10–14 days through second cover.

To limit resistance development, Sovran should be alternated with a fungicide with a different mode of action. No more than two sequential applications of Sovran should be made before another fungicide is applied and no more than four applications of Sovran should be made per season.

The maximum application rate of 450 g EP/ha for the control of powdery mildew is equivalent to an application of 225 g a.i./ha.

2.0 Methods of analysis

2.1 Method for analysis of the active substance as manufactured

A reversed phase—high performance liquid chromatographic (RP–HPLC) method was used for the determination of the active substance and significant impurities (content ≥ 0.1%) in the technical product. The method has been shown to have satisfactory specificity, linearity, precision and accuracy.

2.2 Method for formulation analysis

An isothermal capillary gas chromatographic (GC) method was used for the determination of active substance in the formulation. The method has been shown to have satisfactory specificity, linearity, precision and accuracy.

2.3 Methods for residue analysis

2.3.1 Multiresidue methods for residue analysis

Existing multiresidue methods (MRM) of analysis were found to be suitable for the determination of residues of kresoxim-methyl and its metabolites in fatty (ground beef) and nonfatty (grapes) matrices. Since the MRM analyses for all components of the residues of concern (ROC) in both plant and animal commodities, it may be considered as a singular enforcement method compared with separate methods for plant (350/3-US) and animal (354/1-US and 354/2) matrices (see sections 2.3.2 and 2.3.3). In contrast to method 350/3-US, the MRM also analyses for the parent compound and the metabolite 490M1 separately. Although methods 350/3-US and 354/1-US were also accepted as enforcement methods, method 354/2 was not accepted. Therefore, the submitted MRM is the only method currently accepted for enforcement analyses in meat and meat by-products.

2.3.2 Methods for residue analysis of plants and plant products

The ROC was defined from the apple metabolism studies as the parent compound (kresoxim-methyl) and the metabolites 490M1, 490M2 (free and glucose conjugated) and 490M9 (free and glucose conjugated).

Residues of concern were determined by HPLC with UV detection (270 nm) in apples and apple juice, grapes and its processed commodities (wine, must and marc) and pecans. The analytical method reduces the number of analytes to three by enzymatic cleavage of the glycosides of the metabolites and hydrolysis of kresoxim-methyl to 490M1. Kresoxim-methyl is calculated as 490M1 equivalents. The method limit of quantitation (LOQ) for each analyte in all matrices was reported to be 0.05 parts per million (ppm) and thus 0.15 ppm for all three analytes combined. This method was found to give good recoveries for the analysis of apples (81–105%), apple juice (74–98%), grapes (70–105%), processed grape commodities (76–100%) and pecans (90–130%). The standard deviations measured following spiking at 0.05–5 ppm were indicative of the method having good to satisfactory repeatability. Representative chromatograms of control and spiked samples of apples and grapes showed no interferences from matrix coextractives, or from reagents, solvents and glassware. Good linearity (correlation coefficient, $r = 0.99$ for 490M1, 0.99 for 490M2 and 0.99–1.0 for 490M9) was observed in the range of 0.05–125 µg/mL for the ROC. The interlaboratory validations (ILVs) supported the reliability and reproducibility of the BASF Canada Inc. method for the determination of ROC in apples, grapes and pecans.

2.3.3 Methods for residue analysis of food of animal origin

The ROC for kresoxim-methyl was defined from the lactating goat metabolism studies as kresoxim-methyl and the metabolites BF 490-1, BF 490-2 and BF 490-9 in ruminant tissues and milk.

The petitioner is not proposing a common moiety method. Method 354/1-US (or method 354/1) is proposed to quantify BF 490-9 and BF 490-2 in milk. Method 354/2 is proposed to quantify BF 490-1 and BF 490-9 in liver, BF 490-1 and BF 490-2 in muscle, and BF 490-1, BF 490-2 and BF 490-9 in kidney and fat. Methods 354/1-US and 354/2 determine residues by HPLC with UV detection (270 nm). None of these methods analyses for the parent compound.

The Pest Management Regulatory Agency (PMRA) evaluated the information provided for method 354/1 and found that it is valid. This method was found to give good recoveries for the analysis of whole milk (78–107% for BF 490-2 and 67–118% for BF 490-9). Representative chromatograms of control samples showed interferences for BF 490-2 at the 0.001 ppm fortification level. Because of this interference, the LOQ for both analytes was revised to 0.004 ppm (0.002 ppm for each analyte). The limit of detection (LOD) was determined at 0.002 ppm. Chromatographic peaks for each analyte were well defined and symmetrical. Good linearity ($r = 0.997$ – 0.998) was observed in the range of 0.002–1 ppm for both analytes. Method 354/1-US analyses for the most predominant metabolites in milk, BF 490-9 and BF 490-2, and is recommended as the enforcement method for analyses of milk from animals treated orally with kresoxim-methyl.

The PMRA evaluated the information provided for method 354/2 and found that it is valid for analyses in muscle and liver tissues, but not for analyses in kidney and fat tissues. Recoveries were not always within guideline requirements in kidney and fat samples, especially at the stated LOQ of 0.01 ppm for each analyte. Mean recoveries in all tissues were 69–91% for BF 490-1, 81–93% for BF 490-2 and 83–96% for BF 490-9. In two different validation studies, however, recoveries were beyond the acceptable range of 70–130%, and accuracy was beyond the acceptable range of $\pm 20\%$ in fat samples ($75 \pm 15\%$, coefficient of variation [CV] = 19% for BF 490-1; $81 \pm 20\%$, CV = 24% for BF 490-2; and $96 \pm 26\%$, CV = 27% for BF 490-9 in one study; in the other study, recoveries were $69 \pm 13\%$, CV = 19% for BF 490-1 [$67 \pm 20\%$, CV = 30% at the 0.01 ppm spiking level] and values for BF 490-2 and BF 490-9 were within acceptable ranges). Although the authors argued that detection of BF 490-1 was more important than detection of BF 490-2 and BF 490-9, as evidenced by a cow feeding study, the relatively poor recoveries for BF 490-1 in fat analyses from the two different validation studies does not support the argument for acceptance of this method for fat samples. In one of the two ILVs, kidney analyses indicated that recoveries were slightly beyond the acceptable range at the 0.01 ppm spiking level for all analytes ($69 \pm 1.3\%$ for BF 490-1, $69 \pm 7\%$ for BF 490-2 and $70 \pm 9\%$ for BF 490-9), and the overall accuracy for BF 490-2 analyses (CV = 30%) was beyond the accepted range of 20%. Three trials were attempted before

the author of the ILV accepted these results, on the basis of acceptance of values slightly beyond acceptable ranges. These results were not accepted by the PMRA because they were not within acceptable ranges at the stated LOQ of 0.01 ppm and also because of a lack of overall accuracy for the analysis of BF 490-2. It appears that analyses of analytes in kidney and fat at the 0.01 ppm spiking level are problematical, especially when conducted under good laboratory practice conditions.

Generally, control chromatograms were free from interferences for all analytes in four studies. The LOD was determined to be 0.006 ppm in all tissues tested. Chromatographic peaks for each analyte were well defined and symmetrical. Relatively good linearity ($r = 0.994-0.999$) was observed in the range of 0.002–1 ppm for all analytes.

2.3.4 Analytical methodology for environmental substrates

In soils, one method employing sequential solvent extraction followed by liquid chromatography (LC) and mass spectrometry (MS) (LC/MS/MS) (BASF method no. D9503) has been successfully used to identify and quantitate kresoxim-methyl and the transformation products BF 490-1 and BF 490-5. The LOQ using this method is 0.01 mg/kg soil (0.01 ppm). A second method using sequential solvent extraction followed by GC and electron capture detection (ECD) (BASF method no. D9603) is capable of quantitating kresoxim-methyl and BF 490-1 at 0.5 mg/kg soil (0.5 ppm). BAS 490 F (kresoxim-methyl) and BF 490-1 would be expected to remain stable in soil stored frozen for a period of up to two years.

BAS 490 F and BF 490-1 were determined in aquatic media containing algae and duckweed mixed with different nutrient media (BASF method no. D9209). The method used solid phase extraction to remove residues of BAS 490 F and BF 490-1 from the water followed by GC/ECD. The LOQ was 25 µg test material/L (25 parts per billion [ppb]).

3.0 Impact on human and animal health

3.1 Integrated toxicological summary (see also toxicology summary tables in Appendix I)

A detailed review of the toxicological database available for the new fungicide kresoxim-methyl has been completed. The data submitted were largely complete and comprehensive. Data evaluation records from the U.S. EPA were available for most of the individual study reviews and were utilized as much as possible in this review. A document prepared by the Joint WHO/FAO Meeting on Pesticide Residues (JMPR) (1998) was also available and was utilized to fill in data gaps where any existed. In general, the studies were well conducted and in conformity with currently acceptable international guidelines and protocols.

Metabolism studies in Wistar rats showed technical kresoxim-methyl to be moderately absorbed, then widely distributed and quickly eliminated with no tissue bioaccumulation. After 96 hours (h), only residual radioactivity was detectable in gastrointestinal content and in or on skin (females only). Major routes of elimination in both sexes were through urine (9–33%) and feces (66–81% [35–43% through bile]) and none through exhaled air. Following oral administration, high proportions (73% of dose) of unchanged parent compound were found in feces and none in bile. Systemically available kresoxim-methyl was rapidly and completely metabolized to a total of 32 (major and minor) metabolites, which were identified in urine, feces, bile, plasma and kidneys of rats. The alcohol–acid and phenol–acid of the parent compound and their glucuronides were the predominant final biotransformation products.

Studies of acute toxicity following single dosing showed technical kresoxim-methyl to be of low acute toxicity by the oral, dermal and inhalation routes. Primary eye irritation studies in rabbits showed technical kresoxim-methyl to be a mild eye irritant, while primary dermal irritation studies in rabbits showed that technical kresoxim-methyl was not a dermal irritant. Kresoxim-methyl was not a dermal sensitizer when tested in Guinea pigs according to the maximization test of Magnusson and Kligman. The Sovran[®] formulation (kresoxim-methyl 48.4%) was of low acute toxicity by the oral, dermal and inhalation routes. Eye irritation studies in rabbits showed that Sovran[®] was a mild eye irritant, while dermal irritation studies in rabbits indicated that Sovran[®] was not a dermal irritant. A dermal sensitization study in Guinea pigs showed that Sovran[®] was not a dermal sensitizer.

Short-term toxicity studies in rodents indicated the liver as the target organ, with increases in serum γ -glutamyl transferase (SGGT) occurring in rats at 8000 ppm (577 mg/kg body weight [bw] per day [d]) and an increase in relative (to body weight) liver weight manifesting in mice at 4000 ppm (909 mg/kg bw/d). A one-year feeding study in beagle dogs, however, failed to reveal liver toxicity at doses as high as 25 000 ppm (714 mg/kg bw/d). The no observed adverse effect levels (NOAELs) in subchronic studies were mice, 1000 and 8000 ppm (230 and 258 mg/kg bw/d) for males and females, respectively; rats, 2000 and 16 000 ppm (146 and 1374 mg/kg bw/d) for males and females, respectively; and dogs, 5000 and 25 000 ppm (138 and 761 mg/kg bw/d) in males and females, respectively.

In the chronic toxicity study in rats, liver toxicity manifested, beginning at 8000 ppm, as increases in liver weight and SGGT levels, the presence of gross and microscopic lesions including cysts, masses, eosinophilic cell foci, mixed cell foci and hypertrophy, and an increase in the incidence of hepatocellular carcinomas in both sexes. In a two-year oncogenicity (feeding) study in rats, once again, at 8000 ppm and above, there were treatment related liver lesions including cysts, masses, eosinophilic cell foci, mixed cell foci, hypertrophy, bile duct hyperplasia and cholangiofibrosis, as well as an increase in the incidence of hepatocellular carcinomas in both sexes. The NOAELs in chronic rat studies were 800 ppm (36 and 47 mg/kg bw/d in males and females, respectively). Supplementary hepatic cell proliferation studies in young adult and fully mature rats

showed that treatment with kresoxim-methyl for three weeks induced a 2- to 3-fold increase in hepatocyte proliferation at 16 000 ppm (1140 mg/kg bw/d), with no hepatocyte proliferation (NOAEL) observed at 200 ppm (15 mg/kg bw/d).

In a carcinogenicity study in mice, the NOAEL was 2000 ppm (304 mg/kg bw/d) on the basis of decreased body weight and microscopic liver lesions observed at 8000 ppm (1305 mg/kg bw/d), but there was no evidence of oncogenicity at the highest dose of 8000 ppm (1600 mg/kg bw/d). That the test substance was not carcinogenic in mice suggests that the liver tumour promotion effect observed in the long-term rat studies was rat specific.

Kresoxim-methyl was not mutagenic in either the Ames test using *Escherichia coli* and *Salmonella typhimurium* or in a gene mutation assay with Chinese hamster ovary cells, nor was it clastogenic in a chromosomal aberration assay in primary human lymphocyte cultures or an in vivo mouse micronucleus assay. It also did not cause unscheduled deoxyribonucleic acid (DNA) synthesis in primary rat hepatocytes or unscheduled DNA synthesis ex vivo in rat hepatocytes. These results indicate that kresoxim-methyl is not genotoxic.

Kresoxim-methyl was not teratogenic in rats and rabbits at doses of up to 1000 mg/kg bw/d and did not cause reproductive toxicity in rats fed doses of up to 16 000 ppm (1625 mg/kg bw/d). The NOAEL for developmental toxicity in rats was 1000 ppm (100 mg/kg bw/d) on the basis of reductions in body weight and body weight gain in F_{1b} and F₂ pups and delays of developmental landmarks in F_{1b} and F₂ pups occurring at 4000 ppm. Kresoxim-methyl showed no evidence of neurotoxicity in rats following either acute or subchronic exposure.

Mechanistic studies on induction of liver tumours (S-phase response) in rats, following a single oral gavage dose or three-week feeding, showed that kresoxim-methyl induced an increased cell proliferation (S-phase response) in liver cells after short-term administration. The S-phase induction was reversible within the recovery period.

3.2 Determination of acceptable daily intake (ADI)

3.2.1 Non-carcinogenic endpoints

The recommended acceptable daily intake (ADI) for kresoxim-methyl is 0.36 mg/kg bw/d. The most appropriate study for selection of a toxicity end point for chronic dietary exposure was the two-year dietary study in rats with a NOAEL of 36 mg/kg bw/d, on the basis of body weight decreases and biochemical and pathological liver changes observed at 370 mg/kg bw/d and above. The uncertainty factor of 100 is recommended, on the basis of the fact that the test substance was not genotoxic and did not have teratogenic or reproductive toxicity potential, and that the carcinogenic effect in the liver appeared to be mediated through a non-genotoxic mechanism of tumour promotion.

No additional uncertainty or safety factors were deemed necessary; the database was considered adequate. The apparent increased susceptibility identified in the rat reproductive and developmental toxicity study (decreases in body weights of F_{1b} and F₂ pups and delayed developmental landmarks) occurred at a very high dose, 16 000 ppm (1500 mg/kg bw/d), well above the limit dose at which maternal toxicity was clearly evident.

The U.S. EPA also assigned a reference dose (RfD) of 0.36 mg/kg bw/d non-carcinogenic end points, on the basis of a no observed effect level (NOEL) of 36 mg/kg bw/d (set in the two-year rat study), and using an uncertainty factor of 100.

3.2.2 Carcinogenic endpoints

Chronic toxicity/carcinogenicity, genotoxicity and mechanistic data suggested that the mechanism of liver tumour (hepatocellular carcinomas in rats) induction by kresoxim-methyl was through non-genotoxic “tumour promotion”, mediated by stimulation of prolonged cell proliferation, which appeared to be reversible upon cessation of dosing. However, there was insufficient information to ascertain that the stimulation of liver cell proliferation was a threshold effect. Thus, it was considered most appropriate to utilize the quantitative low dose extrapolation (Q₁^{*}) approach to cancer risk assessment. The Q^{*} value calculated by the U.S. EPA for this purpose was readily available, and was utilized for the cancer risk assessment.

The cancer estimate risk number (Q₁^{*}) assigned by the U.S. EPA for kresoxim-methyl was 2.90×10^{-3} , on the basis of female rat liver tumour development rates from the two-year oncogenicity study (Federal Register, vol. 64, no. 111, June 10, 1999).

3.3 Acute reference dose (ARfD)

In the context of low acute toxicity by the oral, dermal and inhalation routes, and absence of relevant evidence of acute toxicity in the appropriate short-term studies, it is not necessary to propose an acute reference dose (ARfD).

3.4 Toxicology end-point selection for occupational and bystander risk assessment

The end-use product, Sovran[®], is of low acute toxicity by the oral, dermal and inhalation routes. Sovran[®] is a mild eye irritant and is not a dermal irritant or a dermal sensitizer.

In repeated dose toxicology studies, the liver was identified as the target organ. There was no evidence of developmental sensitivity, teratogenicity, reproductive toxicity or neurotoxicity.

Exposure to the mixer, loader or applicator would be short term (i.e., four applications per year) and predominantly via the dermal route. As such, the NOAEL of 1000 mg/kg bw/d, on the basis of absence of toxicity at the highest dose tested from the

21-day dermal toxicology study, was considered most appropriate. A full range of parameters were investigated in this study including clinical signs, body weight gain, hematology, clinical chemistry and macroscopic and microscopic pathology. For non-cancer end points, a margin of exposure of 100 to account for intraspecies and interspecies differences is considered acceptable.

Exposure to the re-entry worker would be of an intermediate term (i.e., several weeks) and predominantly via the dermal route. As such, a study longer than the 21-day dermal toxicology study was considered appropriate, as increased toxicity was observed in female rats and mice following increased duration of exposure. The NOAEL of 146 mg/kg bw/day in the 90-day rat study, on the basis of increases in liver enzymes and liver weights at the next dose, was considered most relevant for the risk assessment for the re-entry workers. For non-cancer end points, a margin of exposure of 100 to account for intraspecies and interspecies differences is considered acceptable.

Kresoxim-methyl caused liver cell carcinomas in rats following long-term feeding. The mechanism of tumour induction was determined to be through tumour promotion by induction of prolonged cell proliferation. Cell proliferation was reversible on cessation of dosing. However, there was insufficient information to ascertain that the stimulation of liver cells proliferation was a threshold effect. Thus, it was considered most appropriate to utilize the quantitative low dose extrapolation (Q_1^*) approach to cancer risk assessment. The Q^* value circulated by the U.S. EPA for this purpose was readily available, and was utilized for the cancer risk assessment.

The Q_1^* assigned by the U.S. EPA for kresoxim-methyl was 2.90×10^{-3} (mg/kg bw/day)⁻¹, on the basis of female rat liver tumour rates from the two-year oncogenicity study (Federal Register, vol. 64, no. 111, June 10, 1999).

3.5 Impact on human health arising from exposure to the active substance or to impurities contained in it

3.5.1 Operator exposure assessment

Sovran[®] is a 50% wettable granular formulation proposed for agricultural use. Applications would be to apple orchards via airblast equipment and would be performed up to four times per year, with a minimum of 10 days between applications. A farmer could typically treat up to 20 ha per day. Sovran would be applied at a typical application rate of 120 g a.i./ha, up to a maximum rate of 225 g a.i./ha.

Dermal Absorption

Potential dermal absorption of kresoxim-methyl was investigated in an in vivo rat study. Male rats were treated dermally with kresoxim-methyl at three dose levels: 0.35 mg/cm², 0.05 mg/cm², and 0.007 mg/cm² for exposure periods of 0.5, 1, 2, 4, 8, and 24 hours. The application site was washed just prior to sacrifice. Two additional groups were exposed for 8 hours and sacrificed after 24 or 72 hours (i.e., 8/24 or 8/72). For these groups, the

application site was washed after 8 hours exposure, and the animals were returned to their cages for an additional 16 or 64 hours before sacrifice. Most of the applied radioactivity was recovered from the skin wash and protective cover.

Due to the number of study limitations (e.g., high percentage of applied dose on protective cover) it was considered appropriate to use a conservative estimate of dermal absorption in the occupational risk assessment. The 8/24 hour high dose absorption value of 24.93 was selected.

Non-Cancer Assessment

Pesticide operator exposure was estimated using the Pesticide Handler Exposure Database (PHED) Version 1.1. The PHED is a compilation of generic mixer, loader and applicator, as well as flagger passive dosimetry data with associated software that facilitates the generation of scenario specific exposure estimates. The PHED estimates meet North American Free Trade Agreement criteria for data quality, specificity, and quantity.

To estimate total dermal and inhalation exposure for airblast application, appropriate subsets of A and B grade data were created from the mixer and loader, as well as the applicator PHED database files. The mixer and loader file was subset for open mixing and dry flowable formulations. The applicator file was subset for application by airblast equipment and tractors or trucks with open cabs. The number of replicates for inhalation and dermal data were acceptable. A best-fit statistical measure was used for the exposure estimates.

The daily exposure estimates are based on the PHED exposure estimates, a dermal absorption of 24.9%, and assuming that farmers treat 20 ha of apples/day at a typical application rate of 120 g a.i./ha. Note that since the NOAEL for mixer/loader/applicator (M/L/A) is based on a dermal toxicity study, exposure estimates are based on total dermal deposition only (inhalation exposure is considered negligible compared to dermal deposition). An average daily exposure (dermal deposition plus inhalation) of 0.034 mg/kg bw/day was derived for mixers, loaders and applicators wearing one layer of clothing plus gloves. The exposure estimates and margins of exposure (MOEs) for farmers mixing, loading and applying Sovran Fungicide are presented in Table 1.

Table 1 Scenario specific exposure estimates for farmers mixing, loading and applying Sovran Fungicide to apples in Canadian orchards

Exposure Scenario	PHED ^a Unit Exp (µg/kg a.i.)	Area Treated (ha/day)	Rate (kg a.i./ha)	Daily Exposure ^b (µg/kg-bw/d ay)	MOE
Mix/Load/Apply Single Layer	1002.81	20	0.12	34.38	29085
Mix/Load/Apply Coveralls	856.86	20	0.12	29.38	34039

^a Since the MOE is based on a dermal toxicity study, the exposure estimates are based on total dermal deposition only (inhalation exposure is considered negligible compared to dermal deposition).

^b Calculated as: (PHED Unit Exposure [µg/kg a.i.] × (Application Rate [kg a.i./ha]) × (area treated [ha])/body weight [70 kg].

The MOEs for farmers mixing/loading/applying Sovran Fungicide to apples in Canadian orchards are acceptable.

Cancer Assessment

A lifetime average daily dose (LADD) was derived for mixer/loader/applicators. The LADD was based on the following assumptions: farmers could mix/load/apply Sovran Fungicide for 4 days/season; and, can treat (on average) 20 ha of apples with airblast equipment per day. The working tenure and lifespan are assumed to be 40 years and 75 years, respectively. The LADD and cancer risk estimates for farmers exposed to Sovran Fungicide as a result of mixing/loading/application to apples are summarized in Table 2.

Table 2 Based on mixer/loader wearing single layer and gloves, and applicator wearing single layer and no gloves

Exposure Scenario	Max No. Days Exposed /Season	Daily Dose ^a (mg/kg-bw/ day)	LADD ^b (mg/kg bw/day)	Q ₁ ^{*c} (mg/kg bw/day)	Cancer Risk
Open Mixing/Loading + Airblast Application of Sovran Fungicide					
M/L/A Single Layer	4	0.00620	3.62×10^{-5}	2.90×10^{-3}	1.05×10^{-7}
M/L/A Coveralls	4	0.00534	3.12×10^{-5}	2.90×10^{-3}	9.05×10^{-8}

^a Based on PHED dermal exposure (adjusted for 24.9% dermal absorption) plus inhalation exposure, and application of 2.4 kg a.i./day (treating 20 ha/day at 0.120 kg a.i./ha). Calculated as: (PHED Unit Exposure [µg/kg a.i.] × (Application Rate [kg a.i./ha]) × (area treated [ha])/body weight [70 kg].

^b Based on an exposure frequency of 4 days/year, a 40 year working tenure, and a life expectancy of 75 years.

^c Q₁^{*} value for kresoxim-methyl based on formation of liver tumours in female rats in a 2-year oncogenicity study.

The cancer risk for farmers applying Sovran Fungicide to apples is acceptable.

3.5.2 Bystanders

For the proposed agricultural use scenario, bystander exposure during and after application was considered minimal compared to mixer/loader/applicator and re-entry worker scenarios and, therefore, not quantified.

3.5.3 Workers

For apples, a range of re-entry activities take place at different stages of cultivation. Thinning, pruning and harvesting were identified as frequent activities which involved high levels of foliar contact. Other re-entry activities such as scouting, weeding and irrigation occur less frequently and involve lower levels of foliar contact.

Exposure estimates for re-entry workers were carried out using dislodgeable foliar residue study conducted on grapes. The dislodgeable foliar residue study was conducted using a kresoxim-methyl formulation on grape vineyards at three locations (Washington, New York and California). Kresoxim-methyl was applied to the grape vineyards using airblast equipment at an application rate of 0.224 kg a.i./ha. Four applications were made, approximately 10 days apart. Sampling occurred immediately before and after each application, and 1, 2, 3, 4, 5, 7, 14, 21, 28 and 35 days after final application. Dislodgeable residues were sampled from grape leaves using a Birkestrand leaf punch sampler. Each sample consisted of 40 leaf punches, and triplicate samples were taken at each time interval, at each site. An untreated control plot was also sampled at each time interval at each site, and an extensive array of field recovery, laboratory recovery, dislodging efficiency, travel recovery and method validation studies were conducted. The analyte was kresoxim-methyl; there was no analysis for transformation products. The application methods, rates and frequency were relevant to the use pattern proposed for Canada. The PMRA considers that the grape dislodgeable foliar residue (DFR) study is an acceptable surrogate for an apple DFR study. Although foliage type is different (grape leaves are smooth whereas apple leaves are waxy), the application methods are similar. However, in the grape DFR study, kresoxim-methyl was applied at a rate of 224 g a.i./ha, while the typical application rate of kresoxim methyl to apples is 120 g a.i./ha. Therefore, to compensate for the differences in application rate, the grape DFR values were multiplied by a correction factor of 120/224 (i.e., the ratio of their application rates).

Dissipation patterns showed pseudo-first-order kinetics with r^2 values greater than 0.9 at all three sites. On the basis of the relative dissipation rates and environmental conditions, the Washington data set was considered most relevant to Canadian conditions, and was used to estimate potential dermal deposition for a worker re-entering a grape vineyard at various post-application time intervals.

Non-Cancer Assessment

Daily exposure estimates for re-entry workers were calculated using the following assumptions: a transfer coefficient (TC) of 3000 cm²/h for high foliar contact activities including thinning, pruning, and harvesting, a dermal absorption of 24.93%, 8 hours of

exposure per day, and a 70 kg body weight. The re-entry exposure estimate and MOE for workers re-entering apple orchards treated with Sovran Fungicide are 0.023 mg/kg bw/day and 6500, respectively. The MOEs for agricultural workers re-entering treated fields are acceptable.

Cancer Assessment

The lifetime average daily dose (LADD) was derived for agricultural workers based on the following inputs: re-entry on 120 days per season, re-entry interval of 48 hours, dermal absorption value of 24.93%, 8 h exposure/day, 70 kg body weight, 40 year working tenure and a 75 year lifespan. Dislodgeable foliar residue data were coupled with a transfer coefficient of 3000 cm²/hour for high foliar contact activities (112 days) and a transfer coefficient of 1000 cm²/hour for low foliar contact activities which could occur during the re-entry intervals (8 days). The resulting LADD for re-entry workers of 1.38×10^{-3} was coupled with the Q1* value of 0.0029 mg/kg bw/day to yield a risk level of 4.0×10^{-6} . This risk level, based on establishment of a 48 hour re-entry interval, is considered acceptable as some of the inputs to the exposure assessment (e.g., exposure frequency, dermal absorption value) are considered conservative.

4.0 Residues (see also residue summary tables in Appendix II)

Metabolism studies submitted demonstrated the fate and disposition of kresoxim-methyl in apples, grapes, wheat, ruminants and rats. The unchanged parent compound accounted for significant residues in wheat, grapes and Mutsu variety apples, but not in Macintosh variety apples. Also, differences in the quantitative nature of residues was demonstrated by comparison of the two apple metabolism studies. The ROC in apples should be defined as parent, 490M1, 490M2 (free and glucose conjugated) and 490M9 (free and glucose conjugated).

The qualitative and quantitative nature of the residue is understood following oral dosing in rats and goats. Kresoxim-methyl was extensively metabolized in both species, with little or no tissue bioaccumulation. On the basis of the animal metabolism studies, the residue of concern was defined as parent compound, 490M1, 490M2 and 490M9. Since the major rat and goat metabolic profiles were similar, a swine metabolism study is not required.

Method 350/3-US, an HPLC method with UV detection (270 nm), was used to determine residues of kresoxim-methyl and predominant metabolites (490M1, 490M2 and 490M9) in plants. This method converts glucosides of 490M2 and 490M9 into their free metabolites and hydrolyses the parent compound to 490M1. Metabolite 490M1 is measured as kresoxim-methyl equivalents. The LOQ for each analyte of the ROC was established at 0.05 ppm, for a total of 0.15 ppm in apples and grapes and their processed commodities, and in pecans. This method was found to give good recoveries and the standard deviations measured with respect to recoveries following spiking at 0.05–5 ppm were indicative of the method having good to satisfactory repeatability. Interlaboratory

validations in apples, grapes and pecans supported the reliability and reproducibility of method 353/3-US.

Methods 354/1-US and 354/2 both use HPLC with UV detection (270 nm) to determine the predominant metabolites (490M1, 490M2 and 490M9) in ruminant tissues. Method 354/1 is considered to be valid by the PMRA for the determination of 490M2 and 490M9 in milk. The LOQ for each analyte was established at 0.002 ppm, for a total of 0.004 ppm in whole milk. This method was found to give good recoveries and is recommended as the enforcement method for analyses of milk from animals treated orally with kresoxim-methyl.

Method 354/2, used for the determination of the predominant metabolites in ruminant tissues, is considered to be valid for analyses in muscle and liver, but not for analyses in kidney and fat. Recoveries were not always within guideline requirements in kidney and fat samples, especially at the stated LOQ of 0.01 ppm for each analyte (total of 0.03 ppm in kidney and fat; total of 0.02 ppm in muscle and liver). The goat metabolism studies indicated that the highest proportions of 490M1, 490M2 and 490M9 were found in muscle, kidney and milk, respectively. Since method 354/2 showed contradictory results for analyses of 490M2 in kidney, as well as for all three analytes in fat, it was concluded that analyses for the most predominant metabolites in these tissues by method 354/2 are not reliable. On the basis of acceptable analyses of the most predominant metabolites in muscle and liver, but unacceptable analyses of the most predominant metabolites in fat and kidney, method 354/2 is not recommended as an enforcement method for meat and meat by-products.

Kresoxim-methyl and its metabolites, 490M1, 490M2 and 490M9, were subjected through the U.S. Food and Drug Administration multiresidue protocols (MRM). Since the MRM analyses for the ROC in both plant and animal commodities, it may be considered as a singular enforcement method compared with separate enforcement methods for plant (350/3-US) and animal (354/1-US and 354/2) matrices. In contrast to method 350/3-US, the MRM analyses for the parent compound and 490M1 separately. Although methods 350/3-US and 354/1-US were also accepted as enforcement methods, method 354/2 was not accepted by the PMRA. Therefore, the submitted MRM is the only method currently accepted by the PMRA for enforcement analyses in meat and meat by-products.

Submitted freezer stability studies in plant matrices indicated that ROC were stable for at least 30 months when stored at less than -5°C in apples and grapes, 12 months when stored at less than -10°C in grapes, apples and apple process fractions, and 6 months when stored at less than -5°C in pecan. The analytical methods (351/2 and 350/3-US = 350/3 ~ D9611) used in the storage stability studies were adequately sensitive and reproducible, with no evidence of interfering residues or background.

Submitted freezer stability studies in ruminant matrices indicated that ROC were stable at -20°C for at least 12 months when stored in whole milk and 6 months when stored in

beef tissues. At 13 months, unacceptable stability was observed for 490M1 in liver (67% relative recovery) and for 490M9 in kidney (67% relative recovery).

The results from supervised field trials (method 350/3-US) in representative Canadian zones indicated that the maximum residues in apples, collected 30 days following the last application of Sovran[®] DF (50% kresoxim-methyl) and treated at 0.88 kg a.i./ha/season (0.98× good agricultural practices [GAP]), were all less than 0.5 ppm. Two residue decline studies followed the same procedures as the supervised field trials, except that samples were collected at 10-, 20-, 30-, 40- and 60-day PHIs. No residues were measured at PHIs of 40 days or more. Considering these results, the proposed maximum residue level (MRL) for the ROC in apples is 0.5 ppm, with a minimum PHI of 30 days.

In a processing study, apples treated with BAS 490 02F (50% kresoxim-methyl) at 1×, 3× or 5× GAP, with a PHI of 30 days, were processed into apple juice and wet apple pomace. A comparison of the residues in the raw agricultural commodity (RAC) with those in each processed fraction resulted in concentration factors of 0.2× for apple juice and 2.6× for apple pomace at a 30-day PHI. The PMRA will recommend an MRL of 0.15 ppm (LOQ) in apple juice to ensure that residues in apple juice do not pose an unacceptable dietary risk. As apple pomace is a livestock feed, an evaluation of the transfer of kresoxim-methyl residues to livestock tissue and milk was assessed in the livestock feeding study.

In the livestock feeding study, dairy cows were fed kresoxim-methyl in a feed premix at levels of 6, 18 and 60 ppm. Results obtained from the analysis of treated animal tissues indicated that 490M1 reached levels of 0.034 ppm in the kidney, and 490M2 and 490M9 did not exceed the LOQ in any tissues when animals were fed a diet containing 6 ppm kresoxim-methyl for 28 days. The anticipated maximum dietary burden in dairy cows resulting from the feeding of wet apple pomace treated with kresoxim-methyl at GAP was not expected to exceed 0.24 ppm. On the basis of the anticipated dietary burden (25× lower than the lowest feeding level of 6 ppm), it is not expected that ROC for kresoxim-methyl will exceed the LOQ for the analytical method for milk, meat and meat by-products. Therefore, the expected residues resulting from the feeding of treated apples to ruminants will be covered by the following MRLs:

milk:	0.004 ppm
meat and meat by-products:	0.03 ppm

These proposed MRLs differ from U.S. tolerances because no tolerance has been proposed for milk in the United States and because definitions for the ROC in food of animal origin differ between the United States and Canada.

Supervised residue trials on pears, grapes and pecans and a processing study in grapes were also submitted.

The results from the supervised crop field trials study in grapes (method 350/3) conducted in the United States showed that the maximum residues in grapes, collected 14 days following the last application of BAS 490 02F (50% kresoxim-methyl) and treated at 0.896 kg a.i./ha/season equivalent to U.S. GAP, were less than 0.793 ppm (highest average field trial [HAFT] = 0.793 ppm). Maximum residues at a 30-day PHI were less than 0.732 ppm. Consequently, an MRL of 1.0 ppm should be established to cover ROC for kresoxim-methyl in or on grapes imported to Canada.

In a processed food and feed study, BAS 490 02F (50% kresoxim-methyl) was applied to grapes at 2.69 kg a.i./ha with PHIs of 1 and 14 days. Data were reported for the 14-day PHI only. The grape samples were processed into grape juice and raisins. A comparison of the residues in the RAC with those in each processed fraction resulted in a concentration factor of 0.1–0.8 for grape juice and 1.5–1.6 for raisins at a 14-day PHI. The maximum combined residues of kresoxim-methyl and its metabolites expected in raisins, on the basis of the HAFT of 0.793 ppm reported from the residue trial study and the maximum concentration factor (1.6×), is 1.27 ppm. The residues expected in grape juice should be covered under the proposed MRL for the RAC (1.0 ppm). An MRL of 1.5 ppm should be established to cover potential residues in or on raisins imported to Canada.

The results from the supervised crop field trials study in pecans (method D9611A ~ method 350/3) conducted in the United States showed that the maximum residues in pecans, collected 44 or 45 days following the last application of BAS 490 02F (50% kresoxim-methyl) and treated at 1.97 kg a.i./ha per season, were all less than 0.15 ppm (LOQ = 0.15 ppm). One residue decline study was also submitted as a portion of these crop residue trials. Sampling occurred at 35-, 55- and 65-day PHIs. Residues were less than 0.15 ppm at all PHIs. Consequently, an MRL of 0.15 ppm should be established to cover ROC for kresoxim-methyl in or on pecans imported to Canada.

The dietary cancer risk from kresoxim-methyl was calculated on the basis of the Q_1^* linear model approach. On the basis of a Q_1^* of 0.0029 mg/kg bw/d, the risk estimate from all dietary sources was estimated to be 2.4×10^{-7} to 1.3×10^{-6} , which was considered to be below the level of concern for lifetime cancer risk, because of the following conservative assumptions used in arriving at this risk estimate: (a) 100% of the crops (apples, grapes and pecans) consumed by Canadians from any source (imported or domestically produced) are treated at the maximum label rate; (b) in addition, there was a maximum transfer of residues into meat and milk; and (c) no allowance has been made for customary culinary practices such as washing, peeling and cooking of either the fruit or the meat and milk. Consequently, the proposed domestic use of kresoxim-methyl on apples and proposed imports of grapes, raisins and pecans treated with kresoxim-methyl does not pose an unacceptable dietary risk to any segment of the population including adults, infants and children, because of the conservative estimate outlined above.

5.0 Fate and behaviour in the environment (see also environmental assessment summary tables in Appendix III)

5.1 Physical and chemical properties relevant to the environment

Kresoxim-methyl has low solubility in water (2.0 mg/L) and will be non-volatile under field conditions (vapour pressure, 2.3×10^{-6} Pa), and from water and moist soil surfaces (Henry's Law Constant, 3.6×10^{-4} Pa·m³/mol). The octanol-water partitioning coefficient ($\log K_{ow} = 3.40$) indicates that there is a potential for bioaccumulation from aquatic systems.

5.2 Abiotic transformation

The rate of hydrolysis of kresoxim-methyl is highly dependent on the ambient pH. Under basic conditions (pH 9), hydrolysis is rapid (half life of 9 h), resulting in the formation of BF 490-1. Under neutral (pH 7) and acidic conditions (pH 5), kresoxim-methyl is more stable and hydrolysis is slower with first-order half-lives of 32 d and 874 d, respectively. Phototransformation is not a major route of transformation on soil or in water with first order half-lives of 70.4 d on soil and 14.8 d to 59.6 d in water (depending on the position of the radiolabel) under conditions of 12 h light:12 h dark.

5.3 Biotransformation

The primary route of transformation of kresoxim-methyl is biotransformation by aerobic micro-organisms in both soil and aquatic systems. The major transformation product produced in soil and water is the acid form, BF 490-1.

In aerobic soil, kresoxim-methyl is transformed rapidly with DT₅₀s ranging from <1 to 4.7 d. In aerobic and anaerobic water/sediment systems, kresoxim-methyl transformed rapidly with DT₅₀s of 1.5 d and 0.9–1.3 d, respectively. Therefore, kresoxim-methyl is non-persistent in soil, water or sediment under either aerobic or anaerobic conditions. Data regarding transformation processes of kresoxim-methyl are summarized in Table 1 of Appendix III.

The major transformation product, BF 490-1, was produced in aerobic soils and has a DT₅₀ ranging from 58.8 to 131 d. Thus, BF 490-1 is classified as moderately persistent in aerobic soils. In aerobic aquatic (water-sediment) systems, BF 490-1 was the major transformation product produced with a DT₅₀ of $\gg 93$ d to 462 d. Therefore, BF 490-1 is considered to be moderately persistent to persistent in aerobic water-sediment systems. As for aerobic water/sediment conditions, BF 490-1 was the major transformation product produced under anaerobic water-sediment conditions with a DT₅₀ ranging from 98–130 d. Therefore, BF 490-1 is classified as moderately persistent in anaerobic water/sediment systems. BF 490-1 is primarily associated with the aqueous phase, therefore, it may be bioavailable to aquatic organisms.

Data on the transformation processes of the major transformation product BF 490-1 are summarized in Table 2 of Appendix III.

5.4 Mobility

Laboratory studies of mobility indicated that kresoxim-methyl exhibits low to moderate mobility in most types of soil (K_{oc} values ranged from 249–567 mL/g OC), but is moderately to highly mobile in sandy soils. Thus, non-bound fractions of kresoxim-methyl may move through soils to the water table to enter the groundwater, especially in sandy soils. Kresoxim-methyl may also run off the soil surface and enter surface waters. Field studies indicated that the residues of kresoxim-methyl were primarily detected in the top 0–15 cm layer of soil; therefore, kresoxim-methyl has a low potential to leach under field conditions.

The potential for kresoxim-methyl to contaminate groundwater was evaluated using the Groundwater Ubiquity Score (GUS) assessment method of Gustafson (1989) and the Expert System for Pesticide Regulatory Evaluation and Simulation (EXPRES) model. The calculated GUS of 1.7 falls into the range for non-leachers (<1.8). This indicates that kresoxim-methyl is unlikely to leach to groundwater. For EXPRES, two indices are calculated, leaching potential (LP; a relative measure of the potential of the pesticide to leach to the water table) and leaching index (LI; a relative measure of the potential migration distance of the pesticide), and compared with four pesticides known from field measurements to have leached to groundwater. Kresoxim-methyl was ranked 50th on the LP scale and 59th on the LI scale of the 130 pesticides in the EXPRES database (Table 5.1). Based on the results of the EXPRES and GUS models and terrestrial field dissipation studies, there is a relatively low probability that kresoxim-methyl will cause groundwater contamination when applied in the field. Data regarding the mobility of kresoxim-methyl are summarized in Table 1 of Appendix III.

Table 5.1 Leaching potential of kresoxim-methyl compared with four pesticides known to leach to groundwater

Pesticide	LP rank	LI rank
Picloram	24	18
Atrazine	45	42
Dicamba	47	46
Kresoxim-methyl	50	59
Dinoseb	70	71

Laboratory studies indicate that BF 490-1 is highly mobile and leached through soils. Based on these laboratory characteristics of persistence and leaching, BF 490-1 would be expected to leach to groundwater. However, field trials indicated that BF 490-1 has a low

potential to leach under field conditions. Not enough information was available to determine the leaching potential of BF 490-1 using the EXPRES model. In aquatic systems, BF 490-1 is found primarily in the water column, where it may be bioavailable to aquatic organisms. Data regarding the transformation processes and mobility of BF 490-1 are summarized in Table 2 of Appendix III.

5.5 Dissipation and accumulation under field conditions

Studies under Canadian and northern U.S. field conditions were conducted at six sites (in Nova Scotia, Ontario, British Columbia, New York, Oregon and California). In the field studies, four applications of BAS 490 F were made at 7-day intervals to bare soil at a rate of approximately 0.26 kg a.i./ha. The DT₅₀ for BAS 490 F ranged from <1 to 11 days in Canada and <1 h to 2.9 days in the northern U.S. The DT₉₀ ranged from 8 to 46 days in Canada and <1 h to 10.9 days in the northern U.S. For all sites, at the end of the 360-d study period, there was no carryover of residue of BAS 490 F. Given the range of DT₅₀s, according to Goring et al. (1975), BAS 490 F is non-persistent in soil under field conditions.

Two transformation products were quantified in the field studies: BF 490-1 (acid) and BF 490-5 (di-acid). These products formed quickly after the last application, increased to a maximum concentration, and dissipated relatively quickly. The DT₅₀s for BF 490-1 (+BAS 490 F) ranged from 12 to 52 days in Canada and 4.9 to 6.8 days in northern U.S. The DT₅₀s for BF 490-5 ranged from <2 to 13 days in Canada and 4 to 18 days in northern U.S. BF 490-1 is classified as slightly to moderately persistent in Canadian soils under field conditions, but is non-persistent in northern U.S. soils. BF 490-5 is non- to slightly persistent under field conditions. BF 490-1 and BF 490-5 amounted to a maximum of 76–97% and 3.6–12.9% of the applied amount, respectively, within a few days of the final application, but were not quantifiable at the end of the 360-d study period. The field studies demonstrated that kresoxim-methyl has a low potential to leach through soils. Furthermore, the field studies indicate that kresoxim-methyl and its transformation products will dissipate in the environment without deep leaching.

Figures correlating the relative decline of the parent compound with the rise and decline of the transformation products were typical of microbial degradation with kresoxim-methyl (BAS 490 F) being transformed to BF 490-1 then BF 490-5 (see Figure 1 in Appendix III). The longer DT₅₀ and DT₉₀ observed for the British Columbia site may result from the generally drier conditions that were present, even with supplemental irrigation. Dry conditions are not conducive to enhancing microbial degradation. In all soils studied, residues of kresoxim-methyl and its transformation products were primarily detected in the top 0–15 cm soil layer, therefore, these compounds have a low potential to leach under field conditions.

5.6 Bioaccumulation

For the parent compound, kresoxim-methyl, bioconcentration factors (BCF) of 220, 430, and 52, were determined for whole fish, viscera, and fillets, respectively. Kresoxim-methyl was rapidly eliminated when fish were moved to clean water, thus, kresoxim-methyl is considered not to accumulate in aquatic organisms.

For the major transformation product BF 490-1, the octanol-water partition coefficient is highly variable and dependent upon environmental conditions (pH). The log K_{ow} of BF 490-1 is 2.95, thus, the log K_{ow} does not meet the Toxic Substances Management Policy (TSMP) criterion of log $K_{ow} \geq 5$. Although the log value of 2.95 (ranging from 2.85 to 3.03) meets the trigger for requiring bioaccumulation studies with fish, at an environmentally-relevant pH (buffered solution at pH 7), the log K_{ow} of BF 490-1 is 0.15, thus, bioaccumulation is unlikely to occur. Therefore, studies on the bioaccumulation of BF 490-1 in fish are not required.

5.7 Summary of fate and behaviour in the terrestrial environment

The fate and behaviour of kresoxim-methyl in soil was assessed by studying the photolysis, biotransformation, and mobility under laboratory conditions. In addition, the dissipation of kresoxim-methyl in the field was assessed. The primary route of transformation of kresoxim-methyl is biotransformation by aerobic microorganisms in both soil and aquatic systems. The major transformation product in soil is an acid, BF 490-1. The DT_{50} of kresoxim-methyl in field studies ranged from <1 to 11 days in Canada and <1 h to 2.9 days in the northern U.S. Therefore, kresoxim-methyl is non-persistent in soil. At all field study sites, no carryover of kresoxim-methyl residues was observed.

Two transformation products, BF 490-1 (acid) and BF 490-5 (di-acid), were quantified in the field studies. The DT_{50} s for BF 490-1 (+BAS 490 F) ranged from 12 to 52 days in Canada and 4.9 to 6.8 days in northern U.S. The DT_{50} s for BF 490-5 ranged from <2 to 13 days in Canada and 4 to 18 days in northern U.S. BF 490-1 is classified as slightly to moderately persistent in Canadian soils under field conditions, but is non-persistent in northern U.S. soils. BF 490-5 is non- to slightly persistent under field conditions. In the field studies, residues of kresoxim-methyl and its transformation products were primarily detected in the top 0–15 cm soil layer, therefore, these compounds have a low potential to leach under field condition.

5.8 Summary of fate and behaviour in the aquatic environment

Laboratory studies examined the abiotic and biotic transformation pathways for kresoxim-methyl in water including hydrolysis, phototransformation and biotransformation under aerobic and anaerobic conditions. The rate of hydrolysis of kresoxim-methyl is highly dependent on the ambient pH with rapid hydrolysis occurring under basic conditions (pH 9). Under neutral (pH 7) and acidic conditions (pH 5),

kresoxim-methyl is more stable and hydrolysis is slower. As in soil, the principal route of transformation of kresoxim-methyl in water is biotransformation by aerobic microorganisms. As in soils, the major transformation product in water is BF 490-1.

Kresoxim-methyl is non-persistent in water and sediment under either aerobic or anaerobic conditions. The major transformation product BF 490-1 is moderately persistent to persistent in aerobic water-sediment systems and persistent in anaerobic water-sediment systems.

5.9 Expected environmental concentrations

5.9.1 Soil

Kresoxim-methyl is proposed for use in Canada on apples at a rate of 0.225 kg a.i./ha, with applications a minimum of 10 days apart, and no more than four applications. The maximum cumulative application rate on soil, taking into account a soil dissipation time 50% (DT_{50}) of 11 days for kresoxim-methyl, is 0.443 kg a.i./ha. Assuming a soil bulk density of 1.5 g/cm³, application at the maximum cumulative rate (443 g a.i./ha) to bare soil with no interception by foliage and uniform mixing in soil over a depth of 15 cm, the expected environmental concentration (EEC) of kresoxim-methyl in soil is 0.20 mg a.i./kg soil dry weight.

5.9.2 Aquatic systems

Expected environmental concentrations in water were calculated by assuming a scenario in which the Canadian label rate (225 g a.i./ha) was applied the maximum recommended number of times (four) at the shortest interval allowed between sprays (10 days). To calculate a maximum cumulative application rate, transformation of the parent compound in soil (runoff) and water (direct overspray) was taken into consideration.

Expected environmental concentration in water from direct overspray:

Using the DT_{50} of 1.6 days in water from the biotransformation in the aerobic water and sediment study, the EEC of kresoxim-methyl in water immediately following the fourth application at 0.225 kg a.i./ha is the equivalent of a cumulative application of 0.228 kg a.i./ha. Assuming a scenario in which a body of water 30 cm deep is oversprayed with the equivalent of a cumulative application of 0.228 kg a.i./ha, the EEC in water is 0.08 mg a.i./L water (see Table 7 of Appendix III).

Concentration in drinking water (runoff):

Expected environmental concentrations of kresoxim-methyl in drinking water were estimated from a U.S. EPA Tier I screening model named GENEEC assuming four applications at 225 g a.i./ha and a minimum 10-day interval between applications. Various parameters from laboratory studies were used in the model. Because the model, GENEEC, does not have an air-blast scenario (by which kresoxim-methyl will likely be applied), a ground application scenario with 1% spray drift and an aerial application

scenario with 5% spray drift were tested. Four generic EECs were generated. The results are presented in Table 5.2. The peak concentration in drinking water is expected to be approximately 13 µg a.i./L.

Table 5.2 Generic expected environmental concentrations calculated by GENEEC model for deep water bodies (2 m)

Application method	Generic EEC (µg a.i./L)			
	Peak	Averaged over 4 days	Averaged over 21 days	Averaged over 56 days
Ground	13.05	7.62	1.75	0.66
Aerial	12.83	7.54	1.74	0.65

Expected environmental concentration in pond water (shallow water) from runoff:
The peak values generated by the GENEEC model were converted to a 30 cm deep pond to estimate the concentration resulting from a runoff episode. The EEC in shallow pond water resulting from runoff is 0.087 mg a.i./L (87 ppb).

5.9.3 Vegetation and other food sources

No data were provided on concentrations of kresoxim-methyl on foliar crops immediately after application. Thus, in the absence of these data, concentrations of kresoxim-methyl on vegetation were estimated using a nomogram developed by the U.S. EPA from the data of Hoerger and Kenaga (1972) and Kenaga (1973) for use in ecological risk assessment (Urban and Cook, 1986) (see Table 3a of Appendix III). A fresh to dry weight conversion was also calculated. The maximum application rate of 900 g a.i./ha was used, which assumes no transformation occurred, as the half-life ($t_{1/2}$) of kresoxim-methyl on vegetation is unknown. The EECs were used to estimate the highest concentration of kresoxim-methyl that may be present in a typical diet of wild birds and some common mammals when exposed to maximum application rates and frequencies (see Table 3b of Appendix III). These concentrations were used to determine the risk to wild birds and mammals.

6.0 Effects on non-target species

6.1 Effects on terrestrial organisms

Kresoxim-methyl and the formulated end product are non-toxic to earthworms, honeybees and wild mammals. The major transformation product, BF 490-1, was non-toxic to earthworms. Exposure to the formulated end product resulted in toxic effects in beneficial predatory mites and ladybird beetles after a single application below the maximum proposed application rate for Canada. Effects were also observed with a terrestrial vascular plant (lettuce). Kresoxim-methyl is practically non-toxic to wild birds;

on a short-term dietary basis, however, dose-related reproductive effects, including damaged (cracked) eggs, infertile eggs, mortality of developing eggs and mortality at hatch (chicks dead in shell), were observed with the bobwhite quail. The NOEL and lowest observed effect level (LOEL) from a reproductive study with the mallard duck were 100 and 500 mg a.i./kg diet, respectively. Data are summarized in Table 4 of Appendix III.

6.2 Effects on aquatic organisms

Kresoxim-methyl was highly toxic to *Daphnia magna*, rainbow trout and bluegill sunfish on an acute basis and affected the survival of juvenile fathead minnows at 160 µg a.i./L. Although toxic to fish, kresoxim-methyl is not expected to bioconcentrate in fish tissues. Effects were observed in the freshwater algal species exposed to kresoxim-methyl. The most sensitive algal end point was a no observed effect concentration (NOEC) of 12.0 µg a.i./L in *Navicula pelliculosa*, a freshwater diatom. No effects were observed in an aquatic vascular plant. In contrast to the active ingredient, the major transformation product, BF 490-1, was practically non-toxic to *Daphnia magna* and rainbow trout on an acute basis. The toxicity of kresoxim-methyl to marine biota was not reviewed because, under the given pattern of orchard use, the risk of kresoxim-methyl entering the marine environment in Canada is minimal. Data are summarized in Table 5 of Appendix III.

6.3 Effects on biological methods of sewage treatment

Data on biological methods of sewage treatment were not required.

6.4 Risk characterization

Separate risk assessments were carried out to examine the environmental fate of kresoxim-methyl and the toxicity to terrestrial and aquatic organisms. The risks to non-target marine invertebrates and fish were not assessed due to the limited risk of kresoxim-methyl entering the marine environment based on the proposed orchard use patterns in Canada. From the review of environmental toxicology, exposure to the major transformation product, BF 490-1, in terrestrial and aquatic systems, did not result in toxicity in the organisms tested (earthworms, *Daphnia magna*, and rainbow trout). Therefore, although the transformation product is moderately persistent in the environment, exposure to it is not expected to result in acute toxicological effects in terrestrial or aquatic organisms.

6.4.1 Environmental behaviour

Leaching Potential

The potential for kresoxim-methyl to contaminate groundwater was evaluated using the Groundwater Ubiquity Score (GUS) assessment method of Gustafson (1989). Using the maximum value of DT₅₀ from the field studies (11 days) and the minimum K_{oc} value from the adsorption/desorption studies (K_{oc} = 249 mL/g OC), the GUS score for kresoxim-

methyl is calculated as 1.7. This indicates that kresoxim-methyl is a non-leacher and is therefore unlikely to present a risk to groundwater.

The potential for kresoxim-methyl to contaminate groundwater was also evaluated using the Expert System for Pesticide Regulatory Evaluations and Simulations (EXPRES) model developed by Environment Canada for use by the Environmental Assessment Division of the PMRA. Using a conservative data set for solubility (2.0 mg/L), vapour pressure (2.3×10^{-3} mPa), biotransformation in soil ($DT_{50} = 4.7$ days, maximum from aerobic soil laboratory studies), and K_{oc} (249 mL/g OC, minimum value), kresoxim-methyl ranked 50th out of 130 pesticides in the EXPRES database on the scale indicating the relative potential for the pesticide to leach to the water table. Similarly, kresoxim-methyl ranked 59th on a scale measuring the relative potential migration distance of the pesticide prior to its degradation in the subsurface. Compared to four pesticides known to leach (atrazine, dinoseb, dicamba, and picloram), there is a relatively low probability that kresoxim-methyl will cause groundwater contamination when applied in the field (Table 1). Factors such as area and quantity of application must also be taken into account when comparing the relative leaching potential of kresoxim-methyl with other pesticides in the database.

Table 1 **Ranked leaching potential of kresoxim-methyl compared to four pesticides known to leach to groundwater**

Pesticide	Rank	
	Potential for leaching	Potential migration distance
Picloram	24	18
Atrazine	45	42
Dicamba	47	46
Kresoxim-methyl	50	59
Dinoseb	70	71

6.4.2 Terrestrial organisms

Wild birds: Wild birds, such as bobwhite quail and mallard duck, could be exposed to residues of kresoxim-methyl as a result of the consumption of sprayed vegetation and (or) contaminated prey. Based on the diet of the bobwhite quail, the EEC of kresoxim-methyl in the diet is 108 mg a.i./kg dry weight. Based on the diet of the mallard duck, the EEC of kresoxim-methyl in the diet is 30.4 mg a.i./kg dry weight.

Based on an acute oral toxicity study with bobwhite quail, it would take 204 d for a wild bobwhite quail to ingest a dose administered by gavage equivalent to that which killed 50% of the individuals in a laboratory population. Similarly, the maximum number of days of intake equivalent to the dose that had no observable effect on the laboratory

population is 204 d. These values indicate that the application of kresoxim-methyl at the maximum proposed label rate will not pose any appreciable risk to wild bird populations that are acutely exposed.

In the acute dietary toxicity studies with the bobwhite quail and mallard duck, the NOECs were each 5000 mg a.i./kg dry weight of diet. As the EECs in the diet of the bobwhite quail and the mallard duck are expected to be 108 mg a.i./kg dry weight and 30.4 mg a.i./kg dry weight, respectively, kresoxim-methyl will not pose any appreciable dietary risk to bobwhite quail or mallard ducks at the proposed maximum application rate. The margins of safety for bobwhite quail and mallard duck are 46.3 and 164, respectively.

From the chronic reproduction tests with bobwhite quail, the lowest observed adverse effect level (LOAEL) was 50 mg a.i./kg diet. A NOEL was not determined in this study. Although the EEC of 108 mg a.i./kg diet is greater than the LOAEL, this would suggest that kresoxim-methyl may result in reproductive effects in bobwhite quail at the proposed maximum application rate. However, based on the proposed use and exposure patterns and the rate of transformation of kresoxim-methyl, it is highly unlikely that wild birds would be exposed to kresoxim-methyl for 26 weeks continuously.

The NOEL from reproduction tests with the mallard duck was 100 mg a.i./kg diet. The NOEL is greater than the EEC of 30.4 mg a.i./kg diet, therefore, kresoxim-methyl will not pose any appreciable reproductive risk to mallard duck at the proposed maximum application rate. The margin of safety is 3.29.

Wild mammals: Wild mammals such as rats and mice could be exposed to kresoxim-methyl residues as a result of the consumption of sprayed vegetation and (or) contaminated prey. Based on the diet of the rat, the EEC in the diet is 454 mg a.i./kg dry weight. For the mouse, the corresponding EEC in the diet is 451 mg a.i./kg dry weight.

From an acute oral toxicity study with the rat, it would take more than 220 d of continuous feeding for a wild rat to attain a dose equivalent to that administered by gavage equivalent which killed 50% of the laboratory population. Similarly, the maximum number of days of intake equivalent to the dose that had no observable effect on the laboratory population is 220 d. These values indicate that the application of kresoxim-methyl at the maximum proposed label rate will not pose any appreciable risk to wild populations of rats that are acutely exposed.

In 90-d dietary toxicity studies with male and female rats, the NOECs were 2000 and 16 000 mg a.i./kg dry weight, respectively. These values are greater than the EEC of kresoxim-methyl in the diet of the rat (454 mg a.i./kg dry weight), thus kresoxim-methyl will not pose any appreciable short-term risk to rats at the proposed maximum application rate. The margins of safety for male and female rats are 4.4 and 35.2, respectively.

The EEC of 454 mg a.i./kg dry weight in the diet of the rat is below the most sensitive NOEC from the reproductive studies with the rat (1000 mg a.i./kg dry weight), therefore, kresoxim-methyl will not pose any appreciable chronic risk to rats at the proposed application rate. The margin of safety is 2.2.

In 90-d dietary toxicity studies with male and female mice, the NOECs were 1000 and 8000 mg a.i./kg dry weight, respectively. These values are greater than the EEC of 451 mg a.i./kg dry weight in the diet of mice, thus kresoxim-methyl will not pose any appreciable short-term risk to mice at the proposed maximum application rate. The margins of safety for male and female mice are 2.2 and 17.7, respectively.

Based on the studies with rats and mice, kresoxim-methyl will not pose any appreciable short-term risk to wild mammals.

Bees: In order to be protective of bees, the lowest LD₅₀ for the active ingredient was selected for the risk assessment. The most sensitive LD₅₀ for honeybees was >25 µg a.i./bee (acute contact), which is equivalent to 28 kg a.i./ha. The maximum seasonal application rate of 0.9 kg a.i./ha is lower than the most sensitive LD₅₀, therefore, kresoxim-methyl will not pose any appreciable risk to bees at the proposed application rate. The margin of safety is 31.1.

Earthworms: The toxicity of the formulants in the end-use product are unknown, therefore, the NOEC for the active ingredient was selected for the risk assessment. The NOEC was 937 mg a.i./kg soil. The EEC of kresoxim-methyl in soil (0.20 mg a.i./kg soil) is below the NOEC, therefore, kresoxim-methyl will not pose any appreciable risk to earthworms at the proposed application rate. The margin of safety is 4690.

Parasites and Predators: For predaceous mites, based on two studies on adult mortality, an LC₅₀ of 1071 g EP/ha was calculated by linear regression ($R^2 = 0.72$). A NOEC was not provided, therefore, one-tenth of the LC₅₀ is used in lieu of an actual value. The NOEC is, therefore, 107.1 g EP/ha. The lifespan of adult predaceous mites is approximately 20 days (Kain and Nyrop 1995, Weeden et al. 1998) therefore, an individual mite could potentially be exposed to three applications of kresoxim-methyl using a minimum 10-d interval between applications during the protonymph and adult life stages. Based on the limited information available on the transformation of kresoxim-methyl on leaf surfaces, the EEC is 1350 g EP/ha (three applications × 450 g EP/ha; assuming that no transformation occurs on leaf surfaces). The EEC is greater than the calculated NOEC ($0.10 \times LC_{50}$), therefore kresoxim-methyl is expected to pose a potential risk to beneficial predaceous mites. The margin of safety is 0.079. Application of kresoxim-methyl to trees containing predaceous mites may have a negative impact on their abundance. It may take up to three years to establish a population of such beneficial predators that is high enough to control pest mites (Weeden et al. 1998).

The most sensitive end point for ladybird beetles was a 60% decline in fertility at an application rate of 300 g EP/ha. According to Hoffman and Frodsham (1993), adult

ladybird beetles may live for weeks or months depending on location, availability of prey, and time of year. An adult living for two months (60 d), could potentially be exposed to four applications of kresoxim-methyl. Therefore, the EEC is 1800 g EP/ha (four applications \times 450 g EP/ha). As for predaceous mites, no transformation of the active ingredient, kresoxim-methyl, was assumed to occur on leaf surfaces due to the limited information available. The EEC used for the risk assessment was, therefore, 1800 g EP/ha. If a single application at 300 g EP/ha results in 60% decline in fertility, then a 50% decline in fertility (EC_{50}) would occur at 250 g EP/ha, assuming the dose-response is linear. A NOEC was not provided, therefore, one-tenth of the LC_{50} is used in lieu of an actual value. The NOEC is, therefore, 25 g EP/ha. The EEC is greater than the calculated NOEC ($0.10 \times LC_{50}$), therefore, kresoxim-methyl is expected to pose a potential risk to ladybird beetles. The margin of safety is 0.014.

Terrestrial vascular plants: In order to be protective of terrestrial plants, the lowest EC_{25} was selected for the risk assessment. The EEC of 900 g a.i./ha. is greater than the most sensitive EC_{25} of 360 g a.i./ha, therefore, kresoxim-methyl would pose a potential risk to some non-target terrestrial plants when applied at the proposed maximum application rate. The margin of safety is 0.4.

6.4.3 Aquatic organisms

Non-target freshwater invertebrates: In order to be protective of *Daphnia magna*, the lowest NOEC of 0.055 mg a.i./L was selected for the risk assessment. Using the EEC of 0.08 mg a.i./L, the margin of safety is 0.69. Therefore, kresoxim-methyl will pose a risk to non-target freshwater invertebrates.

Fish: In order to be protective of fish, the lowest NOEC of 0.087 mg a.i./L was selected for the risk assessment. Using the EEC of 0.08 mg a.i./L, the margin of safety is 1.1 (Table 16). Therefore, kresoxim-methyl does not pose a risk to fish at the proposed maximum label application rate. In addition, the bioconcentration studies showed that kresoxim-methyl is unlikely to accumulate in fish tissues.

The LC_{50} for BF 490-1 was >102 mg BF 490-1/L. Therefore, according to the U.S. EPA classification scheme, the major transformation product is considered practically non-toxic to fish.

Algae: In order to be protective of all non-target freshwater algae, the most sensitive end point was selected for the risk assessment. The lowest end point was the NOEC for a freshwater diatom, which was 0.012 mg a.i./L. Using the EEC of 0.08 mg a.i./L, the margin of safety is 0.15. Therefore, kresoxim-methyl will pose a risk to freshwater algae.

Aquatic vascular plants: The toxicity end point for duckweed was the NOEC of 0.288 mg a.i./L. Because the EEC of kresoxim-methyl in water (0.08 mg a.i./L) is below the NOEC, kresoxim-methyl should not pose any risk to aquatic vascular plants at the proposed application rate. The margin of safety is 3.60.

6.4.4 Summary of risk to terrestrial and aquatic organisms

Margins of safety, using the estimated environmental concentrations and toxicity end points (NOEC), were used to determine the risk of kresoxim-methyl to terrestrial and aquatic non-target organisms (see Tables 6 and 7 of Appendix III). Kresoxim-methyl will not pose a risk to earthworms, honeybees, wild birds, wild mammals, fish or aquatic vascular plants. Kresoxim-methyl may pose a risk, however, to beneficial predators and parasites, some terrestrial plants, *Daphnia magna* and freshwater algae. Risks to terrestrial plants, *Daphnia magna*, and freshwater algae can be mitigated.

There are some toxicological concerns with the major transformation product, BF 490-1, because of its persistence in water and sediments and occurrence in the environment at high concentrations. Although three acute studies were submitted, the chronic effects of BF 490-1 are still unknown.

6.5 Risk mitigation

To mitigate the effects on non-target terrestrial and aquatic species, buffer zones should be observed. Buffer zones are determined by using the most sensitive end point, from submitted toxicity studies, which represents the non-target group at greatest risk. Buffer zones of 3 and 7 m are required to protect sensitive terrestrial and aquatic habitats, respectively.

7.0 Efficacy

7.1 Effectiveness against apple scab caused by *Venturia inaequalis* on apple

Forty-one trials conducted (between 1991–1998) in Canada and the United States (18 and 23, respectively) were submitted in support of the claims on apples. The location and number of these trials were as follows: British Columbia, 4; Nova Scotia, 5; Ontario, 3; Quebec, 6; California, 1; Michigan, 5; New York, 5; Ohio, 6; Pennsylvania, 4; Virginia, 1; and Washington, 1.

7.1.1 Kresoxim-methyl on a protectant schedule

Thirty-one trials included data on kresoxim-methyl on a protectant schedule. Eighteen trials demonstrated that there were significant differences in disease incidence between kresoxim-methyl and the untreated check. Thirteen trials provided data on the basis of more than five applications and lacked statistical analysis; therefore, those trials could be used only as supplemental data.

Application rate:

The data indicated that there were significant differences in disease incidence between kresoxim-methyl (all tested rates) and the untreated check. Fourteen trials provided data on the proposed low rate of 120 g a.i./ha and six trials on the high rate of 180 g a.i./ha.

The high rate provided an average of 96% disease control on fruit and 93% on foliage and the low rate provided an average of 89.5% disease control on fruit and 89% on foliage. Eleven trials demonstrated that 90.0 g a.i./ha consistently provided an equivalent percent disease control to the proposed rate of 120 g a.i./ha (96.2% on fruit and 93.8% on foliage) at low to moderate disease pressure. The rate of 90 g a.i./ha also had a higher percent disease control than the commercial standard (96.4% vs. 91.9% on fruit and 94.4% vs. 92.2% on foliage). Under high disease pressure the 120 and 180 g a.i./ha rates provided acceptable disease control in only two of seven and three of five disease ratings on fruits, respectively.

The available data support an application rate of 90 g a.i./ha under low to moderate disease pressure and 180 g a.i./ha under high disease pressure.

Number of applications and timing:

Eight trials included data on kresoxim-methyl applied at the proposed four applications per season as well as at the commercial standard. In those trials kresoxim-methyl at 90 g a.i./ha provided an average 94.6% (on fruit) and 94.8% (on foliage) disease control compared with 87% (on fruit) and 90.8% (on foliage) for the commercial standard. All trials done under high disease pressure used the proposed number of applications and timing.

The available data support the use of four applications per season of kresoxim-methyl at the rate of 90 g a.i./ha under low to moderate disease pressure and 180 g a.i./ha under high disease pressure at one-half inch green or when environmental conditions become favourable for primary scab and continued on a 10- to 14-day interval through second cover.

7.1.2 Curative applications of kresoxim-methyl

Eight trials (between 1994–1998) conducted in Canada and the United States (six and two, respectively) were submitted in support of the curative activity of kresoxim-methyl on apple scab.

Application rate:

Only one rate, 120 g a.i./ha, applied once, was tested in all trials. In seven trials, kresoxim-methyl applied at 120 g a.i./ha under low to moderate disease pressure provided a level of disease control significantly different from the untreated check. One trial under high disease pressure showed that the 120 g a.i./ha did not provide acceptable disease control; this is consistent with the data reviewed previously for protectant applications.

The available data (curative) support an application rate of 120 g a.i./ha under low to moderate disease pressure and 180 g a.i./ha under high disease pressure.

Application timing:

Seven different application times included in eight trials. There were significant differences between kresoxim-methyl and the untreated check in all the trials except one, which had high disease pressure. Six of eight trials demonstrated that kresoxim-methyl applied 48–120 h after infection provided equal or higher percent disease control on fruit than the commercial standard applied 96 h after infection. Four of six trials demonstrated that kresoxim-methyl applied 72–120 h after infection provided equal or higher percent disease control on foliage than the commercial standard at 96 h after infection. One trial showed that under high disease pressure the 120 g a.i./ha did not provide acceptable disease control 85 h after infection; this finding is consistent with the data reviewed previously for protectant applications.

The available data support the claim that kresoxim-methyl provides disease control if applied at 180 g a.i./ha up to 96 h after the infection period.

7.2 Effectiveness against powdery mildew caused by *Podosphaera leucotricha* on apple

Twenty trials conducted over eight years in Canada and the United States (3 and 17, respectively) were submitted in support of the powdery mildew control claims on apples. The location and number of these trials were as follows: British Columbia, 3; California, 4; New York, 6; Pennsylvania, 2; Virginia, 1; Washington, 3; West Virginia, 1. Fourteen trials demonstrated that there were significant differences in disease incidence between kresoxim-methyl and the untreated check. The remaining six trials provided data from more than six applications of kresoxim-methyl and lacked statistical analysis; therefore, those trials could be used only as supplemental data.

Application rate:

The data indicated that there were significant differences in disease incidence between kresoxim-methyl (all tested rates) and the untreated check. Ten trials provided data on the proposed low rate (120 g a.i./ha), four trials on 225 g a.i./ha and five trials on the proposed high rate (240 g a.i./ha).

Two trials compared the low and high rates under moderate and low disease pressure. There was no significant difference between the two rates; however, 240 g a.i./ha provided a higher level of disease control and the same level of control as the commercial standard in both trials. Four trials compared the 120 and 225 g a.i./ha rates. In all cases, the 225 g a.i./ha gave the same or higher level of disease control on leaves. Under high disease pressure, the 225 g a.i./ha rate provided a significantly higher control of fruit russetting.

The available data support an application rate of 120 g a.i./ha under low to moderate disease pressure and 225 g a.i./ha under high disease pressure.

Number of applications and timing:

Kresoxim-methyl applied at 120 and 225 g a.i./ha, using four applications per season, was significantly different from the untreated check in all cases and provided the same level of disease control as the commercial standard.

The available data support the use of four applications per season of kresoxim-methyl at the rate of 120 to 225 g a.i./ha starting at one-half inch green and continued on a 10 to 14 day interval through second cover.

7.3 Information on the occurrence or possible occurrence of the development of resistance

According to Regulatory Directive DIR99-06 on *Voluntary Pesticide Resistance Management Labelling Based on Target Site/Mode of Action*, the following statements will be incorporated on the Sovran[®] Fungicide label at next printing.

Group 11 FUNGICIDE (on the primary panel)

For resistance management, please note that Sovran[®] Fungicide contains a Group 11 fungicide. Any fungal population may contain individuals naturally resistant to Sovran[®] Fungicide and other Group 11 fungicides. A gradual or total loss of pest control may occur over time if these fungicides are used repeatedly in the same fields. Other resistance mechanisms that are not linked to site of action but specific for individual chemicals, such as enhanced metabolism, may also exist. Appropriate resistance-management strategies should be followed.

To delay fungicide resistance:

- Avoid application of more than two consecutive sprays of Sovran[®] Fungicide or other fungicides in the same group in a season.
- Fungicide use should be based on an IPM program that includes scouting, historical information related to pesticide use and crop rotation and considers cultural, biological and other chemical control practices.
- Monitor efficacy of all fungicides used in the disease management program against the targeted pathogen and record other factors which may influence fungicide performance and (or) disease development.
- If disease continues to progress after treatment with this product, do not increase the use rate. Discontinue use of this product, and switch to another fungicide with a different target site of action, if available.

- Contact your local extension specialist or certified crop advisors for any additional pesticide resistance-management and (or) IPM recommendations for specific crops and pathogens.
- For further information and to report suspected resistance, contact (company representatives) at (toll free number) or at (Internet site).

8.0 Toxic Substances Management Policy considerations

During the review of kresoxim-methyl and Sovran[®] Fungicide, the PMRA has considered the implications of the federal Toxic Substances Management Policy¹ and the PMRA Regulatory Directive DIR99-03 (*The Pest Management Regulatory Agency's Strategy for Implementing the Toxic Substances Management Policy*) and has concluded the following:

Kresoxim-methyl does not meet the criteria for persistence, as its $t_{1/2}$ values in water and sediment systems (up to 1.6 days) and soil (up to 11 days) are below the TSMP Track-1 cut-off criteria for water (≥ 182 days), sediment (≥ 365 days) and soil (≥ 182 days). No data were provided for kresoxim-methyl in air.

Kresoxim-methyl is not bioaccumulative. Studies have shown that the bioconcentration factors (BCF) are 220, 430 and 52, for whole fish, viscera and fillets, respectively, which are below the TSMP Track-1 cut-off criterion for BCF (≥ 5000). In addition, the $\log K_{ow}$ is 3.4, which is below the TSMP Track-1 cut-off criterion of ≥ 5.0 .

The toxicity of kresoxim-methyl is described in Sections 3.0 and 6.0 and Appendices I and III.

Kresoxim-methyl does not contain any by-products or microcontaminants of concern. Impurities of toxicological concerns are not expected to be present in the raw materials nor are they expected to be generated during the manufacturing process. The formulated product does not contain any formulants that are known to contain TSMP Track-1 substances.

In the environment, kresoxim-methyl forms one major transformation product, BF 490-1, which does not meet the criterion for persistence in soil. The DT_{50} of BF 490-1 in soil ranged from 35 to 55 days in field studies, which is below the TSMP Track-1 cut-off criterion for soil (≥ 182 days). However, the half-life of BF 490-1 in aerobic water (calculated at 330–385 days) exceeds the TSMP Track-1 cut-off criterion for persistence in water (≥ 182 days). A definitive half-life for BF 490-1 in sediment could not be determined. Persistence in any medium is a trigger for an examination of the

¹ The federal Toxic Substances Management Policy is available through Environment Canada's web site at www.ec.gc.ca/toxics.

bioaccumulation potential of BF 490-1: the log K_{ow} of BF 490-1 was examined and determined to be 2.95. As the log K_{ow} value does not meet TSMP criterion for bioaccumulation, the major transformation product BF 490-1 will not meet all the criteria for a Track-1 substance under the TSMP. Therefore, BF 490-1 cannot be considered to be a Track-1 substance.

In conclusion, the four TSMP Track-1 criteria for anthropogenicity, persistence, bioaccumulation, and toxicity are not met for either the parent compound, kresoxim-methyl, or for the major transformation product, BF 490-1. Thus, kresoxim-methyl and BF 490-1 are not considered to be Track-1 substances.

9.0 Proposed regulatory decision

The PMRA has carried out an assessment of available information in accordance with Section 9 of the Pest Control Products (PCP) Regulations and has found it sufficient pursuant to Section 18(b), to allow a determination of the safety, merit and value of the active ingredient kresoxim-methyl and the end-use product Sovran[®] Fungicide. The Agency has concluded that the use of the active ingredient kresoxim-methyl and the end-use product Sovran[®] Fungicide in accordance with the label has merit and value consistent with Section 18(c) of the PCP Regulations and does not entail an unacceptable risk of harm pursuant to Section 18(d). Based on the considerations outlined above, therefore, the use of the active ingredient kresoxim-methyl and the end-use product Sovran[®] Fungicide, for the control of apple scab and powdery mildew in apple orchards, is proposed for full registration, pursuant to Section 13 of the PCP Regulations.

The PMRA will accept written comments on this proposal up to 45 days from the date of publication of this document to allow interested parties an opportunity to provide input into the proposed registration decision for this product.

List of abbreviations

a.i.	active ingredient
ADI	allowable daily intake
ARfD	acute reference dose
BCF	bioconcentration factors
bw	body weight
CAS	Chemical Abstracts Service
CEPA	<i>Canadian Environmental Protection Act</i>
CV	coefficient of variation
d	day
DFR	dislodgeable foliar residue
DNA	deoxyribonucleic acid
DT ₅₀	dissipation time 50%
EC ₂₅	concentration effective against 25% of test organisms
EC ₅₀	median effective concentration
ECD	electron capture detection
EEC	expected environmental concentration
EP	end-use product
EPA	Environmental Protection Agency (U.S.)
EXPRES	Expert System for Pesticide Regulatory Evaluation and Simulation
FAO	Food and Agriculture Organization
FOB	functional observational battery
GAP	good agricultural practices
GC	gas chromatography
GIT	gastrointestinal tract
GST-P	glutathione S-transferase, placental form
GUS	Groundwater Ubiquity Score
h	hour
ha	hectare
HAFT	highest average field trial
HPLC	high performance liquid chromatography
ILV	interlaboratory validation
JMPR	Joint WHO/FAO Meeting on Pesticide Residues
K_{oc}	organic carbon adsorption coefficient
K_{ow}	octanol–water partition coefficient
LADD	lifetime average daily dose
LC	liquid chromatography
LC ₅₀	median lethal concentration
LD ₅₀	median lethal dose
LI	leaching index
LOAEL	lowest observed adverse effect level
LOD	limit of detection
LOQ	limit of quantitation
LP	leaching potential
MAS	maximum average score

M/L/A	mixer/loader/applicator
MOE	margin of exposure
MRL	maximum residue limit
MRM	multiresidue method
MS	mass spectrometry
nm	nanometre
NOAEL	no observed adverse effect level
NOEC	no observed effect concentration
NOEL	no observed effect level
OC	organic carbon content
PAI	pure active ingredient
pH	potential hydrogen
PHED	Pesticide Handlers Exposure Database
PHI	preharvest interval
pK_a	dissociation constant
PMRA	Pest Management Regulatory Agency
ppb	parts per billion
ppm	parts per million
Q_1^*	cancer estimate risk number
PIS	primary irritation score
r	correlation coefficient
r^2	coefficient of determination
R^2	regression coefficient
RAC	raw agricultural commodity
RfD	reference dose
ROC	residue of concern
RP	reversed phase
SGGT	serum γ -glutamyl transferase
TC	transfer coefficient
TGAI	technical grade of active ingredient
TSMP	Toxic Substances Management Policy
$t_{1/2}$	half-life
U.S.	United States
UV	ultraviolet
WHO	World Health Organization

Appendix I Toxicology

Metabolism			
<p>Technical kresoxim-methyl was moderately well absorbed in the GIT. Peak plasma levels were achieved in 1–8 h. It was widely distributed (majority remains in the GIT) with no sex related differences. It was rapidly excreted ($\pm 90\%$ administered dose in 48 h), mostly (66–81%) through feces, 9–33% in urine and none in expired air. It has negligible potential for accumulation, <1% administered dose remaining in the carcass by 120 h. It was completely metabolised to 34 metabolites, ultimately conjugated and eliminated as to sulfates and glucuronides. The alcohol–acid, and phenol–acid of the parent compound, and their glucuronides were the predominant final biotransformation products. The parent compound was the most toxicologically significant.</p>			
Study	Species and Strain and Doses	LD₅₀; mg/kg bw or LC₅₀; mg/L	Target Organ and Significant Effects and Comments
Acute studies: technical			
Oral	Rats, Wistar, 5/sex 5000 mg/kg bw (limit test)	Median Lethal Dose (LD ₅₀) > 5000 mg/kg bw in males and females	No mortality and no treatment related clinical signs or necropsy findings, low acute toxicity.
Dermal	Rats, Wistar, 5/sex 2000 mg/kg bw (limit test)	LD ₅₀ > 2000 mg/kg bw in males and females	No mortality, erythema at test site, no treatment related clinical signs or necropsy findings, low acute toxicity.
Inhalation	Rats, Wistar, 5/sex 2.04 and 5.6 mg/L	Median Lethal Concentration (LC ₅₀) > 5.6 mg/L	No mortality, increased respiration, nasal and ocular discharge occur during exposure, low acute toxicity.
Skin irritation	Rabbits, Vienna White, 2 males, 4 females 500 mg dose	primary irritation score (PIS) (24 and 48 h) = 0.0	Slight erythema in 1/6 animals one hour after dosing, resolved in 24 h, non-irritating to skin.
Eye irritation	Rabbits, Vienna White 0.1 mL (39 mg) dose	maximum average score (MAS) = 4/110	Slight erythema and ocular discharge in all animals within one hour, resolved within 72 h, mild eye irritant.
Skin sensitization (maximization test) of Mugnason and Kligman	Guinea pigs, Dunkin Hartley 0.1 mL intradermal, 5% test material induction and topical 0.3 g, 50% of test material challenge DNCB Positive Control	No erythema or edema 24 or 48 h after challenge	No evidence of sensitization, not a dermal sensitizer.
Acute studies: Sovran® BAS 490 02F (end-use product [EP])			
Oral	Rats, Wistar, 5/sex 5000 mg/kg bw (limit test)	LD ₅₀ > 5000 mg/kg [bw] in males and females	No mortality, some animals developed diarrhea, which resolved readily, no necropsy findings, low toxicity.
Dermal	Rats, Wistar, 5/sex 2000 mg/kg bw (limit test)	LD ₅₀ > 2000 mg/kg bw in males and females	No mortality and no treatment related clinical signs nor necropsy findings, low toxicity.
Inhalation	Rats, Wistar, 5/sex 2.04 and 5.6 mg/L	LC ₅₀ > 5.7 mg/L	No mortality, increased respiration in all animals, low acute toxicity.
Skin irritation	Rabbits, Vienna White 500 mg dose	PIS (24 and 48 h) = 0.0	Slight erythema in 1/6 animals one hour after dosing, resolved in 24 h, non-irritating.

Study	Species and Strain and Doses	LD ₅₀ ; mg/kg bw or LC ₅₀ ; mg/L	Target Organ and Significant Effects and Comments
Eye irritation	Rabbits, Vienna White 0.1 mL (39 mg) dose	MAS = 5/110	Slight erythema and ocular discharge in all animals within one hour, resolved within 72 h, mild eye irritant.
Skin sensitization (Buehler method)	Guinea pigs, Dunkin Hartley 0.5 mL of 60% test material, topical induction, and challenge DNCB Positive Control data used	No erythema or edema 24 or 48 h after challenge	No evidence of sensitization, not a dermal sensitizer.
Short-term toxicity			
Study	Species/Strain and Doses	NOAEL (mg/kg bw)/day	Target Organ and Significant Effects and Comments
90-d dietary	Mice, CrIBR, 10/sex/dose 0, 250, 1000, 4000 and 8000 ppm (0, 57, 230, 909 and 1937 mg/kg bw for males; 0, 80, 326, 1326 and 2583 mg/kg bw for females)	NOAEL = 1000 ppm (230 mg/kg bw/d for males) and 8000 ppm (2583 mg/kg bw/d for females)	Lowest observed adverse effect level (LOAEL) males = 4000 ppm on the basis of decreased bw and increased relative liver weight LOAEL females > 8000 ppm, on the basis of absence of toxic effects at highest dose tested.
21-d repeat-dose dermal	Rats, Wistar, 10/sex/dose 0 and 1000 mg/kg bw/d, 6 h/d	NOAEL = 1000 mg/kg/d in males and females	LOAEL > 1000 mg/kg bw/d, on the basis of absence of toxicity at highest dose tested (limit test).
90-d dietary	Rats, Wistar, 10/sex/dose 0, 500, 2000, 8000 and 16 000 (0, 36, 146, 577 and 1170 for males; 0, 43, 172, 672 and 1374 for females)	NOAEL = 2000 ppm (146 mg/kg/d) in males; 16 000 ppm (1374 mg/kg/d) in females	LOAEL males = 8000 ppm, on the basis of increased SGGT (also seen in other studies) LOAEL females > 16 000 ppm on the basis of absence of toxicity at highest dose.
12-month gavage	Dogs, beagle, 4/sex/dose 0, 1000, 5000, 25 000 (0, 27,138, 714 mg/kg bw/d for males; 0, 30, 146, 761 mg/kg bw/d for females)	NOAEL = 5000 ppm (138 mg/kg/d) in males; 25 000 ppm (761 mg/kg/d) in females	LOAEL males = 25 000 ppm, on the basis of decreased bw, decreased bw gain and decreased food efficiency LOAEL females > 25 000 ppm on the basis of absence of toxicity at highest dose.

Study	Species/Strain and Doses	NOAEL (mg/kg bw)/day)	Target Organ and Significant Effects and Comments
Chronic toxicity and oncogenicity			
18-month feeding	Mice, CrlBr, 50/sex/dose 0, 400, 2000 and 8000 ppm (0, 60, 304 and 1305 mg/kg bw/d in males; 0, 81, 400 and 1662 mg/kg bw/d in females)	Chronic effects NOAEL = 2000 ppm (304 mg/kg bw/d) in males; 400 ppm (81 mg/kg bw/d) in females Oncogenicity No oncogenic effects in either sex	LOAEL males = 8000 ppm (1305 mg/kg bw/d) on the basis of decreased bw and liver and adrenal amyloidosis. LOAEL females = 2000 ppm (400 mg/kg bw/d) on the basis of decreased bw. Not carcinogenic in mice.
Two-year feeding	Rats, Wistar, 20/sex/dose 0, 200, 800, 8000 and 16 000 ppm (0, 9, 36, 370 and 746 mg/kg bw in males; 0, 12, 48, 503 and 985 mg/kg bw in females)	Chronic effects NOAEL = 800 ppm (36 mg/kg bw/d) in males; 800 ppm (48 mg/kg bw/d) in females Oncogenicity NOAEL = 800 ppm (36 and 48 mg/kg bw in males and females, respectively)	LOAEL males = 8000 ppm on the basis of increased SGGT, increased liver weight and liver histopathology. LOAEL females = 8000 ppm on the basis of decreased bw, decreased bw gain and liver histopathology. LOAEL for liver carcinomas in both sexes was 8000 ppm (370 and 503 mg/kg bw in males and females, respectively).
Two-year feeding	Rats, Wistar, 50/sex/dose 0, 200, 800, 8000 and 16 000 ppm (0, 9, 36, 375 and 770 in males; 0, 12, 47, 497 and 1046 females)	Chronic effects NOAEL = 800 ppm (36 and 47 mg/kg for males and females, respectively) Oncogenicity NOAEL = 800 ppm (36 and 47 mg/kg for males and females, respectively) LOEL = 8000 ppm (375 and 497 mg/kg for males and females, respectively)	LOAEL in males = 8000 ppm (375 mg/kg bw/d)* on the basis of decreased bw and liver lesions. LOAEL in females = 8000 ppm (497 mg/kg bw) on the basis of decreased bw, decreased bw gain and liver histopathology. Increased incidence of liver carcinomas occurred in both sexes at 8000 ppm (375 and 497 mg/kg for males and females, respectively) and above.
Neurotoxicity			
Acute neurotoxicity	Rats, Wistar, 10/sex/dose 0, 500, 1000 and 2000 mg/kg bw via gavage	NOAEL = 2000 mg/kg bw/d for males and females, no acute neurotoxicity	LOAEL > 2000 mg/kg bw/d No effect on motor activity or functional observational battery (FOB) at highest dose, no other treatment related effect observed.

Study	Species/Strain and Doses	NOAEL (mg/kg bw)/day	Target Organ and Significant Effects and Comments
Subchronic neurotoxicity (90-d)	Rats, Wistar, 10/sex/dose 0, 1000, 4000 and 16 000 ppm (0, 72, 292 and 1180 mg/kg bw/d) in the diet	NOAEL (neurotoxicity) = 1180 mg/kg bw/d for males and females NOAEL (systemic toxicity) = 4000 ppm (292 mg/kg bw/d) No subchronic neurotoxicity	LOAEL (neurotoxicity) > 1180 mg/kg bw/d. No effect on motor activity or FOB at highest dose. LOAEL (systemic toxicity) = 16 000 (1180 mg/kg bw/d) on the basis of significant reductions in bw and bw gain.
Reproductive and developmental toxicity			
Multigeneration	Rats, Wistar, 25/sex/dose F ₀ ; 25/sex/dose, F ₁ 0, 50, 1000, 4000 and 16 000 ppm (0, 4.75, 95.4, 386 and 1552.3 males; 0, 5.3, 104.7, 426.9 and 1696.8 females)	Systemic effects Parental systemic NOAEL = 1000 ppm (104 mg/kg bw/d) Reproductive and developmental effects NOAEL = 1000 ppm (104 mg/kg bw/d)	LOAEL for parental systemic toxicity = 4000 ppm, on the basis of decreased bw, of F ₀ and F ₁ parental animals, decreased kidney weights F ₀ and increased SGGT F ₀ males. LOAEL for reproductive and developmental toxicity = 4000 ppm, on the basis of decreased pup weights (F _{1b} and F ₂) and delayed developmental landmarks (unfolding of ears [F _{1b}], auditory canal [F ₂] and eye opening [F _{1b}]). There were no effects on fertility, or reproductive performance at the highest dose 1696 mg/kg bw.
Teratogenicity	Rats, Wistar, 25/dose 0, 100, 400 and 1000 (mg/kg bw/d) via gavage	Maternal NOAEL = 1000 mg/kg bw/d Developmental NOAEL = 1000 mg/kg bw/d No teratogenic effects up to the highest dose tested	LOAEL maternal > 1000 mg/kg bw/d. No maternal toxicity highest dose tested. LOAEL developmental > 1000 mg/kg bw/d. No fetal toxicity at highest dose tested, no teratogenic effects at highest dose tested.
Teratogenicity	Rabbits, Himalayan, 15/dose 0, 100, 400 and 1000 mg/kg bw/d via gavage	Maternal NOAEL = 1000 mg/kg bw/d Developmental NOAEL = 1000 mg/kg bw/d No teratogenic effects up to the highest dose tested	LOAEL maternal > 1000 mg/kg bw/d. No maternal toxicity at highest dose. LOAEL developmental > 1000 mg/kg bw/d. No fetal toxicity at highest dose, no teratogenic effects at the highest dose tested.
Genotoxicity			
Study	Species/Strain and Doses	Doses Employed	Significant Effects and Comments
Ames test, point mutation	<i>Salmonella typhimurium</i> , TA 98 and TA 100 <i>Escherichia coli</i>	0, 20, 100, 500, 2500 and 5000 µg/plate ± S9	Negative
Ames test, point mutation	<i>Escherichia coli</i> CM 881 (WP2 trp uvrA pKM 101)	0, 20, 100, 500, 2500 and 5000 µg/plate ± S9	Negative

Study	Species/Strain and Doses	Doses Employed	Significant Effects and Comments
Mammalian chromosomal aberration (in vitro)	Human peripheral lymphocytes	0, 10, 20 and 40 µg/mL ± S9	Negative
Micronucleus assay (in vivo)	Mice CrIbR (5/sex/dose) single interperitoneal dose	0, 500, 1000 and 2000 mg/kg bw with cells harvested at 16, 24, 48 and 72 h post-treatment	Negative for micronuclei, clinical signs of toxicity were observed in all dose groups after 30 minutes.
UDS in vitro (DNA damage and repair)	Rat hepatocytes (Wistar)	0.33, 1.0, 3.33, 10.0, 33.3 and 100.0 µg/mL	Negative
UDS ex vivo (DNA damage and repair)	Rats, Wistar, single dose oral gavage	0, 20, 200 and 1000 mg/kg bw	Negative
UDS ex vivo (DNA damage and repair)	Rats, Wistar, 3/group, three-week feeding	0, 200 and 16000 ppm (0, 4.79 and 441.87 mg/kg bw)	Negative
Mammalian cytogenetics (in vitro)	CHO/HGPRT	0.1, 0.5, 1.0, 5.0, 10.0 and 100.0 µg/mL ± S9 or 1.0, 2.15, 4.64, 10.0, 21.5, 46.4 and 100.0 µg/mL ± S9	Negative
Special studies			
Study	Species/Strain and Doses	NOAEL (mg/kg bw)/day	Target Organ/Significant Effects and Comments
Hepatocyte proliferation S-phase response (single dose)	Rats, Wistar, 3 males/dose 0, 20, 200 and 1000 mg/kg bw by gavage	NOAEL = 20 mg/kg bw, increased hepatocyte proliferation at 200 mg/kg bw and above	LOAEL = 200 mg/kg bw on the basis of two-fold increase in S-phase response at 200 mg/kg bw/d and above after three weeks.
Hepatocyte proliferation S-phase response (three-week dosing)	Rats, Wistar, young adult, 3-5/dose 0, 200 and 16 000 ppm (0, 15 and 1140 mg/kg bw/d) in the diet	NOAEL = 200 ppm (15 mg/kg bw), 2-3 fold increase in hepatocyte proliferation at 1140 mg/kg bw	LOAEL = 16 000 ppm (1140 mg/kg bw/d) on the basis of increased S-phase response (hepatocyte proliferation).
Hepatocyte proliferation (S-phase response) (three-week dosing and two-week recovery (from JMPR report)	Rats, Wistar, 16 months old, 3-5/dose 0, 200 and 16 000 ppm mg/kg bw/d in diet	NOAEL = 200 ppm (15 mg/kg bw), 2-3 fold increase in hepatocyte proliferation at 1140 mg/kg bw	LOAEL = 16 000 ppm (1140 mg/kg bw/d) on the basis of increased S-phase response. S-phase induction was reversible within the recovery period.
Tumour initiation	Rats, Wistar, 10/sex 2388 mg/kg bw via gavage	NOAEL (tumour initiation) > 2388 mg/kg bw Not a tumour initiator	LOAEL > (tumour initiation) 2388 mg/kg bw No increase in glutathione S-transferase, placental form (GST-P) positive foci in liver.
Tumour promotion	Rats, Fischer, at 0, 200, 800, 8000 and 16 000 ppm (0, 10.7, 42.5, 430 and 886 mg/kg bw/d) for six weeks, via diet	NOAEL (tumour promotion) = 800 ppm (42.5 mg/kg bw/d) A possible tumour promoter	LOAEL (tumour promotion) = 8000 ppm (430 mg/kg bw/d) on the basis of a dose related increase in GST-P positive foci at 8000 ppm and above.

Study	Species/Strain and Doses	NOAEL (mg/kg bw)/day	Target Organ/Significant Effects and Comments
<p>Recommendation for ADI for non-carcinogenic end points The ADI was 0.36 mg/kg bw/d, on the basis of the NOAEL of 36 mg/kg bw/d set in a two-year rat study and using a 100-fold uncertainty factor.</p> <p>Q₁* value: The Q₁* assigned by the U.S. EPA for kresoxim-methyl was 2.90×10^{-3}, on the basis of female rat liver tumour rates from the two-year oncogenicity study (Federal Register, vol. 64, no. 111, June 10, 1999).</p> <p>ARfD: An ARfD was deemed unnecessary. No acute end point was selected, as no adverse effects resulting from single exposure were identified in the short-term toxicity studies and acute neurotoxicity study in rats, nor in developmental toxicity studies in the rat and the rabbit.</p>			

Appendix II Residues

<p>Plant metabolism The unchanged parent compound accounted for significant residues in wheat, grapes and Mutsu variety apples, but not in Macintosh variety apples. Also, differences in the quantitative nature of residues were demonstrated by comparison of the two apple metabolism studies. The ROC in apples was defined as parent, 490M1, 490M2 (free and glucose conjugated) and 490M9 (free and glucose conjugated).</p>					
Matrix	Total rate (kg a.i./ha)	PHI (days)	Identified total radioactive residues [¹⁴ C] kresoxim-methyl label		
Apples (MacIntosh)	3.91	14	0.69 ppm		
Apples (Mutsu)	2.4	14	0.32 ppm		
<p>Confined crop rotation studies Not applicable.</p>					
<p>Multiresidue methods for residue analysis Recoveries of kresoxim-methyl residues in grapes ranged from 83 to 106% at spiking levels of 0.05–0.5 ppm using Protocols D and E. Recoveries in ground beef ranged from 107 to 121% at the same spiking levels using Protocol F.</p>					
<p>Methods for residue analysis of plants and plant products Data gathering method 350/3-US: HPLC with UV detection (LOQ = 0.05 ppm per analyte; LOD = 0.025 ppm per analyte).</p>					
Matrix	Apples	Apple juice	Grapes	Grape wine, juice (must) and marc	Pecans
Spiking levels (ppm)	0.05–5.0	0.05–5.0	0.05–5.0	0.05–5.0	0.05–0.15
Mean recoveries (%)	81–105	74–98	70–105	76–100	90–130
<p>Interlaboratory validation Interlaboratory validations indicated good reliability and reproducibility.</p>					
<p>Acceptability as an enforcement method Recommended. Enforcement method equivalent to data gathering method.</p>					
<p>Freezer storage stability tests Metabolism studies indicated that residues of kresoxim-methyl were stable in homogenized apple and grape samples when stored at less than –5°C for 30 and 34 months, respectively. Stability of kresoxim-methyl residues from the freezer storage stability studies are shown below.</p>					
Matrix	Storage interval (months)	Temperature (°C)	Spiking levels (ppm)	Corrected recovery in stored samples (%)	
apple	1–24	–20	1	98	
apple	2–12	less than –10	0.34–1.0	80–118	
wet apple pomace	2–12	less than –10	1	84–102	
apple juice	2–12	less than –10	1	78–105	
grape	2–12	less than –10	1	92–114	
grape	2–9	less than –10	0.34–0.52	75–118	

Freezer storage stability tests				
Metabolism studies indicated that residues of kresoxim-methyl were stable in homogenized apple and grape samples when stored at less than -5°C for 30 and 34 months, respectively. Stability of kresoxim-methyl residues from the freezer storage stability studies are shown below.				
Matrix	Storage interval (months)	Temperature (°C)	Spiking levels (ppm)	Corrected recovery in stored samples (%)
pecan	2-6	less than -10	1	97-100
pecan	6	less than -10	0.25-0.47	82-90
Animal metabolism				
In goats, kresoxim-methyl was extensively metabolized, with the predominant metabolites being 490M1, 490M2 and 490M9. Little or no parent compound was observed in tissues. The ROC was defined as kresoxim-methyl, 490M1, 490M2 and 490M9.				
Matrix	Dosing levels (mg/kg bw/d)	Percent of administered dose (ppm)		
Tissues	0.25, 0.31, 25.0	<0.10-0.11 (0.19-21.41)		
Milk	0.25, 0.31, 25.0	0.03-<0.10 (0.003-0.191)		
Feces	0.25, 0.31, 25.0	18-25		
Urine	0.25, 0.31, 25.0	59-70		
Methods for residue analysis of animal matrices				
Data gathering method 354/1-US for milk: HPLC with UV detection (LOQ = 0.002 ppm per analyte; LOD = 0.001 ppm per analyte).				
Matrix	Whole milk			
Spiking levels (ppm)	0.002-0.1			
Mean recoveries (%)	67-118			
CV (%)	2.3-23.0			

Interlaboratory validation Interlaboratory validations indicated good reliability and reproducibility.						
Acceptability as an enforcement method Recommended. Enforcement method equivalent to data gathering method.						
Data gathering method 354/2 for animal tissues: HPLC with UV detection (LOQ = 0.01 ppm per analyte; LOD = 0.002 ppm per analyte).						
Matrix	Skeletal muscle	Liver	Kidney	Fat		
Spiking levels (ppm)	0.01–1.0	0.01–1.0	0.01–1.0	0.01–1.0		
Mean recoveries (%)	80–96	72–97	69–118	53–109		
CV (%)	3.3–14.8	3.4–19.1	8.5–30.3	8.9–26.9		
Interlaboratory validation Interlaboratory validations indicated good reliability and reproducibility for skeletal muscle and liver, but poor reliability and reproducibility for kidney and fat.						
Acceptability as an enforcement method Not recommended (MRM recommended for analyses of animal tissues).						
Freezer storage stability tests						
Matrix	Storage interval (months)	Temperature (°C)	Spiking levels (ppm)	Corrected recovery in stored samples (%)		
Milk	12	less than –20	0.02	108–113		
Liver	13	less than –20	0.10–0.12	67–77		
Kidney	13	less than –20	0.10–0.12	67–101		
Skeletal muscle	13	less than –20	0.10–0.12	90–94		
Subcutaneous fat	13	less than –20	0.10–0.12	82–95		
Cattle feeding study Lactating dairy cows fed kresoxim-methyl in a feed premix for 28–29 days at doses of 0, 6, 18 or 60 ppm.						
Dose level (mg/kg/d)	Highest measured residue concentration (ppm)					
	Whole milk	Peritoneal fat	Subcutaneous fat	Skeletal muscle	Kidney	Liver
6	<0.002	<0.01	<0.01	<0.01	0.034	nd
18	<0.004	0.041	<0.01	<0.01	0.156	0.08
60	<0.004	0.134	0.03	<0.01	0.387	0.04

Hen feeding study Because there are no poultry feed items associated with this petition, no data depicting the magnitude of kresoxim-methyl residues in poultry commodities were required.										
Number of residue field trials by region										
Zones	1	0.04	2	5	0.208	5B	9	10	11	Total
Required	1	1		4		3			3	12
Submitted	4		2		4		2	2	6	20
Supervised residue trials on apples										
Commodity and portion analysed	Formulation	Application			PHI (days)	Highest residues measured (ppm) (several replicates of each sample)				
		No.	Total rate (kg a.i./ha)	Percent GAP						
U.S. trials										
Apples	490 02F	8	0.9	100	30	0.22				
	490 02F	152	0.88	98	30	0.43				
	490 02F	16	0.88	98	10, 20	0.35				
	490 02F	16	0.88	98	40, 60	0.15				
Processing studies Apples were treated at 4.48 kg a.i./ha (5× GAP) with a PHI of 30 days. Grapes were treated at 2.69 kg a.i./ha (3× U.S. GAP) with a PHI of 14 days.										
Matrix	Concentration factor		Expected residue (ppm)		Proposed MRL					
whole apple	—		<0.5		0.5					
apple juice	0.2		<0.15		0.15					
whole grape	—		<0.8		1					
grape juice	0.8		0.6		1					
raisins	1.6		1.3		1.5					
Dietary cancer risk assessment Using DEEM™ Software (Version 6.77) and on the basis of food consumption data from the 1994–1996 Continuing Survey of Food Intake by Individuals ($Q_1^* = 0.0029$ mg/kg bw), Tier II using supervised trial mean residues for raw agricultural commodities, proposed MRLs for processed commodities and EEC for drinking water.										
	Total Population	All infants (<1 year)	Children (1–6 years)	Children (7–12 years)	Males (13–19 years)	Males (20+ years)	Seniors 55+			
Lifetime risk	3.88×10^{-7}	7.16×10^{-7}	1.29×10^{-6}	5.41×10^{-7}	3.40×10^{-7}	2.71×10^{-7}	2.45×10^{-7}			

No acute dietary risk assessment was conducted, since an acute exposure hazard was not identified in the toxicology review.

Proposed MRLs

Commodity	Proposed Canadian MRL (ppm)	U.S. tolerance (ppm)
Apples	0.5	0.5
Grapes	1	1
Pecans	0.15	0.15
Apple juice	0.15	None
Raisins	1.5	1.5
Meat and meat by-products of cattle, goats, hogs, horses, sheep	0.03	0.01
Milk	0.004	None

Appendix III Environmental Assessment

Table 1 Summary of transformation and mobility data for kresoxim-methyl and the formulated end-use product

Title	Value	Comments
Soil: kresoxim-methyl		
Phototransformation on soil (25°C)	$t_{1/2} = 70.4$ days*	Not a principal route of transformation
Aerobic soil biotransformation	Label A: $DT_{50} \sim 15$ h Label B: $DT_{50} = 4.7$ days	Kresoxim-methyl is non-persistent
Mobility (adsorption or desorption)	Loam: organic carbon adsorption coefficient (K_{oc}) = 249 mL/g organic carbon content (OC)	Low to moderate mobility in loam
	Sand: $K_{oc} = 320$ mL/g OC	Moderate to high mobility in sand
	Loamy sand: $K_{oc} = 541$ mL/g OC	Low to moderate mobility in loamy sand
	Clay: $K_{oc} = 567$ mL/g OC	Low mobility in clay
Soil: end-use product		
Canadian field dissipation (0.26 kg a.i./ha × four applications)	Nova Scotia: $DT_{50} = 1$ day Ontario: $DT_{50} < 1$ day British Columbia: $DT_{50} = 11$ days	Sovran® is non-persistent in soil under field conditions
U.S. field dissipation (0.269 kg a.i./ha × four applications)	New York: $DT_{50} = 4.5$ h Oregon: $DT_{50} = 2.9$ days California: $DT_{50} < 1$ h	Sovran® is non-persistent in soil under field conditions
Water: kresoxim-methyl		
Hydrolysis (25°C)	pH 5: $t_{1/2} = 874$ days pH 7: $t_{1/2} = 32$ days pH 9: $t_{1/2} = 9$ h	Principal route of transformation at pH 9
Phototransformation in water	Label A: $t_{1/2} = 14.8$ days* Label B: $t_{1/2} = 59.6$ days*	Not a principal route of transformation
Aerobic water and sediment biotransformation	Loam system: $DT_{50} = 1.5$ days Sand system: $DT_{50} = 1.6$ days	Kresoxim-methyl is non-persistent
Anaerobic sediment/water biotransformation	Label A: $DT_{50} = 0.9$ days Label B: $DT_{50} = 1.3$ days	Kresoxim-methyl is non-persistent

* Under conditions of 12 h light : 12 h dark

Table 2 Summary of transformation and mobility data for the major transformation product, BF 490-1

Title	Value	Comments
Soil		
Aerobic soil biotransformation	Label A: $DT_{50} = 131$ days Label B: $DT_{50} = 58.8$ days	Moderately persistent
Mobility in U.S. soils (adsorption and desorption)	Loam: $K_{oc} = 33$ mL/g OC	High to very high mobility in loam
	Sand: K_{oc} not determined	High mobility was reported in sand
	Loamy sand: $K_{oc} = 69$ mL/g OC	High to very high mobility in loamy sand
	Clay: $K_{oc} = 44$ mL/g OC	High to very high mobility in clay
Mobility in German standard soils (adsorption and desorption)	Sandy loam with low OC: K_{oc} not determined	Very high mobility in sandy loam soils with low organic carbon (0.90%) content
	Sandy loam with high OC: $K_{oc} = 24$ mL/g OC	High to very high mobility in sandy loam soils with high organic carbon (2.60%) content
	Loamy sand: K_{oc} not determined	Very high mobility in loamy sand
	Clayey loam: $K_{oc} = 17$ mL/g OC	High to very high mobility in clayey loam
Mobility (leaching)	not determined	High mobility in aged soils
Canadian field dissipation (0.26 kg a.i./ha \times four applications)	Nova Scotia: $DT_{50} = 55$ days Ontario: $DT_{50} = 35$ days British Columbia: $DT_{50} = 56$ days	BF 490-1 is moderately persistent in soil under field conditions
Water		
Aerobic water and sediment biotransformation	Loam system: $DT_{50} \sim 462$ days Sand system: $DT_{50} > 93$ days	Moderately persistent to persistent
Anaerobic sediment and water biotransformation	Label A: $DT_{50} = 98$ days Label B: $DT_{50} = 130$ days	Moderately persistent

Table 3a Maximum EECs of kresoxim-methyl on vegetation and other food sources immediately after four applications at the Canadian maximum label rate of 0.225 kg a.i./ha

Environmental Compartment	EEC fresh weight (mg a.i./kg) ^a	Fresh/Dry weight ratios	EEC dry weight (mg a.i./kg)
short range grass	193	3.3 ^b	636
leaves and leafy crops	101	11 ^b	1110
long grass	88.2	4.4 ^b	388
forage crops	46.8	5.4 ^b	253
small insects	46.8	3.8 ^c	178
Pods with seeds	9.63	3.9 ^c	37.6
large insects	8.01	3.8 ^c	30.4
grain and seeds	8.01	3.8 ^c	30.4
fruit	5.58	7.6 ^c	42.4

^a Based on correlations reported in Hoerger and Kenaga (1972) and Kenaga (1973).

^b Fresh/dry weight ratios from Harris (1975).

^c Fresh/dry weight ratios from Spector (1956).

Table 3b Expected environmental concentrations (mg a.i./kg dw) of kresoxim-methyl on food sources and in the diet of wild birds and mammals immediately after four applications at the Canadian maximum label rate of 0.225 kg a.i./ha (no transformation)

Organism	Food item	Percent of diet	EEC		
			Food type	Each	Total
Bobwhite quail	small insects	301555	17825330.4	53.4	108
	forage crops			38.0	
	grain			16.7	
Mallard duck	large arthropods	3070	30.4	9.12	30.4
	grain		30.4	21.3	
Mouse	short grass	255025	63630.41	15915.22	451
	grain and seeds		11	77	
	leaves and leafy crops				
Rat	short grass	702010	636	445	454
	grain and seeds		30.4	6.08	
	large insects		30.4	3.04	

Table 4 Summary of effects of kresoxim-methyl on terrestrial non-target species

Organism	Organism and study	NOEC or NOEL	LC ₅₀ or LD ₅₀	Interpretation and comments
Birds	bobwhite quail; acute oral	NOEL = 2150 mg a.i./kg bw	LD ₅₀ >2150 mg a.i./kg bw	practically non-toxic*
	bobwhite quail; acute dietary	NOEC = 5000 mg a.i./kg diet	LC ₅₀ >5000 mg a.i./kg diet	practically non-toxic*
	mallard; acute dietary	NOEC = 5000 mg a.i./kg diet	LC ₅₀ >5000 mg a.i./kg diet	practically non-toxic*
	bobwhite quail; reproduction	LOAEL = 50 mg a.i./kg diet NOEL not determined	not calculated	effects observed
	mallard; reproduction	LOEL = 500 mg a.i./ kg diet NOEL = 100 mg a.i./ kg diet	not calculated	—
Mammals	rat; acute oral	NA	LD ₅₀ : >5000 mg a.i./kg bw	low acute toxicity
	rat exposed to EP; acute oral	NA	LD ₅₀ : >5000 mg a.i./kg bw	low acute toxicity
	mouse; 90-d dietary	NOAEL = 1000 mg/kg dw of diet in males; 8000 mg/kg dw of diet in females	NA	increased relative liver weight in males
	rat; 90-d dietary	NOAEL = 2000 mg/kg dw of diet in males; 16 000 mg/kg dw of diet in females	NA	increased levels of SGGT in males
	rat; multigenerational reproduction	Systemic effects NOAEL = 104 mg/kg bw/d in females Reproductive effects NOAEL = 1552.3 mg/kg bw/d in males; 1696.8 mg/kg bw/d in females	NA	decreased kidney, body, and pup weight; increased SGGT in males; delayed developmental landmarks
Invertebrates	earthworms; acute	≥937 mg a.i./ kg	>937 mg a.i./ kg	—
	earthworms; acute exposure to BF 490-1	≥1000 mg BF 490-1/kg	>1000 mg BF 490-1/kg	—
	earthworms; acute exposure to EP	250 mg EP/kg	644 mg EP/kg	—
	honeybees; acute contact	0–25 µg a.i./bee	>25 µg a.i./bee	relatively non-toxic**
	honeybees; acute contact with EP	413.5 µg EP/bee	>413.5 µg EP/bee	relatively non-toxic**
	honeybees; acute oral with EP (two studies)	(1) not reported (2) 410 µg EP/bee	(1) >98 µg EP/bee (2) >410 µg EP/bee	relatively non-toxic**

Organism	Organism and study	NOEC or NOEL	LC ₅₀ or LD ₅₀	Interpretation and comments
	predators and parasites	not determined	not determined	effects include mortality, reduction in fertility, decline in number of offspring, variable recovery
Terrestrial plants	soybean, lettuce, radish, tomato, cucumber, cabbage, oat, ryegrass, corn, onion; seed germination, seedling survival, plant height, and dry weight (three studies)	(1) not determined (2) 400 g a.i./ha (3) 390 g a.i./ha	(1) not determined (2) concentration effective against 25% of test organisms (EC ₂₅) = 360 g a.i./ha (3) EC ₂₅ > 390 g a.i./ha	significant reduction in dry weight of lettuce

* According to U.S. EPA classification scheme

** Classification according to Atkins et al. 1981

Table 5 Summary of effects of kresoxim-methyl on aquatic non-target species

Organism	Organism and study	NOEC	LC ₅₀ or EC ₅₀	Interpretation and comments
Freshwater invertebrates	<i>Daphnia magna</i> ; 48-h, flow-through	160 µg a.i./L	332 µg a.i./L	highly toxic*
	<i>Daphnia magna</i> ; acute, static with BF 490-1	not calculated	>100 mg BF 490-1/L	practically non-toxic*
	<i>Daphnia magna</i> ; 21-d	55–107 µg a.i./L	not calculated	effect on reproduction at concentrations greater than 107 µg a.i./L
Freshwater fish	Rainbow trout; acute, flow-through	104 µg a.i./L	190 µg a.i./L	highly toxic*
	Rainbow trout; acute, static with BF 490-1	102 mg BF 490-1/L	>102 mg BF 490-1/L	practically non-toxic*
	Bluegill sunfish; acute, flow-through	388 µg a.i./L	499 µg a.i./L	highly toxic*
	Fathead minnow; early life-stage toxicity	87 µg a.i./L (survival of larvae and growth)	>87 µg a.i./L	survival at the highest treatment (160 µg/L) was significantly less than controls
	Rainbow trout; bioaccumulation study, flowthrough	NA	NA	bioconcentration of BAS 490 F unlikely
Freshwater algae	<i>Navicula pelliculosa</i> (diatom); Tier II	12 µg a.i./L	29.2 µg a.i./L	—
	<i>Selenastrum capricornutum</i> (green alga); Tier II	12.2 µg a.i./L	59.4 µg a.i./L	—
	<i>Anabaena flos-aquae</i> (blue-green alga); Tier II	295 µg a.i./L	>295 µg a.i./L	—
Vascular plants	<i>Lemna gibba</i> (duckweed); Tier II	288 µg a.i./L	>288 µg a.i./L	—

* According to U.S. EPA classification scheme

Table 6 Summary of risk assessment to terrestrial non-target species

Organism	Type of study	NOEC or NOEL	EEC	Margin of safety	Comments
Earthworms	artificial soil	937 mg a.i./kg soil	0.20 mg a.i./kg soil	4690	no risk
Bees	acute contact	28 kg a.i./ha	0.90 kg a.i./ha	31.1	no risk
Predatory mites	laboratory exposure	107.1 g EP/ha	1350 g EP/ha	0.079	potential risk
Ladybird beetles	laboratory exposure	25 g EP/ha	1800 g EP/ha	0.014	potential risk
Bobwhite quail	acute oral	2150 mg a.i./kg bw	108 mg a.i./kg dw	204 days	no risk
	acute dietary	5000 mg a.i./kg dw	108 mg a.i./kg dw	46.3	no risk
	chronic reproduction	50 mg a.i./kg dw (LOAEL)	108 mg a.i./kg dw	not applicable	no risk on the basis of exposure and use patterns and rate of transformation
Mallard duck	acute dietary	5000 mg a.i./kg dw	30.44 mg a.i./kg dw	164	no risk
	chronic reproduction	100 mg a.i./kg dw	30.44 mg a.i./kg dw	3.29	no risk
Rats	acute oral toxicity	5000 mg a.i./kg bw	454 mg a.i./kg dw	>220 days	no risk
	90-d dietary (male)	2000 mg a.i./kg dw		4.4	no risk
	90-d dietary (female)	16 000 mg a.i./kg dw		35.2	no risk
	chronic reproduction	1000 mg a.i./kg dw		2.2	no risk
Mice	90-d dietary (male)	1000 mg a.i./kg dw	451 mg a.i./kg dw	2.2	no risk
	90-d dietary (female)	8000 mg a.i./kg dw		17.7	no risk
Non-target terrestrial plants (lettuce)	Tier II	0.36 kg a.i./ha (EC ₂₅)	0.90 kg a.i./ha	0.4	potential risk in the absence of mitigation

Note 1: For predaceous mites, the NOEC was estimated as 10% of the LC₅₀ calculated from a linear regression model on the basis of mortality of adult mites from Ufer 1994 and Kühner 1993 ($R^2 = 0.72$). The EEC for predaceous mites was on the basis of three applications of the EP at 450 g EP/ha over the adult lifespan of approximately 20 days (Kain and Nyrop 1995, Weeden et al. 1998).

Note 2: For ladybird beetles, the NOEC was estimated as 10% of the EC₅₀, as calculated from the ratio of 60% decline in fertility at an application rate of 300 g EP/ha, assuming a linear dose–response. The EEC for ladybird beetles is the maximum seasonal application rate because over the predicted lifespan of the adult beetle of several weeks to months (Hoffmann and Frodsham 1993), the adult could potentially be exposed to the maximum recommended four applications of the EP (1800 g EP/ha).

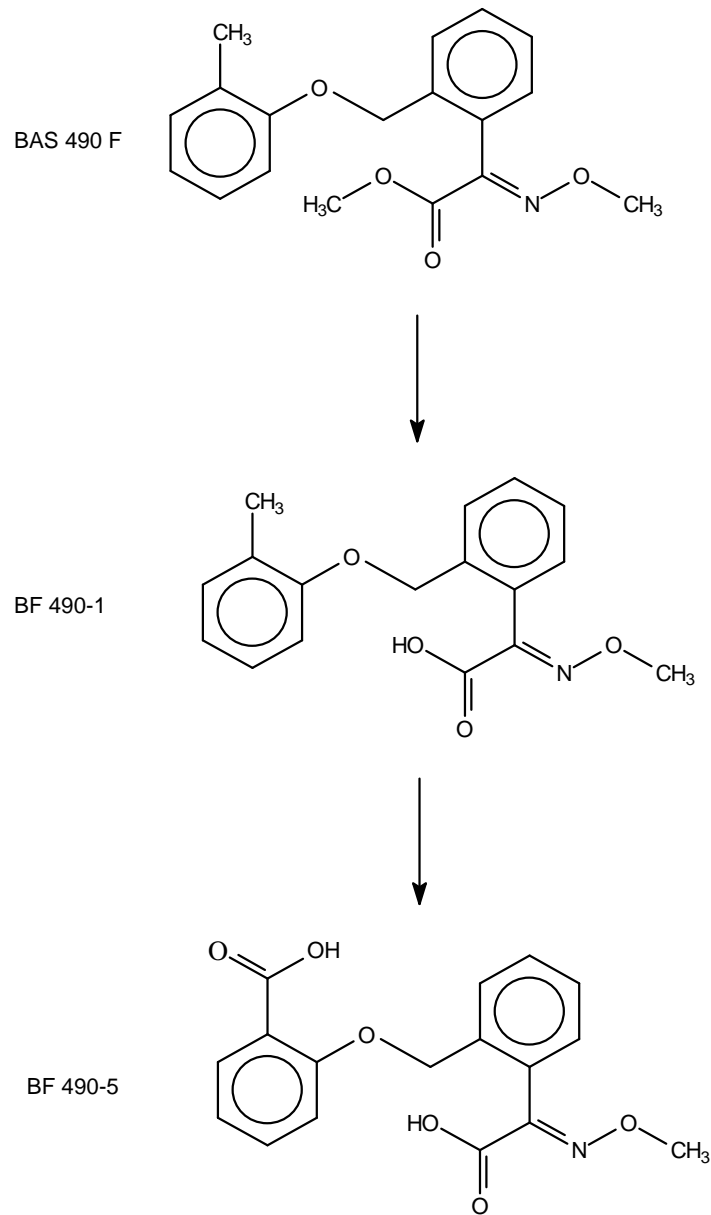
Note 3: The assessment of the risk of kresoxim-methyl to wild mammals is on the basis of the evaluation of mammalian toxicity studies by the Health Evaluation Division.

Note 4: For acute toxicity studies with birds and mammals, the margin of safety is reported with the units of days. LD₅₀ and NOEC values, as well as food consumption and mean body weights of control animals, were used to determine the amount of time required for a wild animal to accumulate a toxic dose of kresoxim-methyl if exposed to food sources that are contaminated with kresoxim-methyl.

Table 7 Summary of risk assessment to aquatic non-target species

Organism	Type of study	NOEC or NOEL	EEC	Margin of safety	Comments
Non-target freshwater invertebrates (<i>Daphnia magna</i>)	chronic flow-through	0.055 mg a.i./L	0.08 mg a.i./L	0.69	potential risk in the absence of mitigation
Fish (fathead minnow)	early life-stage toxicity	0.087 mg a.i./L	0.08 mg a.i./L	1.1	no risk
Algae (freshwater diatom)	Tier II	0.012 mg a.i./L	0.08 mg a.i./L	0.15	potential risk in the absence of mitigation
Aquatic vascular plants (duckweed)	Tier II	0.288 mg a.i./L	0.08 mg a.i./L	3.6	no risk

Figure 1. Environmental transformation pathway of kresoxim-methyl (BAS 490 F) for field studies, as proposed by the applicant.



References

- Atkins, E.L., Kellum, D. and Atkins, K.W. 1981. Reducing pesticide hazards to honey bees: mortality prediction techniques and integrated management strategies. University of California Division of Agricultural Sciences Leaflet No. 2883. 22 pp.
- Gustafson, D.I. 1989. Groundwater ubiquity score: a simple method for assessing pesticide leachability. *Environmental Toxicology and Chemistry*, **8**: 339–357.
- Hoerger, F. and Kenaga, E.E. 1972. Pesticide residues on plants: correlation of representative data as a basis for estimation of their magnitude in the environment. *In* (F. Coulston and F. Korte, eds.) *Environmental quality and safety: chemistry, toxicology and technology*. Vol. I. Global aspects of chemistry, toxicology and technology as applied to the environment. Georg Thieme Publishers, Stuttgart, and Academic Press, New York. pp. 9–28.
- Hoffmann, M.P. and Frodsham, A.C. 1993. *Coccinella septempunctata* (Coleoptera: Coccinellidae). *In* *Biological control: a guide to natural enemies in North America*. Cornell University. From M.P. Hoffmann and A.C. Frodsham. 1993. Natural enemies of vegetable insect pests. Cooperative Extension, Cornell University, Ithaca, N.Y. <http://www.nysaes.cornell.edu/ent/biocontrol/predators/c7.html>. Accessed on Sept. 1, 1999.
- Kenaga, E.E. 1973. Factors to be considered in the evaluation of the toxicity of pesticides to birds in their environment. *In* (F. Coulston and F. Korte, eds.) *Environmental quality and safety: global aspects of chemistry, toxicology and technology as applied to the environment*. Vol. II. Georg Thieme Publishers, Stuttgart, and Academic Press, New York. pp. 166–181.
- Kain, D.P. and Nyrop, J.P. 1995. Predatory mites: *Typhlodromus pyri* (Scheuten), *Amblyseius fallacis* (German), *Typhlodromus occidentalis* (Nesbitt) (Acari: Phytoseiidae), and *Zetzellia mall* (Ewing) (Acari: Stigmaeidae). Insect Identification Sheet No. 123. <http://www.nysaes.cornell.edu/ipmnet/ny/fruits/FruitFS/predmites.html>. Accessed on Sept. 1, 1999.
- Kühner, C. 1993. Study of the side effects of BAS 490 02 F on the predatory mite *Typhlodromus pyri*, SCHEUTEN: (Acari Phytoseiidae) in the laboratory. Arbeitsgemeinschaft, GAB Biotechnologie GmbH und IFU Umweltanalytik GmbH, Eutinger, Germany, Dec. 15, 1993, BASF Canada, Inc., Toronto, Ontario, BASF Reg. Doc. No. 93/11615. 23 pp. Unpublished. Submitted to Canada July 28, 1997, DACO 9.2.5.
- Ufer, A. 1994. Mortality of different life stages of the predatory mite *Typhlodromus pyri* Scheuten (Phytoseiidae) after treatment with BAS 490 02 F. BASF Aktiengesellschaft, Limburgerhof, Germany, November 1994, BASF Canada, Inc., Toronto, Ontario, BASF Reg. Doc. No. 94/11069, 16 pp. Unpublished. Submitted to Canada July 28, 1997, DACO 9.2.5.

Urban, D.G. and Cook, N.J. 1986. Ecological risk assessment. United States Environmental Protection Agency, EPA 540.9-85-001. 96 pp.

Weeden, X., Shelton, X. and Hoffman, X. (eds.). 1998. *Galendromu (=Typhlodromus) pyri* (Acarina: Phytoseiidae) In Biological control: a guide to natural enemies in North America. Cornell University. Adapted from D. Kain and J. Nyrop (1995). Predatory mites. Insect Identification Fact Sheet No. 23. Cooperative Extension, Cornell University, Ithaca, N.Y. <http://www.nysaes.cornell.edu/ent/biocontrol/predators/galpyri.html>. Accessed on Sept. 1, 1999.