

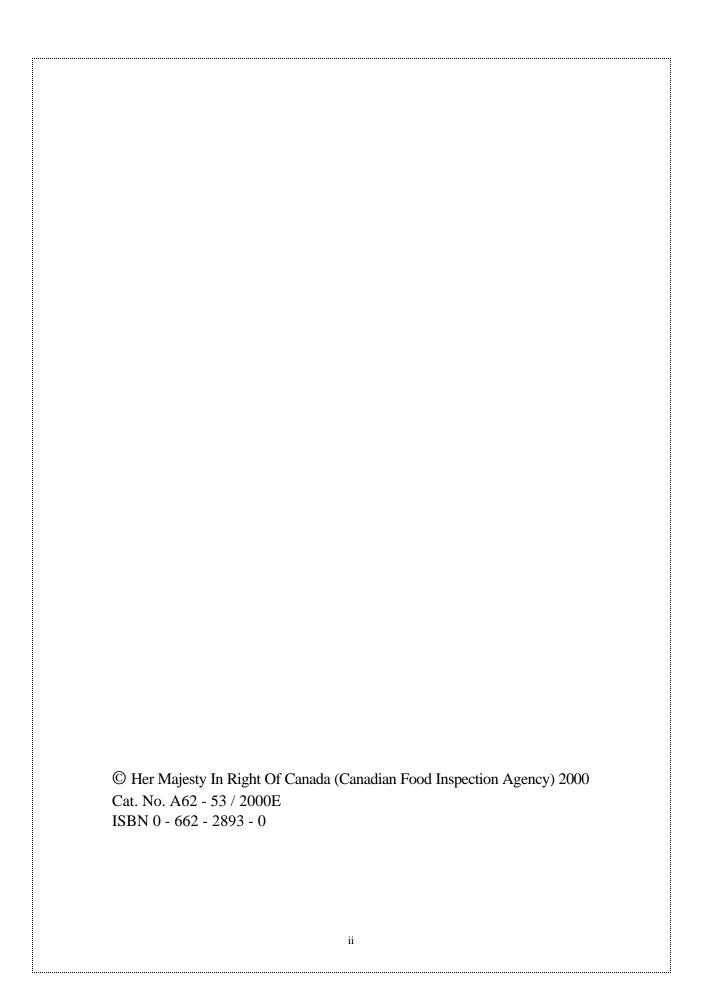
Canadian Microbiological Baseline Survey

of

Chicken Broiler
and
Young Turkey
Carcasses

June 1997 - May 1998





ENQUIRIES

General questions concerning this report should be directed to the CFIA spokespersons:

English:

Dr Gary Thiessen, D.V.M., M.Sc., (**EDITOR**) Acting Chief Poultry Inspection Programs Food of Animal Origin Division, Canadian Food Inspection Agency

telephone	(613) 225-2342 ext. 4689
facsimile	(613) 228-6636
e-mail address	"gthiessen@em.agr.ca"

French:

Ms. Sylvye Des Marchais, Special Advisor to the President and Communications MPIP,

Canadian Food Inspection Agency,

3100 Laframboise Blvd., Saint-Hyacinthe, Quèbec, Canada J2S 4Z4

telephone	(450) 773-6639 ext. 153
facsimile	(450) 774 8522
e-mail address	"desmarchaiss@em.agr.ca"

INTERNET ACCESS

This report and other related information are available from the Internet such as:

- fact sheets on the causes and prevention of foodborne diseases associated with microbial pathogens
- GENERIC MODELS for poultry slaughtering operations, intended for use as a guide by plants developing their HACCP (Hazard Analysis Critical Control Program) system
- the meat hygiene Manual of Procedures (MOP),
- the Meat Inspection Act & Regulations, by contacting the

CFIA website: "www.cfia-acia.agr.ca"

ACKNOWLEDGMENTS

Oversight was provided by the Modernized Poultry Inspection Project (MPIP) Science and Technology Committee comprised of staff from the participating organizations (with their telephone numbers) who supplied the following expertise as listed:

Canadian l	Food Inspection Agency (CFIA);
C	Dr Yves Labbé, former Chief, Poultry Inspection Programs
C	Dr Jean-Robert Bisaillon, veterinary epidemiologist
C	Dr Jean Kamanzi, veterinary microbiologist
C	Dr Gary Thiessen, veterinary poultry specialist
C	Mr Roger Trudel, statistician
Health Pro	tection Branch (HPB) of Health Canada 1-519-822-3300
C	Dr Rebecca Irwin, veterinary microbiology & risk assessment
Canadian l	Poultry and Egg Processors Council (CPEPC) 1-613-724-6605
C	Mr Martin Pelletier, technical director, CPEPC
C	Mr Larry Binning, Cold Springs Farm Limited
C	Dr Keith McMillan, veterinarian, Lilydale Cooperative Ltd.
C	Mr Wayne Sprung, microbiologist, Maple Leaf Poultry
C	Mr Murray Hunt, research & development, Olymel S.E.C./L.P.
Further Po	ultry Processors Association of Canada (FPPAC) 1-613-739-7850
C	Mr Robert de Valk, food industry consultant
Chicken F	armers of Canada (CFC)
C	Mr Mike Dungate, chicken producer specialist
Canadian 7	Furkey Marketing Agency (CTMA) 1-905-564-3100
C	Mr Sateesh Singh, turkey producer specialist

Computer analysis and reports on slaughter volumes by poultry class, by region and by plant were provided by Mr Gary Woito using the CFIA data base.

The project was jointly funded by the CPEPC and by the Agri-Food Trade 2000 Program administered by the Market and Industry Services Branch (MISB) of Agriculture & Agri-Food Canada (AAFC) with Mr Earl New as the project contact person (telephone 613-724-9511, internet address "newe@em.agr.ca") .

Sample collection and/or oversight was provided by on-site CFIA staff at the participating federally registered slaughtering establishments.

Courier services were provided under contract by Purolator Courier Ltd..

Laboratory services were provided under contract by Silliker Laboratories of Canada, Ltd..

Table of contents

SUMMARY 1
INTRODUCTION
OBJECTIVES
Program Design Relative to Objectives
SURVEY DESIGN
Sampling plan
Sample Collection and Handling
Laboratory procedures
Statistical analysis
RESULTS4
Sampling Profile
Salmonella spp5
Escherichia coli (E. coli) Biotype I
Aerobic Plate Count (APC)
CONCLUSIONS
TABLES
FIGURES
REFERENCES

LIS	7
Table 1.	Canadian Poultry Microbiological Baseline Survey, June 1997 -May 1998:
	Chicken broilers, sampling plan and number of carcasses received and suitable for testing
Table 2.	Canadian Poultry Microbiological Baseline Survey, June 1997 -May 19 Young turkeys, sampling plan and number of carcasses received and suitable for testing
Table 3.	Canadian Poultry Microbiological Baseline Survey, June 1997 -May 1999 Chicken broilers and young turkeys, number of sample carcasses requested, not received, not suitable for testing and suitable for laboratory testing
Table 4.	Canadian Poultry Microbiological Baseline Survey, June 1997 -May 1999 Proportion of sampled chicken broiler and young turkey carcasses shown to be positive for Salmonella bacteria upon laboratory testing
Table 5.	Canadian National Microbiological Poultry Baseline Survey, June 1997 -May 1998:
	Summary statistics for specified bacteria counts from samples of chicken broiler and young turkey carcasses
Table 6.	Canadian Poultry Microbiological Baseline Survey, June 1997 -May 1998:
	Distribution of Salmonella (per ml) enumerated in rinse fluids
	from chicken broiler carcasses testing positive for the presence of Salmonella bacteria
Table 6a.	Canadian Poultry Microbiological Baseline Survey, June 1997-May 1999 Distribution of Salmonella (per cm² of surface area) as calculated from rinse fluids from chicken broiler carcasses testing positive for

Table 7.	Canadian Poultry Microbiological Baseline Survey, June 1997-May 1998:
	Salmonella spp. distribution (per ml) enumerated in rinse fluids from young turkey carcasses which tested positive for the presence of Salmonella bacteria
Table 7a.	Canadian Poultry Microbiological Baseline Survey, June 1997-May 1998: Salmonella spp. distribution (per cm²) as calculated from rinse fluids from young turkey carcasses which tested positive for the presence of Salmonella bacteria
Table 8.	Canadian Poultry Microbiological Baseline Survey, June 1997-May 1998: Escherichia coli (Biotype I) distribution (per ml) enumerated in rinse fluids from chicken broiler carcasses
Table 8a.	Canadian Poultry Microbiological Baseline Survey, June 1997-May 1998: Escherichia coli (Biotype I) distribution (per cm²) as calculated from rinse fluids from chicken broiler carcasses
Table 9.	Canadian Poultry Microbiological Baseline Survey, June 1997 -May 1998: Escherichia coli (Biotype I) distribution (per ml) enumerated in rinse fluids from young turkey carcasses
Table 9a.	Canadian Poultry Microbiological Baseline Survey, June 1997 -May 1998: Escherichia coli (Biotype I) distribution (per cm²) as calculated from rinse fluids from young turkey carcasses
Table 10.	Canadian Poultry Microbiological Baseline Survey, June 1997 -May 1998: Distribution of Aerobic Plate Count (APC) @35°C (CFU/ml) enumerated in rinse fluids from chicken broiler carcasses

Table 10a.	Canadian Poultry Microbiological Baseline Survey, June 1997 -May 1998 Distribution of Aerobic Plate Counts (APC) @35°C (CFU/ cm²) as calculated from rinse fluids from chicken broiler carcasses 15
Table 11.	Canadian Poultry Microbiological Baseline Survey, June 1997 -May 1998: Distribution of Aerobic Plate Counts @35°C (CFU/ml) enumerated in rinse fluids from young turkey carcasses
Table 11a.	Canadian Poultry Microbiological Baseline Survey, June 1997 -May 1998: Distribution of Aerobic Plate Counts @35°C (CFU/cm²) as calculated from rinse fluids from young turkey carcasses
FIGURES	
Figure 1.	Distribution of <i>Salmonella</i> Bacteria Enumerated in Chicken Broiler and Young Turkey Carcass Rinse Fluids
Figure 2.	Number of <i>Salmonella</i> Bacteria Estimated on Chicken Broiler and Young Turkey Carcasses
Figure 3.	Seasonal Incidence of <i>Salmonella</i> Bacteria on Chicken Broiler and Young Turkey Carcasses
Figure 4.	Escherichia coli (Biotype I), Distribution (per ml) in Rinse Fluids from Chicken Broiler & Young Turkey Carcasses 21
Figure 5.	Aerobic Plate Counts @35°C, Distribution (per ml) in Rinse Fluids from Chicken Broiler & Young Turkey Carcasses

Canadian Microbiological Baseline Survey of Chicken Broiler and Young Turkey Carcasses June 1997 - May 1998

SUMMARY

A national survey was conducted to evaluate the prevalence and levels of *Salmonella spp.*, *Escherichia coli* (Biotype 1) and Aerobic Plate Counts (APC) on chicken broiler and young turkey carcasses under current processing practices. Carcasses from federally registered establishments across Canada were evaluated using the rinse sampling procedure developed by the United States Department of Agriculture (USDA). Results were expressed as (colony forming units) CFU/ml of rinse fluid which were then calculated as approximate CFU/cm² of carcass surface area. The average percentage of carcasses testing positive for Salmonella (USDA qualitative test) was 21.1% for chicken broilers and 19.6% for young turkeys. The estimated number of Salmonella rinsed from sampled carcasses (using the USDA qualitative MPN method) was less than 100 from 97% of chicken broiler and 98% of young turkey carcasses collected after chilling but prior to cut-up and/or packaging. The 80th percentile (used as "m" in a 3 class sampling plan by the USDA) for results expressed as CFU/ml was 21 *E. coli* for chicken broiler carcasses and 23 for young turkeys. Similarily, the 98th percentile (or "M") for *E. coli* bacteria per ml of rinse fluid was 950 CFU/ml for broiler chicken carcasses and 350 CFU/ml for young turkeys. The geometric mean for APC was 971 and 1,306 CFU/ml of rinse fluid for chicken broiler and young turkey carcasses respectively.

INTRODUCTION

Consumer groups concerned with food safety, poultry growers and processors desiring to adapt to rapidly evolving new technologies and processes, and regulatory agencies worldwide supplementing organoleptic - based inspection methods with requirements to control foodborne pathogenic bacteria all recognize the need to better integrate science into existing inspection programs. The development of HACCP (Hazard Analysis and Critical Control Points) systems for poultry slaughtering operations has provided just such an opportunity. However, the implementation of HACCP has highlighted the need for microbial guidelines or standards to objectively quantify the critical limits or pass/fail criteria for use in HACCP plans as part of the verification activities at applicable CCP's (Critical Control Point) to demonstrate ongoing process control over e.g. evisceration and chilling operations. For example, the USDA conducted a "Nationwide Broiler Chicken Microbiological Baseline Data Collection Program" during 1994/5 and a similar program for young turkeys in 1997/8 to assist them in developing performance based microbiological guidelines or standards for the implementation of HACCP. CFIA and national poultry industry associations made a joint decision in 1996 to conduct a similar national baseline survey of poultry carcasses from federally inspected slaughtering establishments.

OBJECTIVES

- 1. To provide current data on the prevalence and levels of *Salmonella spp.*, *Escherichia coli* (Biotype 1) and Aerobic Plate Counts (APC) on chicken broiler and young turkey carcasses under commercial processing practices.
- 2. To use the information to calculate science based *Escherichia coli* (*E. coli*) guidelines and *Salmonella spp*. standards for incorporation into the MPIP policy and for use by plants developing HACCP systems to
 - C enhance the safety of poultry meat products and
 - facilitate continued trade in poultry products with e.g. the United States (US).

Program Design Relative to Objectives:

The survey was modelled after the Nationwide Broiler Chicken Microbial Baseline Data Collection Program, Working Draft, July 16, 1994, United States Department of Agriculture (USDA). Sample collection and laboratory testing procedures were based on the aforementioned document and the "Pathogen Reduction; Hazard Analysis and Critical Control Point (HACCP) Systems" regulatory amendments as published in the Federal Register (61 FR 38806) on July 25, 1996 (internet address "http://www.fsis.usda.gov/OA/haccp/ imphaccp.htm").

SURVEY DESIGN

Sampling plan:

It was determined that a sample size of 750 chicken broiler and 542 young turkey carcasses would sample the Canadian poultry production at roughly 10 times the ratio as used in the US baseline survey (USDA, 1996). An additional 20% was added to compensate for any samples which could not be submitted (e.g. plant closed) and for carcasses which might arrive at the lab either too warm (>10°C) or too late (more than 24 h after collected).

Within a province, sampling units (carcasses) were allocated proportional to slaughter volume in each establishment. Plants assigned less than one sample were excluded. Within an establishment, a day was randomly selected for collection of each carcass. Based on 1995 slaughter volumes, the survey design ensured that 99.9% of the chicken broilers, and 99.6% of young turkeys slaughtered under federal inspection were eligible to be sampled.

Schedules for a 52 week period were distributed to the Veterinarian-in-Charge (VIC) at each poultry establishment included within the survey.

Sample Collection and Handling:

A carcass with no parts trimmed or missing was randomly selected under the supervision of government inspection staff. Sterile technique was carefully observed during sample collection and packaging. Each carcass was selected after chilling, at the end of the drip line or at the last readily accessible point prior to cut-up or packaging. Samples were double-bagged and shipped in precooled insulated containers with frozen jell pack(s) sufficient to maintain carcass temperature above freezing but below 10°C during transit. Samples were to be processed at the laboratory by the day after collection.

Laboratory procedures:

After determining the temperature and weight, each carcass was shaken for one minute within a sterile bag containing 400 ml of Butter field's diluent for chicken broiler and 600 ml for young turkey carcasses.

The rinse fluid was then sampled according to following bacteriological tests:

- APC incubated for $48 \pm 3 \text{ h}$ @ $35 \pm 1\text{EC}$ using 3M PetrifilmTM (JAOAC, 1990),
- C E. coli count incubated for $48 \pm 4 \text{ h}$ @ $35 \pm 1\text{EC}$ using 3M PetrifilmTM (JAOAC, 1991),
- Salmonella *qualitative* test- official Food Safety and Inspection Service (FSIS),USDA method, and Salmonella *quantitative* test- official FSIS/USDA Most Probable Number (MPN) method, but only on samples positive to the salmonella qualitative test.

Silliker Laboratories of Canada, Ltd. has been accredited by Agriculture and Agri-Food Canada (AAFC) for various microbial tests. To assure conformance to the above listed procedures, one (1) announced and one (1) unannounced audit were performed by representatives of CFIA, Health Canada and CPEPC at the beginning, and during the course of the survey, respectively.

Statistical analysis:

A specific numerical value was required to permit statistical analysis. Therefore samples which had some of the target bacteria present, but in very low quantities (reported as <1), were assigned a numerical value between zero (0) and 1.0 ie 0.1, 0.2, 0.3,....prior to further analysis.

Counts expressed in CFU/ml were used to calculate percentiles required for 3 class sampling plans ie the 80th (equivalent to "m") and the 98th percentile (equivalent to "M"). However APC or E. coli, expressed as CFU/ml,

were transformed into log_{10} prior to calculating other summary statistics such as the median, geometric mean, the standard error & confidence intervals, etc.

Data from carcass rinse fluids, expressed as colony forming units (CFU)/ml, was also converted to counts per cm² by using the formula published in the FSIS/USDA microbial baseline survey (USDA, 1996) as follows:

```
Total colony forming units (CFU) of bacteria / Total Surface Area (cm<sup>2</sup>) = (\# CFU/ml \text{ recovered } x \text{ ml used to rinse the carcass}) / ((0.87 \text{ x w}) + 635)
```

```
The denominator is a formula reported by N. L. Thomas (1978); 

Total\ Carcass\ Surface\ Area\ (cm^2) = 0.87w + 635 where "w" is the weight of the carcass in grams.
```

The geometric mean of the *Salmonella spp.*, *E coli* or APC counts was determined by calculating the antilog of the mean bacteria count log ₁₀.

The Upper and Lower 95% Confidence Limits of the average or mean bacteria count (each count expressed as or transformed into log 10) was calculated using the following formula:

```
mean of the counts transformed into \log_{10} \pm 1.96 \ x (standard deviation of the counts transformed into \log_{10} \pm square root of the number of values).
```

The 95% Confidence Interval (C.I.) of the geometric mean was then obtained by calculating the antilog of the confidence limits as obtained from the preceding formula. The standard error (S.E.) was also contained within the preceding formula since the S.E. is defined as the:

```
(standard deviation of the counts transformed into log 10 ÷ square root of the number of values).
```

All aforementioned summary statistics were obtained by the use of ExcelTM version 7 including the largest and smallest values which were readily obtained by ordering the data set in ascending or descending order. Then the number of

values within each consecutive range, as contained in the tables and figures, were manually calculated by using the associated row numbers in the large Excel TM spreadsheet which contained all the test results which had been entered directly by the laboratory.

RESULTS

Sampling Profile:

Prior slaughter volume and the corresponding number of carcasses requested for both chicken broilers and young turkeys are presented in Tables 1 and 2.

Carcasses were tested from a total of 36 chicken and 14 turkey slaughtering plants. The remaining seven (7) chicken and eight (8) turkey establishments, which were unable to participate in the survey, accounted for only 3.3 % of the chicken and 4.3 % of the requested turkey samples. Among participating plants, the number of low volume plants scheduled for 1-3 samples was 10 for chickens and 6 for turkeys.

From 901 chicken carcasses scheduled for selection as samples, 774 (or 86%) were suitable for testing by the laboratory (Table 1). Similarly, of 651 turkey carcasses included in the national sampling plan, 506 (or 78% of requested samples) qualified for testing (Table 2). The remaining carcasses were in transit too long, or arrived too warm or were not received for testing (Table 3).

Of the tested chicken carcasses, 90% were supplied by 25 chicken slaughtering establishments. Similarly, 90% of the tested turkey carcasses came from 7 turkey slaughtering establishments.

The average weight of the carcasses tested at the laboratory was 1.4 kg for broiler chickens and 5.3 kg for young turkeys.

Salmonella spp:

The average percentage of carcasses testing positive for Salmonella (qualitative test) was 21.1 % for sampled chicken broiler carcasses and 19.6 % for young turkeys (Table 4).

The geometric mean salmonella count from the MPN method was 0.08 CFU/ml of rinse fluid for chicken broiler carcasses and 0.08 CFU/ml for young turkey carcasses (Table 5).

Tables 6-7a and Figure 1, modelled after the USDA-FSIS baseline surveys, present the prevalence or frequency of occurrence of *Salmonella spp*. in the rinse fluids from chicken broiler and young turkey carcasses. Data is presented such that each level or interval in the Tables and Figures encompasses one log cycle.

The estimated number of salmonella bacteria (MPN method) on sampled Canadian poultry carcasses was very low. Less than 0.30 Salmonella cells per ml of rinse fluid were enumerated from 98% of chicken and turkey carcasses (Tables 6 & 7, Figure 1). Similarly, 98% of chicken and turkey carcasses had below 0.30 Salmonella cells per cm² of surface area as sampled by the rinse method (Tables 6a & 7a). Moreover, 61% of chicken carcasses (Tables 6 & 6a, Figure 1) and 68% of turkey carcasses (Tables 7 & 7a, Figure 1) found to be positive by the qualitative method (reported as Salmonella positive) had levels of salmonella below the level of detection as tested by the quantitative MPN method.

Under 100 salmonella CFU per carcass were isolated from 96.9% of chicken and 96.0% of turkey carcasses (Figure 2).

The seasonal incidence of salmonella positive carcasses was relatively constant throughout summer, fall and winter (20-22%), but dropped to 15% in the spring (Figure 3) for young turkey carcasses. A seasonal peak in the summer was observed for chicken broiler carcasses (Figure 3).

Escherichia coli (*E. coli*) Biotype I :

Tables 8 -9a and Figure 4 present the prevalence, or frequency of occurrence, of Escherichia coli

(Biotype 1) in chicken broiler and young turkey carcass rinse fluids such that each level or interval encompasses one log cycle.

The 80th and 98th percentile for E. coli bacteria, (used as a general indicator of hygiene during evisceration operations), was 21 and 950 CFU/ml respectively for chicken broilers. Corresponding counts were 23 and 350 CFU/ml respectively for young turkeys (Table 5).

Aerobic Plate Count (APC):

Tables 10 - 11a and Figure 5 present the prevalence, or frequency of occurrence, of APC in chicken broiler and young turkey carcass rinse fluids such that each level or interval encompasses one log cycle.

The geometric mean per ml of carcass rinse fluid for APC was 971 CFU/ml (or log_{10} 2.99) for chicken broilers and 1,306 CFU/ml (or log_{10} 3.12) for young turkeys (Table 5).

CONCLUSIONS

Canadian results for salmonella incidence and for *E. coli* & APC counts were similar to those reported in the US poultry baseline surveys.

Summary statistics from the national baseline surveys should be considered by Canadian poultry establishments developing or reviewing their plant specific HACCP system.

The national percentage of carcasses positive for *Salmonella spp*.(% +ve), such as provided by this survey, may be used as a reference to measure the effectiveness of any future pathogen reduction initiatives.

TABLES

Table 1. Canadian Poultry Microbiological Baseline Survey, June 1997 -May 1998: Chicken broilers, sampling plan and number of carcasses received and suitable for testing

Region	1995 Slaughter Volume	Proportion of National Slaughter	Number of Samples Requested	Number Suitable for Testing	Proportion of National Samples
Atlantic	39,892,896	8.9 %	80	70	9.0 %
Québec	133,512,917	29.8 %	269	233	30.1 %
Ontario	133,386,898	29.8 %	266	232	30.0 %
Mid-West	27,849,760	6.2 %	56	47	6.1 %
Alberta	43,038,015	9.6 %	86	63	8.1 %
British Columbia	70,444,222	15.7 %	144	129	16.7 %
Total	448,124,708	100.0 %	901	774	100.0 %

Table 2. Canadian Poultry Microbiological Baseline Survey, June 1997 -May 1998:

Young turkeys, sampling plan and number of carcasses received and suitable for testing.

Region	1995 Slaughter Volume	Proportion of National Slaughter	Number of Samples Requested	Number Suitable for Testing	Proportion of National Samples
Atlantic	1,039,113	5.0 %	33	15	2.9 %
Québec	5,359,393	25.9 %	168	138	27.3 %
Ontario	8,089,200	39.1 %	255	188	37.1 %
Mid-West	2,060,367	10.0 %.	65	51	10.1 %
Alberta	1,802,840	8.7 %	57	50	9.9 %
British Columbia	2,347,487	11.3%	73	64	12.7 %
Total	20,698,400	100.0 %	651	506	100.0 %

Table 3. Canadian Poultry Microbiological Baseline Survey, June 1997 -May 1998: Chicken broilers and young turkeys, number of sample carcasses requested, not received, not suitable for testing and suitable for laboratory testing.

Number	Chicken Broilers	Young Turkeys
Requested	901	651
Not Received	32	67
Temperature >10°C	86	68
Transit Time >24 hr	9	10
Suitable for Testing	774	506

Table 4. Canadian Poultry Microbiological Baseline Survey, June 1997 -May 1998: Proportion of sampled chicken broiler and young turkey carcasses shown to be positive for Salmonella bacteria upon laboratory testing.

	Chicken Broilers Young Turkeys	
% Salmonella +ve \pm S.E.	21.1 ± 1.5	19.6 ± 1.8
C. I., % Salmonella +ve	18 -24	16 -23

%+ve -percentage of sampled carcasses testing positive for Salmonella spp.

S.E. -Standard Error using the binomial distribution

C. I. -95% Confidence Interval

(There is a 95% probability that the true (or exact) percentage of chicken broiler carcasses testing positive for Salmonella bacteria from the entire population of Canadian chicken broilers slaughtered in federally registered plants during 1997-98 was somewhere within the range of 18 -24 %)

Table 5. Canadian National Microbiological Poultry Baseline Survey, June 1997 -May 1998: Summary statistics for specified bacteria counts from samples of chicken broiler and young turkey carcasses

		CFU/ml of carcass rinse fluid		CFU/cm ² of carcass surface area	
Bacteria	Summary Statistics	Chicken Broilers	Young Turkeys	Chicken Broilers	Young Turkeys
MPN Salmonella spp. (quantified positive samples only)	Geometric Mean C.I. Geometric Mean Log ₁₀ Mean ± S.E. Median Value Maximum Value	0.08 0.06 -0.12 -1.07 ± 0.07 0.04 >110	0.08 0.06 -0.10 -1.11 ± 0.05 0.09 0.40	0.008 0.004 -0.013 -2.12 ± 0.12 0.009 >0.30	0.006 0.005 -0.008 -2.21 ± 0.06 0.007 0.032
Escherichia. coli Biotype I (E. coli)	Geometric Mean C.I. Geometric Mean Log ₁₀ Mean ± S.E. Median Value Maximum Value m (80th percentile) M (98th percentile) % of counts # 1,000	16.22 14.48 -18.73 1.22 ± 0.03 13 8,000 73 927 98.4	9.33 8.16 -10.63 0.97 ± 0.03 9 3,000 23 350 98.8	3.54 3.12 -4.03 0.55 ± 0.03 3 1,658 16 208 99.9	1.12 0.98 -1.28 0.05 ± 0.03 1 433 3 51 100.0
Aerobic Plate Count (APC) @ 35°C	Geometric Mean C.I. Geometric Mean Log ₁₀ Mean ± S.E. Median Value Maximum Value	971 844 -1,066 2.99 ± 0.02 870 290,000	1,306 1,123 -1,519 3.12 ± 0.03 1,100 520,000	210 192 -231 2.32 ± 0.02 182 57,732	158 135 -185 2.20 ± 0.03 130 34,599

CFU/ml or cm²-colony forming units (estimated number of bacteria) per millilitre of rinse fluid or per square centimetre of surface MPN-most probable number (closely approximates the number of bacteria), quantitative method applied to positive samples only Geometric Mean-antilog of the average of the bacteria counts which had been transformed into log_{10} ; C.I.-95% confidence interval Median Value -50th percentile; 80th percentile-maximum value for 80 percent of the eg *E. coli* counts

Table 6. Canadian Poultry Microbiological Baseline Survey, June 1997 -May 1998: Distribution of Salmonella (per ml) enumerated in rinse fluids from chicken broiler carcasses testing positive for the presence of Salmonella bacteria.

Range MPN CFU/ml	Number of Samples	Percent of Total	Cumulative Number	Cumulative Percent
<0.031	99	60.7	9.9159161e+13	60.7
0.030 -0.30	60	36.8		97.5
0.301 -3.0	2	1.3		98.8
3.01 -30.0	1	0.6		99.4
$>30.0^{2}$	1	0.6		100.0
Total	163	100.0		

MPN-Most Probable Number (closely approximates the number of bacteria)

CFU/ml -colony forming units (estimated number of bacteria) per millilitre of carcass rinse

fluid

Table 6a. Canadian Poultry Microbiological Baseline Survey, June 1997-May 1998: Distribution of Salmonella (per cm² of surface area) as calculated from rinse fluids from chicken broiler carcasses testing positive for the presence of Salmonella bacteria.

Range MPN CFU/cm ²	Number of Samples	Percent of Total	Cumulative Number	Cumulative Percent
<0.011	99	60.7		60.7
0.01 -0.30	50	30.7	9.9149160e+13	91.4
0.0301 -0.30	11	6.7		98.1
0.3001 -3.0	2	1.3		99.4
$3.001 - 30.0^2$	1	0.6		100.0
Total	163	100		

MPN-Most Probable Number (closely approximates the number of bacteria)

CFU/cm² -colony forming units (estimated number of bacteria) per square centimetre of carcass surface area

¹ Negative by quantitative MPN method

² Actual maximum level reported was A>110" MPN/ml which was evaluated as 110 CFU/ml

¹ Negative by quantitative MPN method

² Actual maximum level reported was A>110" MPN/ml which was evaluated as 110 CFU/ml

Table 7. Canadian Poultry Microbiological Baseline Survey, June 1997-May 1998: Salmonella spp. distribution (per ml) enumerated in rinse fluids from young turkey carcasses which tested positive for the presence of Salmonella bacteria.

Range MPN CFU/ml	Number of Samples	Percent of Total	Cumulative Number	Cumulative Percent
<0.031	67	67.7	679799	67.7
0.030 -0.30	30	30.3		98.0
0.301 -3.0	2	2.0		100.0
3.01 -30.0	0	0.0		
>30.02	0	0.0		
Total	99	100		

MPN-Most Probable Number (closely approximates the number of bacteria)

CFU/ml -colony forming units (estimated number of bacteria) per millilitre of carcass rinse

fluid

Table 7a. Canadian Poultry Microbiological Baseline Survey, June 1997-May 1998: Salmonella spp. distribution (per cm²) as calculated from rinse fluids from young turkey carcasses which tested positive for the presence of Salmonella bacteria.

Range MPN CFU/cm ²	Number of Samples	Percent of Total	Cumulative Number	Cumulative Percent
<0.011	67	67.7	67	67.7
0.01 -0.03	31	30.3	97	98.0
0.0301 -0.30	1	2.0	99	100.0
0.3001 -3.0	0	0.0		
3.001 -30.0	0	0.0		
>30.0	0	0.0		
Total	99	100.0		

MPN-Most Probable Number (closely approximates the number of bacteria)

CFU/ml -colony forming units (estimated number of bacteria) per square centimetre of carcass surface area

¹ Negative by quantitative MPN method

² Actual maximum level reported was **A**>110" MPN/ml

¹ Negative by quantitative MPN method

Table 8. Canadian Poultry Microbiological Baseline Survey, June 1997-May 1998: Escherichia coli (Biotype I) distribution (per ml) enumerated in rinse fluids from chicken broiler carcasses.

Range CFU/ml	Number of Samples	Percent of Total	Cumulative Number	Cumulative Percent
<11	31	4.1	31	4.1
1 -10	306	39.5	337	43.6
11 -100	316	40.8	653	84.4
101 -1,000	110	14.2	763	98.6
1,001 -10,000	11	1.4	774	100.0
10,001 -100,000	0	0.0		
100,001 -1,000,000	0	0.0		
Total	774	100.0		

fluid

Table 8a. Canadian Poultry Microbiological Baseline Survey, June 1997-May 1998: Escherichia coli (Biotype I) distribution (per cm²) as calculated from rinse fluids from chicken broiler carcasses.

Range CFU/cm ²	Number of Samples	Percent of Total	Cumulative Number	Cumulative Percent
<0.011	31	4.0		4.0
0.01-1	137	17.7	3.1168579e+16	21.7
1.01 -10	410	53.0		74.7
10.01 -100	153	19.8		94.5
100.01 -1,000	42	5.4		99.9
1,000.01 -10,000	1	0.1		100.0
10,000.01 -100,000	0	0.0		
Total	774	100		

CFU/cm² - colony forming units (estimated number of bacteria) per square centimetre of carcass surface area

¹ Negative by test method

¹ Negative by test method

Table 9. Canadian Poultry Microbiological Baseline Survey, June 1997 -May 1998: Escherichia coli (Biotype I) distribution (per ml) enumerated in rinse fluids from young turkey carcasses

Range CFU/ml	Number of Samples	Percent of Total	Cumulative Number	Cumulative Percent
<11	26	5.1	26	5.1
1 -10	271	53.6	297	58.7
11 -100	175	34.6	472	93.3
101 -1,000	28	5. 5	500	98.8
1,001 -10,000	6	1.2	506	100.0
10,001 -100,000	0	0.0		
100,001 -1,000,000	0	0.0		
Total	506	100.0		

fluid

Table 9a. Canadian Poultry Microbiological Baseline Survey, June 1997 -May 1998: Escherichia coli (Biotype I) distribution (per cm²) as calculated from rinse fluids from young turkey carcasses

Range CFU/cm ²	Number of Samples	Percent of Total	Cumulative Number	Cumulative Percent
<0.011	0	0.0	0	0.0
0.01 -1	255	50.4	255	50.4
1.01 -10	205	40.5	460	90.0
10.01 -100	41	8.1	501	99.0
100.01 -1,000	5	1.0	506	100.0
1,000.01 -10,000	0	0.0		
10,000.01 -100,000	0	0.0		
Total	506	100		

CFU/cm² -colony forming units (estimated number of bacteria) per square centimetre of carcass surface area

¹ Negative by test method

¹ Negative by test method

Table 10. Canadian Poultry Microbiological Baseline Survey, June 1997 -May 1998: Distribution of Aerobic Plate Count (APC) @35°C (CFU/ml) enumerated in rinse fluids from chicken broiler carcasses.

Range CFU/ml	Number of Samples	Percent of Total	Cumulative Number	Cumulative Percent
<11	0	0.0	0	0.0
1 -10	0	0.0	0	0.0
11 -100	21	2.7	21	2.7
101 -1,000	413	53.4	434	56.1
1,001 -10,000	307	39.7	741	95.8
10,001 -100,000	29	3.7	770	99.5
100,001 -1,000,000	4	0.5	774	100.0
Total	774	100		

fluid

¹ Negative by test method

Table 10a. Canadian Poultry Microbiological Baseline Survey, June 1997 -May 1998: Distribution of Aerobic Plate Counts (APC) @35°C (CFU/ cm²) as calculated from rinse fluids from chicken broiler carcasses.

Range CFU/cm ²	Number of Samples	Percent of Total	Cumulative Number	Cumulative Percent
<11	0	0.0	0	0.0
1 -10	0	0.0	0	0.0
10.01 -100	225	29.1	225	29.1
100.01 -1,000	460	59.4	685	88.5
1,000.01 -10,000	84	10.9	769	99.4
10,000.01 -100,000	5	0.6	774	100.0
100,000.01 -1,000,000	0	0.0		
Total	774	100		

CFU/cm² -colony forming units (estimated number of bacteria) per square centimetre of carcass surface area

¹ Negative by test method

Table 11. Canadian Poultry Microbiological Baseline Survey, June 1997 -May 1998: Distribution of Aerobic Plate Counts @35°C (CFU/ml) enumerated in rinse fluids from young turkey carcasses

Range CFU/ml	Number of Samples	Percent of Total	Cumulative Number	Cumulative Percent
<11	0	0.0	0	0.0
1 -10	0	0.0	0	0.0
11 -100	25	4.9	25	4.9
101 -1,000	214	42.3	239	47.2
1,001 -10,000	168	33.2	407	80.4
10,001 -100,000	90	17.8	497	98.2
100,001 -1,000,000	9	1.8	506	100.0
Total	506	100.0		

¹ Negative by test method

Table 11a. Canadian Poultry Microbiological Baseline Survey, June 1997 -May 1998: Distribution of Aerobic Plate Counts @35°C (CFU/cm²) as calculated from rinse fluids from young turkey carcasses

fluid

Range CFU/cm ²	Number of Samples	Percent of Total	Cumulative Number	Cumulative Percent
<11	1	0.2	1	0.2
1 -10	17	3.3	18	3.5
10.01 -100	206	40.7	224	44.2
100.01 -1,000	205	40.5	429	84.7
1,000.01 -10,000	57	11.3	486	96.0
10,000.01 -100,000	20	4.0	506	100.0
100,000.01 -1,000,000	0	0.0		
Total	506	100.0		

CFU/cm² -colony forming units (estimated number of bacteria) per square centimetre of carcass surface area

¹ Negative by test method

FIGURES

Figure 1. Distribution of Salmonella bacteria
Enumerated in Chicken & Turkey Carcass Rinse Fluids

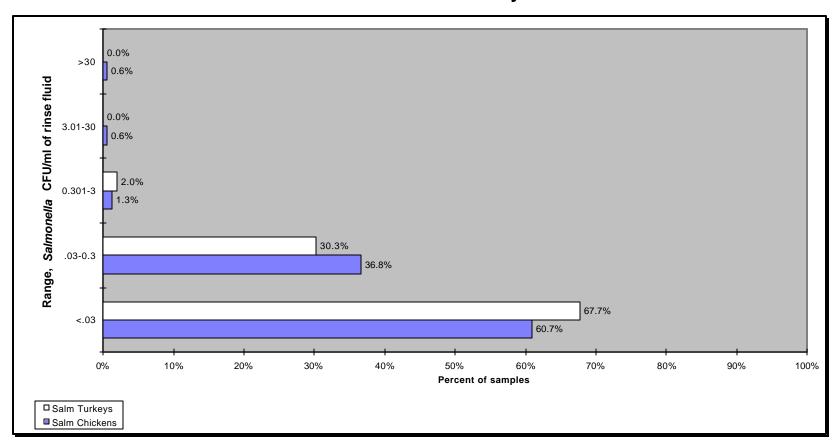


Figure 2. Distribution of Salmonella bacteria;
Number estimated on chicken & turkey carcasses

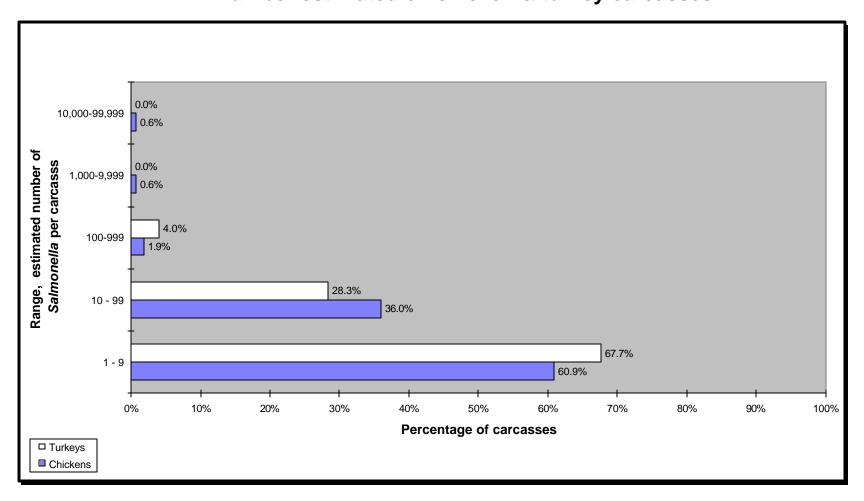


Figure 3. Seasonal Incidence of Salmonella sp. on Canadian Chicken Broiler and Young Turkey Carcasses

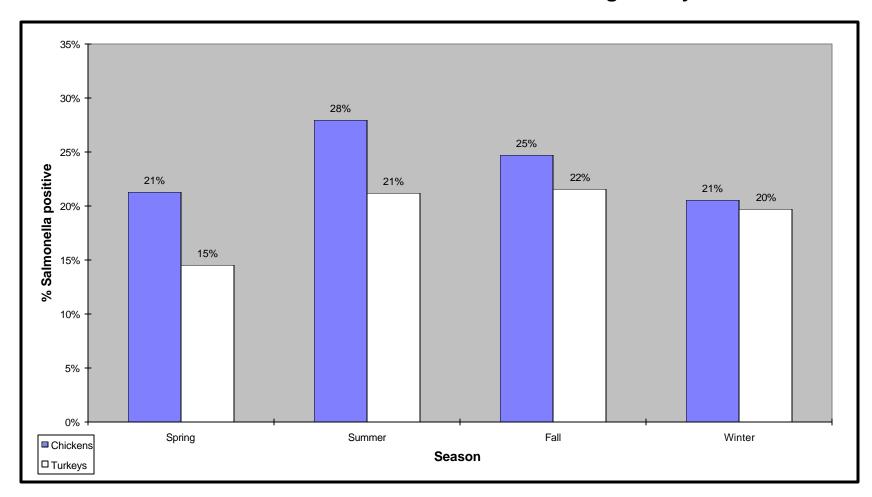


Figure 4. Escherichia coli (Biotype I)
Distribution in Chicken and Turkey Carcass Rinse Fluids

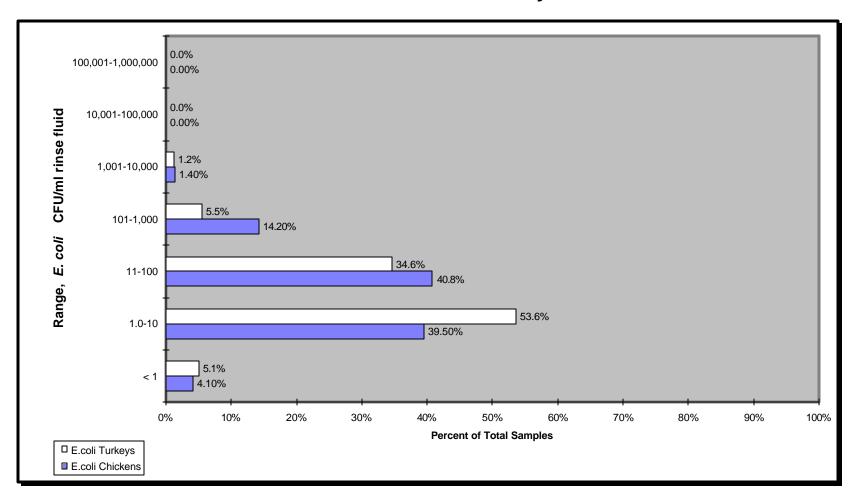
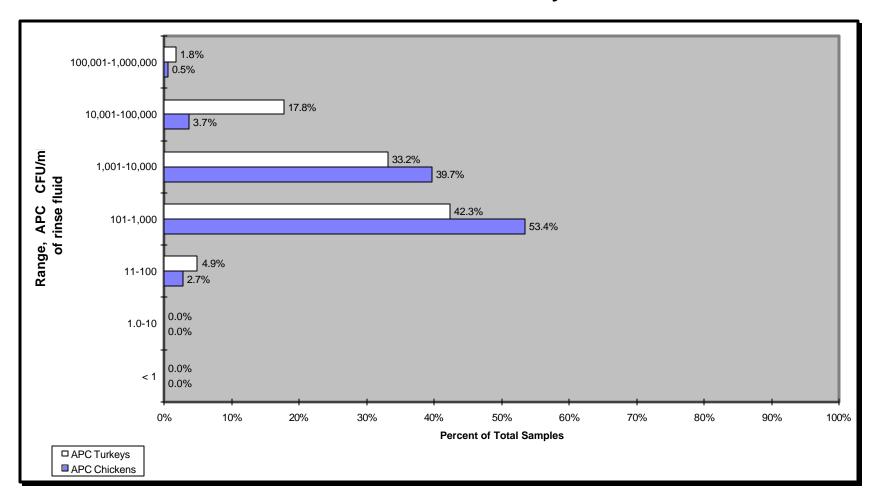


Figure 5. Aerobic Plate Count grown at 35° Centigrade
Distribution in Chicken and Turkey Carcass Rinse Fluids



REFERENCES

Bean, N. H., J. S. Goulding, M. T. Daniels and F. J. Angulo. 1997. Review: Surveillance for food borne disease outbreaks - United States, 1988-1992. J. Food Prot. 60:1265-1286.

Bryan, F. L. 1980. Food borne diseases in the United States associated with meat and poultry. J. Food Prot. 43:140-150.

Curiale, M. S., T. Sons, J. S. McAllister, B. Halsey and T. L. Fox. 1990. Dry rehydratable film for enumeration of total aerobic bacteria in foods; Collaborative study. J. Assoc. Off. Anal. Chem. 73:242-248.

Curiale, M. S., T. Sons, D. McIver, J. S. McAllister, B. Halsey, D. Roblee and T. L. Fox. 1991. Dry rehydratable film for enumeration of total coliforms and *Escherichia coli* in foods; Collaborative study. J. Assoc. Off. Anal.Chem. 74:635-648.

Lammerding, A. M., M. M. Garcia, E.D. Mann, Y. Robinson, W. J. Dorward, R.B. Truscott and F. Tittiger. 1988. Prevalence of *Salmonella spp*. and thermophilic *Campylobacter* in fresh pork, beef, veal and poultry in Canada. J. Food Prot. 51:47-52

Thomas, N. L..1978. Observations of the relationship between the surface area and weight of eviscerated carcasses of chickens, ducks and turkeys. J. Food Technol. 13:81-86.

Todd, E. C. D.. 1992. Food borne disease in Canada - a 10 year summary from 1975 to 1984. J. Food Prot. 55:123-132.

USDA-FSIS. 1994. Concept paper, Nationwide broiler chicken microbiological baseline data collection program. United States Department of Agriculture, Food Safety and Inspection Service, Microbiology Division, Washington, D.C.

USDA-FSIS. 1996. Nationwide young turkey microbiological baseline data collection program. United States Department of Agriculture, Food Safety and Inspection Service, Microbiology Division, Washington, D.C.

USDA-FSIS. 1996. Nationwide broiler chicken microbiological baseline data collection program, July 1994 - June 1995. United States Department of Agriculture, Food Safety and Inspection Service, Science and Technology, Microbiology Division, Washington, D.C.

USDA-FSIS. 1996. Pathogen reduction; hazard analysis and critical control point (HACCP) systems; final rule. Fed. Regist. 61:38806.