How To Submit Samples To The Crop Protection Laboratory

The accuracy of our laboratory's diagnosis of your sample is dependent on the quality of the sample submitted and the inclusion of relevant information. Below are some tips on how to sample, package and submit specimens, as well as information on how to fill out the submission form.

Weed Samples

Sampling

- Choose several samples showing as many growth stages as possible.
- Collect whole samples including roots where possible.
- Ideally, flowering and fruiting stages should be represented as the most distinctive morphological features are seen at these stages.
- Record the habitat, plant features (height, flower colour, odor etc.), abundance (single plant to common species), growth habit (erect, creeping etc.), nature (annual, biennial, perennial), and any factor that might be significant.

Preserving and Packaging

- Plants can be placed between dry absorbent paper (e.g. newspaper) on a flat surface and pressed by adding weight on top of the plants in the paper. Pressed plants should be checked for moisture and replace the paper if wet. More frequent inspection is required in the early stages of pressing. Once the plants are dry they should be placed in rigid containers that do not allow the specimens to be damaged in shipping. Placing dried specimens between rigid pieces of cardboard is effective.
- Plants requiring urgent identification may be shipped immediately, without pressing, loosely wrapped in dry paper towel to prevent damage during shipping. A rigid cardboard container containing packing materials to prevent movement in shipping is appropriate.

<u>Note</u>: although the above conditions are ideal we will attempt to work with plant specimens at any growth stage and in less than ideal conditions. On occasion a lone seed may be enough to identify a plant to the species level. However some identifications will be more difficult and a positive identification may not be possible without all growth stages being represented.

Insect Samples

Sampling

- The sample to be submitted should include several specimens showing as many life stages as possible. The larva or nymphs and adults are the most useful for identification and rarely will eggs or pupae be of much value for specific identifications. Choose as many stages as possible of the larvae or nymph.
- Include information associated with the specimen the habitat (animal pest, plant pest, substrate), location (house, yard, field etc.), type of injury caused to host, numbers (few to numerous), and distribution (scattered, common throughout etc.).

Preserving and Packaging

• Soft-bodied stages of insects should be preserved in 70 per cent alcohol or maintained in substrate (medium in which the insects are living). Alcohol may be obtained in most drug stores and larger quantities are available from lab supply companies.

- Place insects in well-sealed vials or jars (plastic) and then place these in rigid containers with packing materials to prevent breakage during shipping.
- If the insects are to be maintained in their substrate, the insect and substrate should be placed in a large plastic bag that will maintain the humidity of the sample and allow some free air.
- If the substrate is soil or of a loose abrasive structure, the insects should be removed and placed in alcohol. If this is not convenient, then the substrate should be tightly enclosed in plastic to avoid injury to the insects in transit.
- Hard bodied insects (i.e. most beetles) may be placed in a rigid container (vial, pill bottle, jar, etc. and lightly supported with tissue paper to avoid damage during shipping.
- Insects with easily removed scales such as butterflies and moths should ideally be pinned to a small piece of material such as Styrofoam and packed in a rigid position to avoid injury in transit.
- When shipping in live insects, provide the appropriate host plant for nourishment.
- Most insects may be killed by placing them in a freezer overnight this can facilitate handling.

Plant Disease Samples

Sampling

- Choose specimens showing various stages of disease symptoms (light to severe symptom). Include some healthy specimens for comparison.
- If possible, whole plants, including roots, should be submitted (many symptoms seen in above ground plant parts may result from root infection).
- Record the plant parts affected, symptoms observed, distribution within the plant population, and cropping history.
- If there is any possibility of herbicide being implicated, please provide information on chemical use both in the present cropping season and for the previous four years if available.
- It is important to note the fertilizer regimen, including timing of application, rate of application, and the form of the fertilizer used. The previous year's fertilizer applications should also be noted.

Packaging and handling

- Wrap specimens in dry paper towel.
- If the sample includes the root system with soil attached, put this portion into a plastic bag and tie it off at the base of the stem. Leave the above ground parts loosely packed in dry paper towel.
- Submit the sample in a rigid container, loosely packed in dry packing material such as newspaper.
- Note: if a viral infection is suspected, please submit living plants when possible, e.g. dig up infected plants and place in a container and transport to the lab immediately. If this is not possible wrap the detached plant material that is showing symptoms in slightly moistened paper towel and submit to the lab as quickly as possible.

Herbicide Resistance Testing

Sampling

- If possible, collect only mature, healthy seed from the suspect plants. Green or diseased seed will not germinate properly and will impede the diagnosis or make testing impossible.
- A minimum of a thousand seeds per herbicide group is desirable. When there is a need for several groups of herbicides to be tested, number of seeds should be increased accordingly.
- Collect seed from specimens that appear to have survived the herbicide application. If other labeled weeds have been controlled in the area, there is stronger reason to suspect resistance.

- Note the present crop and crops from the previous four years, the herbicide used and its rate of application, the timing of application (month, year, growth stage of crop, growth stage of weed), climatic conditions around the time of application, the weed's distribution and level of control.
- Do not submit seed that has been treated with a preharvest herbicide.
- Indicate the chemical group(s) that you want to be tested.
- Samples are processed on a first-come first-serve basis with the duration of the test varying with the group and type of weed being tested.

Packaging and handling

- ALLOW SEEDS TO AIR DRY BEFORE SHIPPING! This is done by placing the seeds in paper bags and allowing them to air dry for a few weeks in a dry environment. This prevents mold developing in shipment. Note: It is not critical to get the samples to the lab quickly, as testing will not commence until January because the seeds must have time to break their dormancy period.
- Seed should be relatively clean with as little foreign material as possible.
- Once the seeds are dry, they should be packaged in any durable container, including strong plastic bags and then placed in sturdy cardboard containers (or equivalent) prior to shipping to the lab.

NOTES (For All Samples Submitted)

- Never add water to the samples or the packaging as this will result in rotting or saprophytic growth. (*The only exception is for disease samples where viral causal agents are suspected. For specimens of this nature, the sample needs to be maintained in as fresh a state as possible. For above ground parts showing symptoms, wrap individual parts between lightly moistened paper towels and ship to the lab as quickly as possible).*
- If possible, avoid sending in fresh specimens over the weekend to prevent decay in shipping.
- A sample cannot be too large, but it can be too small.
- The information about the sample may be as significant as the sample itself, so provide as much information as possible.
- When possible, use a Crop Protection Diagnostic Form available on this web site.

The Crop Protection Diagnostic Form

This form should accompany all samples. If the form is not available, please write down the information requested above and submit this data with the sample. The form will simplify the recording of information. The form is divided into six sections and should be filled out as follows:

Section 1

- Fill out completely for any sample submitted.
- The check box at the bottom is for confidentiality.

Section 2

- Section 2 need only be filled in if the grower and submitter are different.
- Leave the date received blank, as the lab will fill this out upon receipt of the sample.
- If you would like to receive control measures for the problem, indicate by using this check box.

Fees area

- If unsure whether a disease sample should be visual or cultural, you may attach a note to process as required.
- Please leave the total charges and explanation area blank.

Section 3

- The type of agriculture should be checked off for all samples.
- The other relevant information area should be used to indicate any additional information that may be useful for the diagnosis.
- Fertilizer, timing, and weather are significant for disease and chemical injury samples.

Section 4

• Fill out this section **ONLY** if the sample submitted is for weed identification.

Section 5

• Fill out this section **ONLY** if the sample being submitted is for insect or disease identification.

Section 6

• Fill out this section **ONLY** if the sample being submitted is for herbicide resistance testing, or for visual analysis where chemical injury may be an issue.

Section 7

• Fill out this section **ONLY** if the sample is for herbicide resistance testing.

The Diagnosis and Recommended Treatment area is to be left blank, as this area is used for reply/analysis.

NOTES:

- If there is insufficient space on the form for all the pertinent information, attach a separate piece of paper with these additional details.
- A hand-drawn map showing the distribution or the area of concern may be attached with the form; this is particularly significant where <u>herbicide</u> drift is suspected.
- The length of time it takes to make a diagnosis of specimens submitted depends on the nature of the submission. For example, weed or insect identifications are based on morphological features and are usually complete within one day of being received. If the causal agent for a disease submission is not visually determinable, culturing is required and typically takes about a week to complete, but can take longer depending on the nature of the pathogen.

Call us at 306-787-8130 if there are any questions. It is better to inquire before the submission rather than risk impairing the value of the sample.

Saskatchewan Agriculture, Food	Cr	Crop Protection Diagnostic Form Send samples to:				Inquiry Number			
Revitalization	Ser					Foog (CST) will be added)			
	Cro	op Prote	ction Laboratory		Check appropriate box(es)				
	308	35 Alber	t Street						
	Reg	egina, SK S4S 0B1 (306) 787-8130			Plant Identification				
					\Box Disease ID (Visual)				
					La Disease ID (Cultured) \$40				
1. Date Submitted			2. Date Received		Herbicide Resistance				
Address		'	Address		Group 3 - Green Foxtail \dots \$55				
Town/City		· ,	Town/City		Group 1	- Wild Oats	\$100		
Postal Code		Postal Code			Any Other				
Telephone		Telephone			Total Charges				
Send Reply to Owner/G	rower ONI	LY	Require control meas	ures?	Explanation				
3. General Information		To be co	mpleted for disease iden	ntification or herb	icide tolerance	testing applica	ations.		
Plasse check one		Fertilizer (Date and Rate) Previous Year							
Commercial Agricult	uro	When w	ere symptoms first notic	ed?					
Greenhouse	uie	Weather conditions prior to appearance of symptoms:							
Home/Garden/Yard		Other relevant information:							
U Other (Describe)									
4. Weed/Plant Identificat	tion								
Habitat		Featu	ires	Abundance	Habit	Nature	Specific Information		
Annual Crop/Fallow	Plant Hei	ight	Plant Colour	1 Plant	🗌 Herb	Annual 🗌			
Forage/Rangeland	Plant Tex	ture	Plant Odour	Few	🗌 Shrub	Biennial			
Park/Garden/Yard	Type of I	Root	Flower Odour	Many	Tree	Perennial			
☐ Body of Water	Flower C	Colour	Fruit Size		Erect				
Non-crop Land	our	Fruit Type		Creepir	ng				
5. Insect and Disease Ide	ntification	(Section	six must also be comple	eted for disease di	agnosis or her	bicide resistanc	e testing.)		
Host Name \rightarrow Plant		/	Animal	_ Food/Fabric		_ Other (Spec	uity)		
Parts affected	propriate	ouxes	vmntoms		Distribution		Cronning History		
	/Fruit	Wilting		ion Cro		Scattered	Veor		
Stoms/Branchas		Stunting		es Most of Field Vear					
		Juning				Field Marging	Veer		
		Chawad Laguage Apparent Crowth				Detebu	Veer		
L 110wers		Cheweur				1 ateriy			
6. Pesticide Application	Made to F	ield (Pres	eeding, in crop, pre and	post harvest)	7. Herbicid	e Resistance T	esting		
Year Crop Product Rate of Product Application Date						Due to the dormancy characteristics of wild oat			
	Mo Yr					and other weed seeds, the test results may not be			
(Present)					available for four or more months from the time		nonths from the time		
— ———					the sample i	s received. \checkmark			
•									
(Previous)					Weed				
▼					Herbicide group to be tested for				
					Crop variety				
					Date seeded	l			
▼									
					Herbicide A	Application Inf	formation V		
					Date applied				
					Rate of app	lication			
					Temperature: at application				
weeds Data (Most common weeds present)	Stage	at Application Control (Excellent, Good, Fair, Poor			1 Week Bef	ore 1	Week After		
1)	1)		1)						
2)	2)		2)						
3)	3)		3)						
4)	_ 4)		4)						
By providing information on	this form, yo	u are givin	g Saskatchewan Agriculture,	Food and Rural Re	vitalization conser	nt to use your info	ormation for program		
delivery, development and/or	evaluation p	urposes. It	may also be used to provide	e you with informatio	on on additional p	programs from wh	ich you may benefit.		
Diagnosis/Identifica	tion and	Recom	mended Treatment						
Scientific Name Common Name									