

Ecotoxicology of Nanoparticles: Issues and Approaches

Presented by

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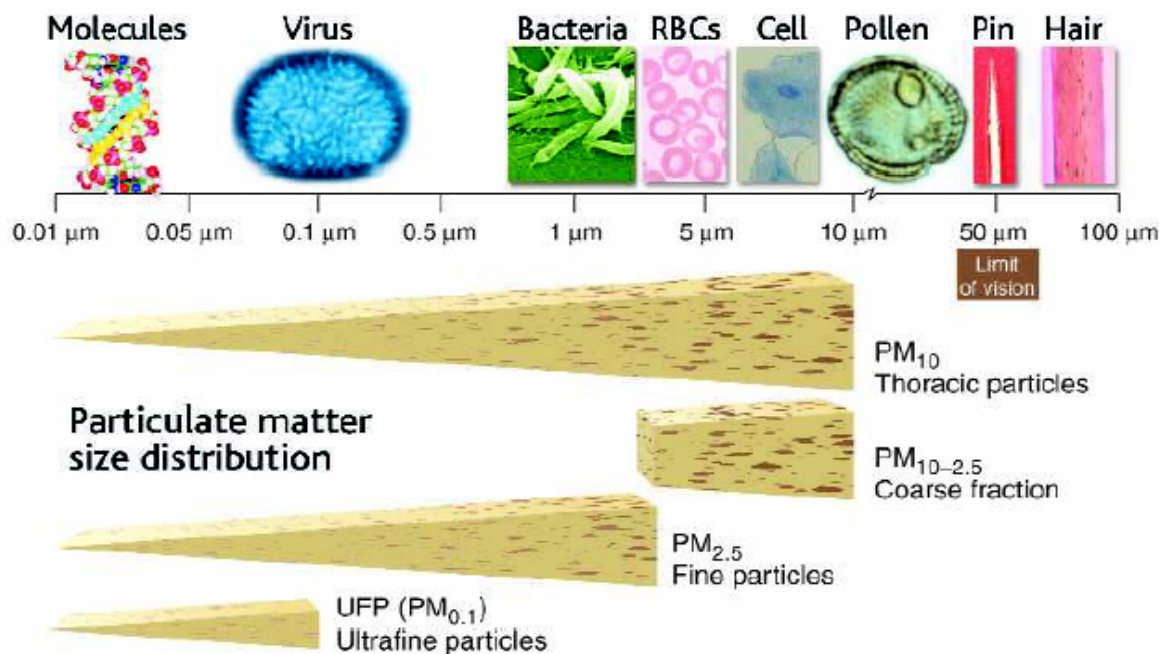
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Outline of presentation

- **Nanoparticles and the Environment**
- **Ecotoxicological Risk Assessment:**
 - **Approaches and measurement tools**
 - **Use of standardized toxicity test methods**
- **Ecotoxicology of NPs using single species toxicity tests**
- **Bioaccumulation of single NPs and mixtures**
- **Issues and Priorities**
- **Recommendations and Closing remarks**

Nanotechnology and Nano(eco)toxicology

- Nanotechnology uses matter sized “*at dimensions of roughly 1 to 100 nanometers.*” (US- National Nanotechnology Initiative)



- Nano(eco)toxicology: Hazardous effects of Engineered NPs on ecological receptors
- Bioavailable NPs
- Nanotoxicity \propto Size, shape, surface area, etc.

Sources of exposure to ultrafine particles and nanoparticles

Table 1. UFPs/NPs (< 100 nm), natural and anthropogenic sources.

Natural	Anthropogenic	
	Unintentional	Intentional (NPs)
Gas-to-particle conversions	Internal combustion engines	Controlled size and shape, designed for functionality Metals, semiconductors, metal oxides, carbon, polymers Nanospheres, -wires, -needles, -tubes, -shells, -rings, -platelets Untreated, coated (nanotechnology applied to many products: cosmetics, medical, fabrics, electronics, optics, displays, etc.)
Forest fires	Power plants	
Volcanoes (hot lava)	Incinerators	
Viruses	Jet engines	
Biogenic magnetite: magnetotactic bacteria, protists, mollusks, arthropods, fish, birds	Metal fumes (smelting, welding, etc.)	
human brain, meteorite (?)	Polymer fumes	
Ferritin (12.5 nm)	Other fumes	
Microparticles (< 100 nm; activated cells)	Heated surfaces	
	Frying, broiling, grilling	
	Electric motors	

- Oberdörster et al. (2005)

**** Emerging area of toxicology and ecotoxicology ****

Nanotechnology applications in the environmental sector

- **New production methods that are more environmentally friendly and consume less resources**
- **New, environmentally friendly materials**
- **More efficient methods for the production and consumption of energy**
- **New methods for the preparation of water, soil decontamination, and new environmental sensors.**

Nanotechnology and Nano(eco)toxicology

- Need quantitative data on ecotoxicology of NPs (dose-response relationships).
- Ecotoxicological risk assessments should account for:
 - toxic effects of the NP on an ecological receptor
 - the probability of exposure of that NP to ecological receptor,
 - and possible release of NP into the environment.

Ecotox Risk \propto Toxicity x Exposure
- Need for standardized methods

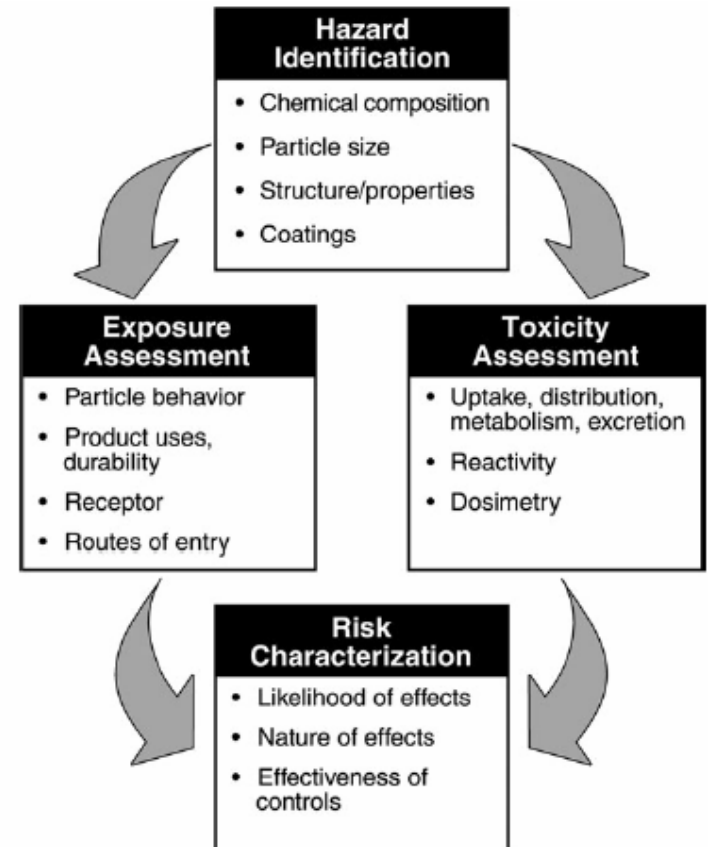
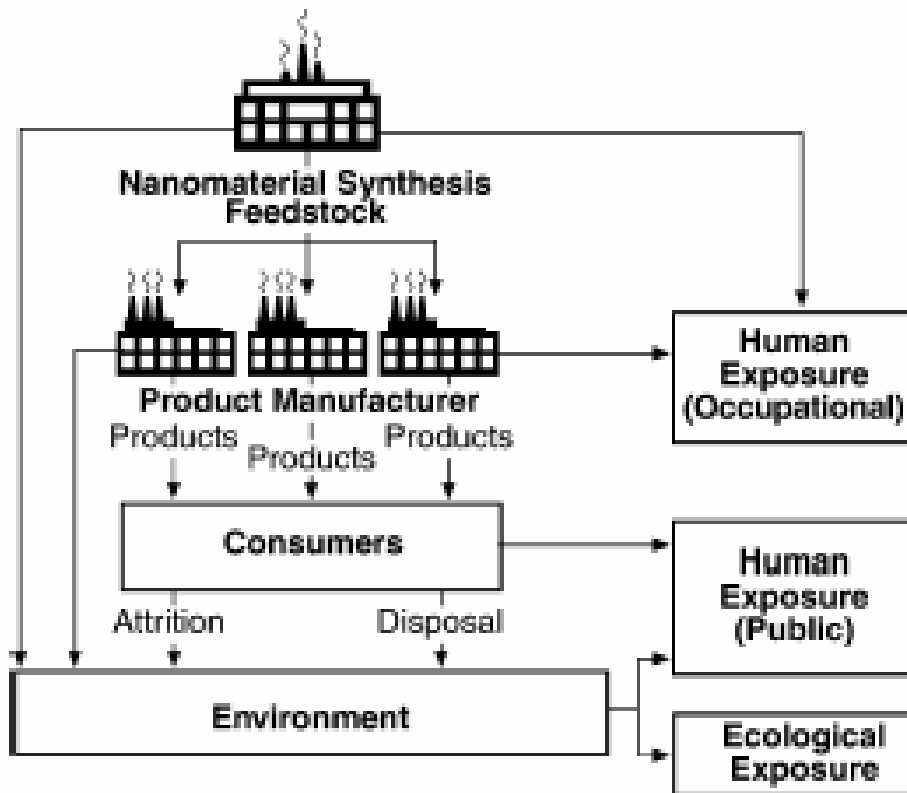


FIG. 4. Risk assessment framework for nanomaterials.

Nano(eco)toxicology and Life Cycle Impact Assessment



LCIA Environmental concerns (e.g., ISO 14042):

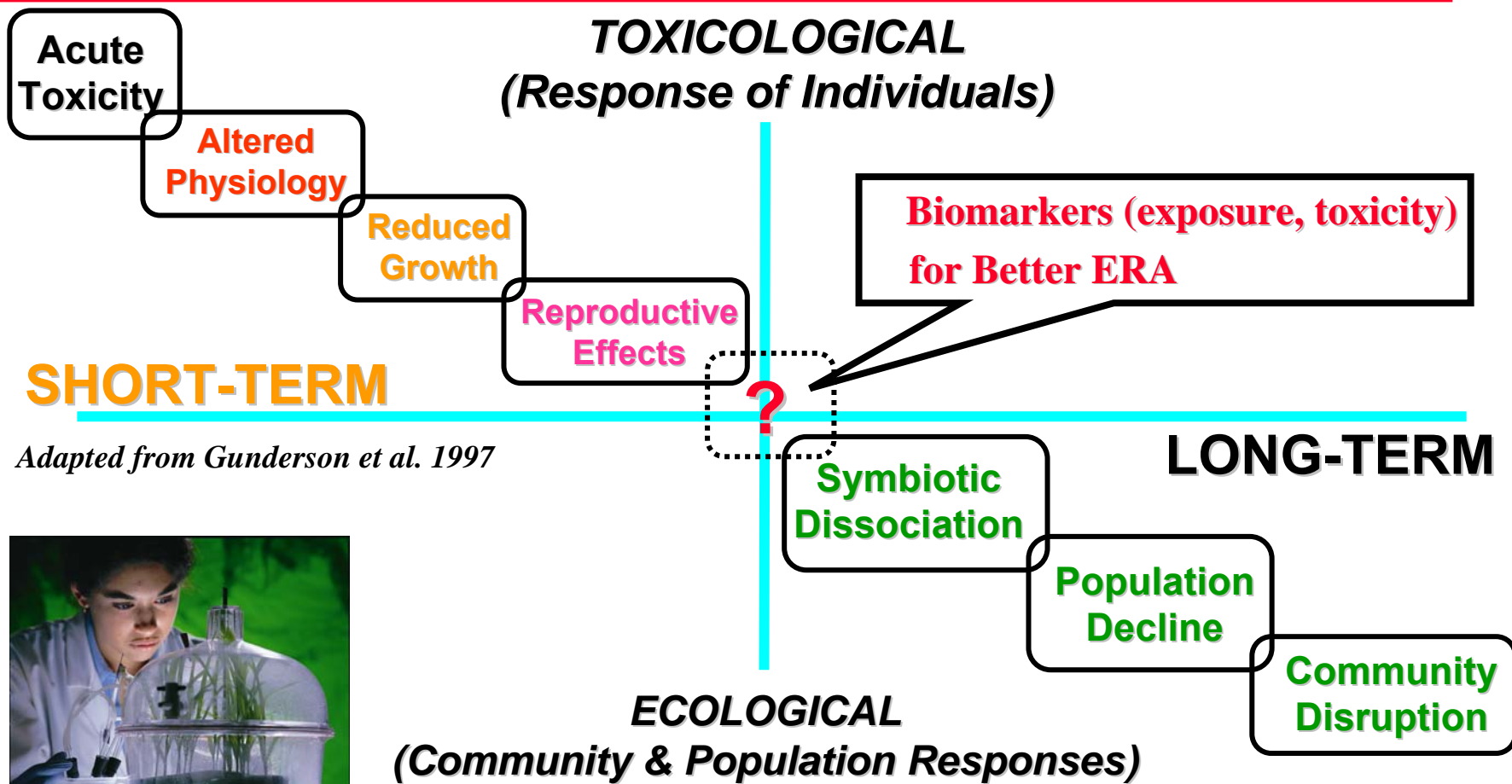
- *Direct Effects*: Toxic effects on ecological receptors.
- *Indirect Effects*: Bioaccumulation and biomagnification of NPs in the food chain. Other impacts (eutrophication, GHG).
- Contamination of water and soil from release and improper disposal of NPs and related products

Water, Soil, and Air

FIG. 5. Potential for release and exposure to nanoscale substances.

Tsuji et al. 2005

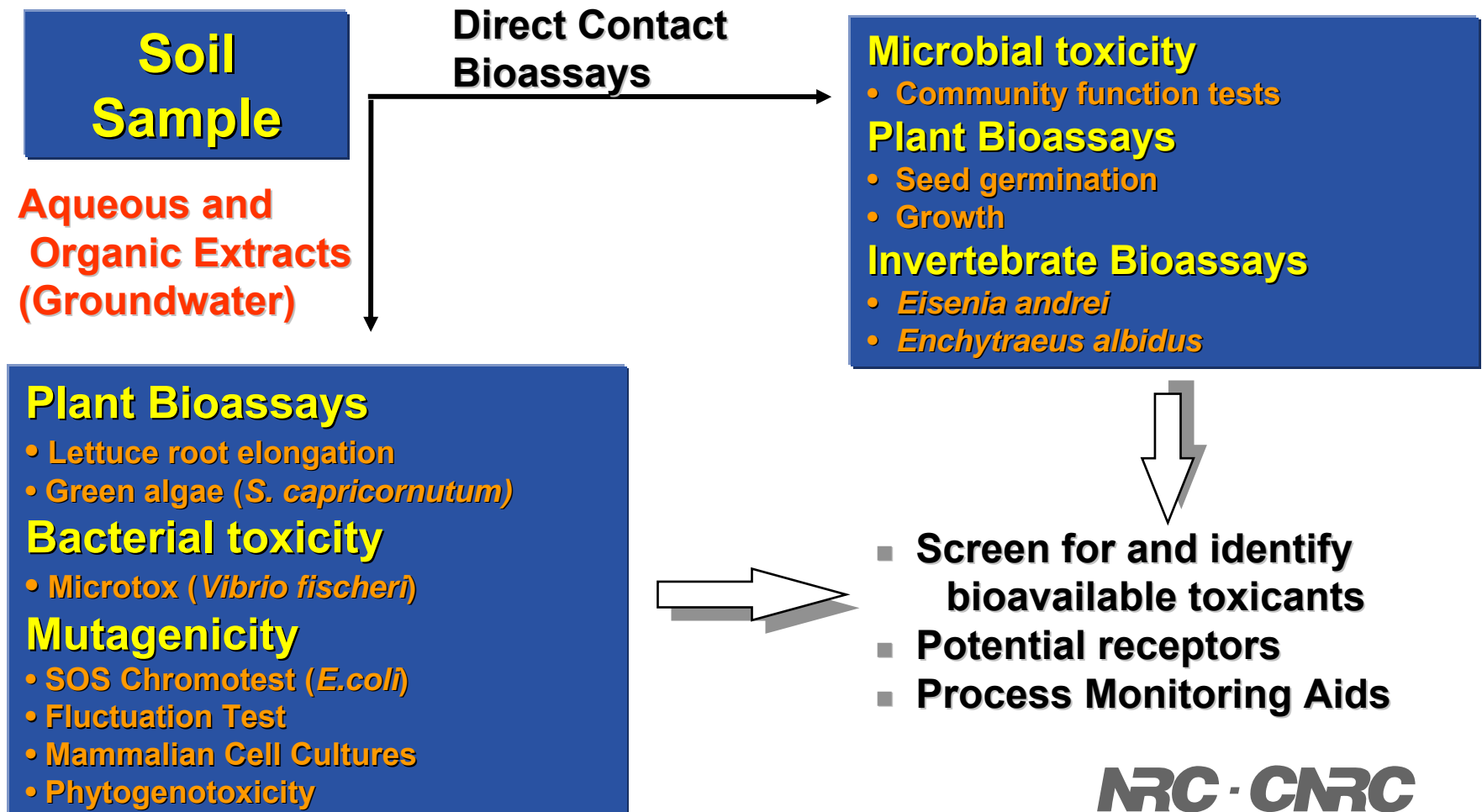
Establishing a Chain of Ecotoxicological Evidence for Potential Toxicity and Bioaccumulation



Adapted from Gunderson et al. 1997

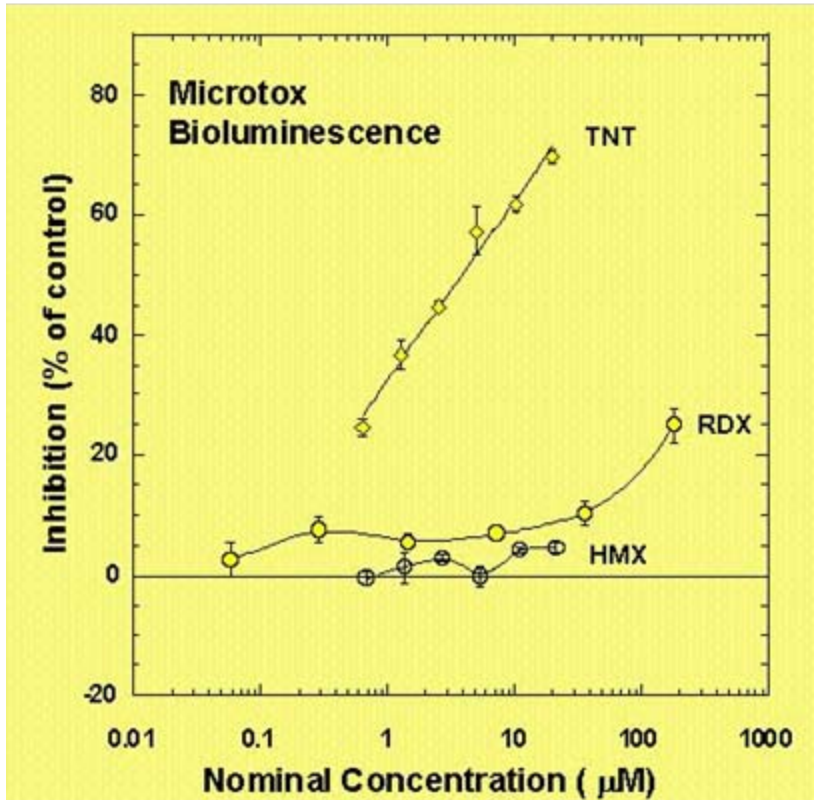


Ecotoxicological Hazard Assessment- Screening for Bioavailable Toxicants using Ecotoxicity Tests

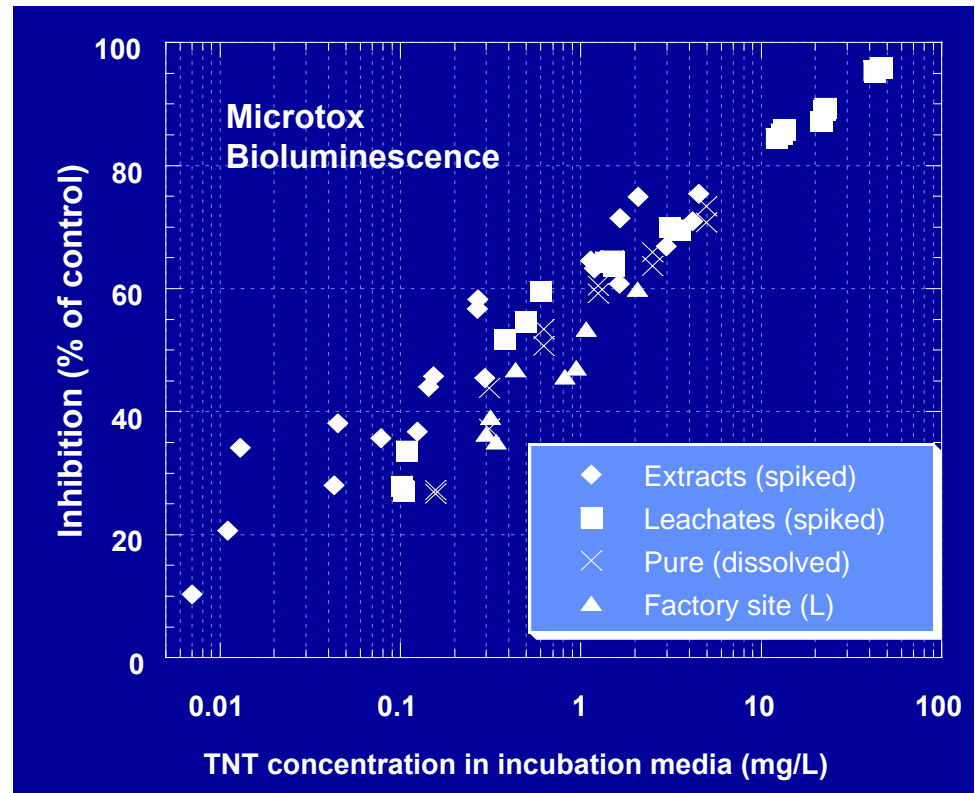


From Laboratory to Field: Toxicity to Vibrio fischeri using field soils having mixed contaminants

Pure Compounds

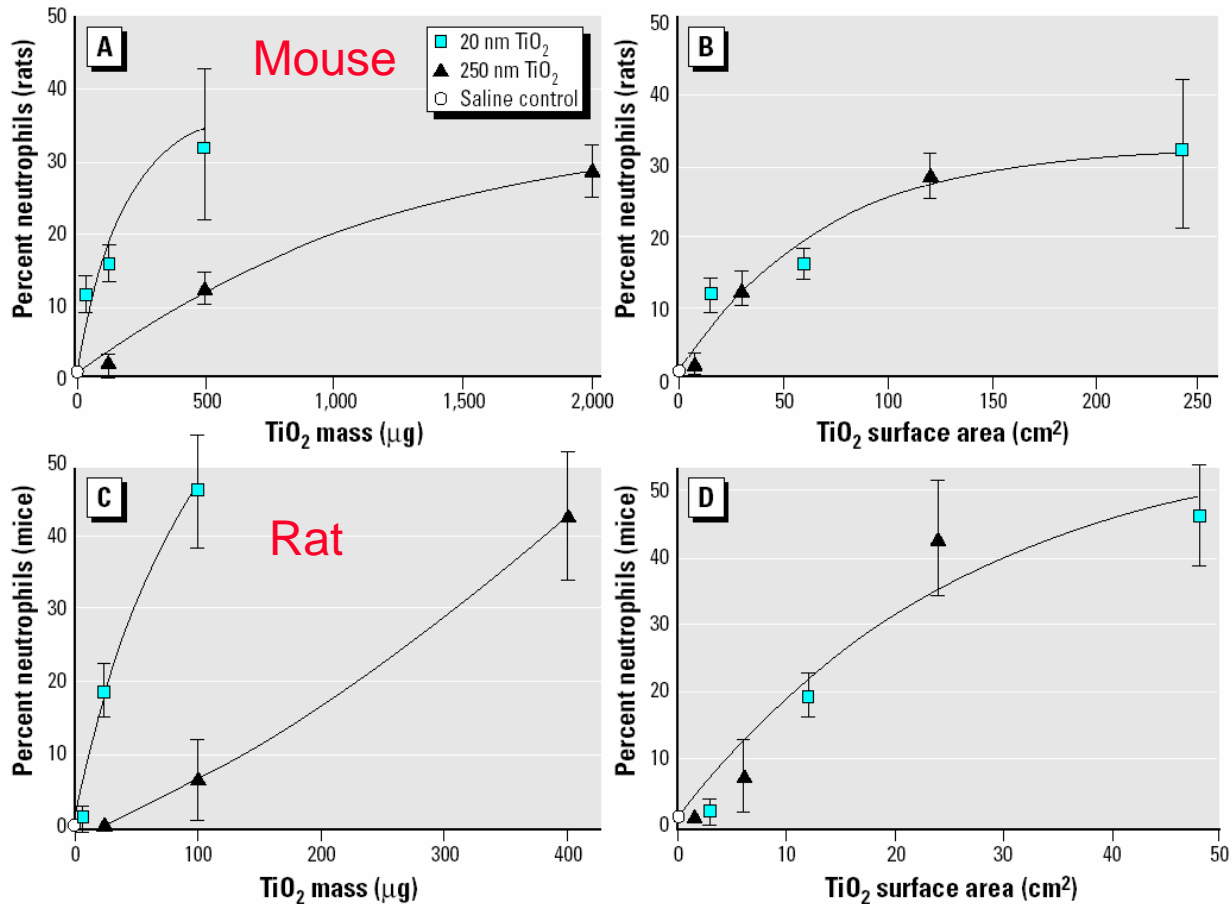


Soil leachates and extracts from a contaminated site



Dose-metric: Mass vs. Surface area

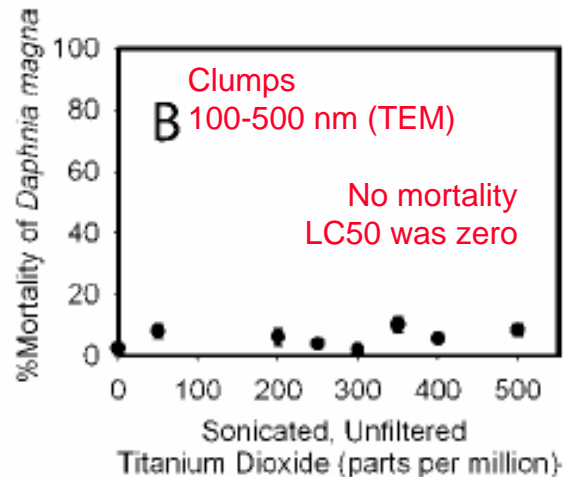
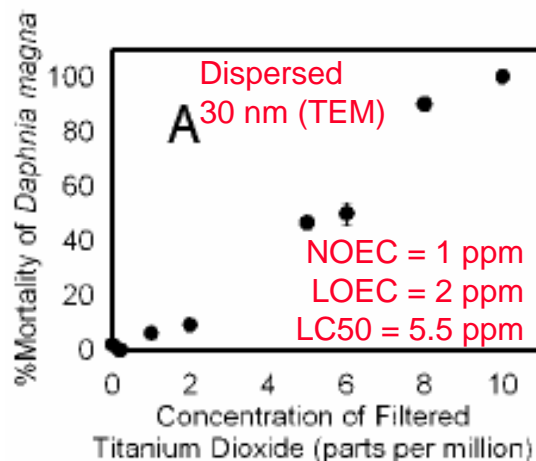
Effect of TiO₂ particle size on toxicity



- Oberdörster et al. (2005)

Sample preparation influences TiO_2 toxicity: 48-h Daphnia magna lethality test (USEPA Method 2024) (Lovern and Klaper, 2006)

- Freshwater *Daphnia* spp.,
- Filter-feeder, food chain
- 48-h acute mortality test (USEPA Method 2024)
- Titanium dioxide in deionized water \pm THF
- Passed through 0.22 μ m nylaflo filter
- Concentrations: UV spectroscopy (325-350 nm)



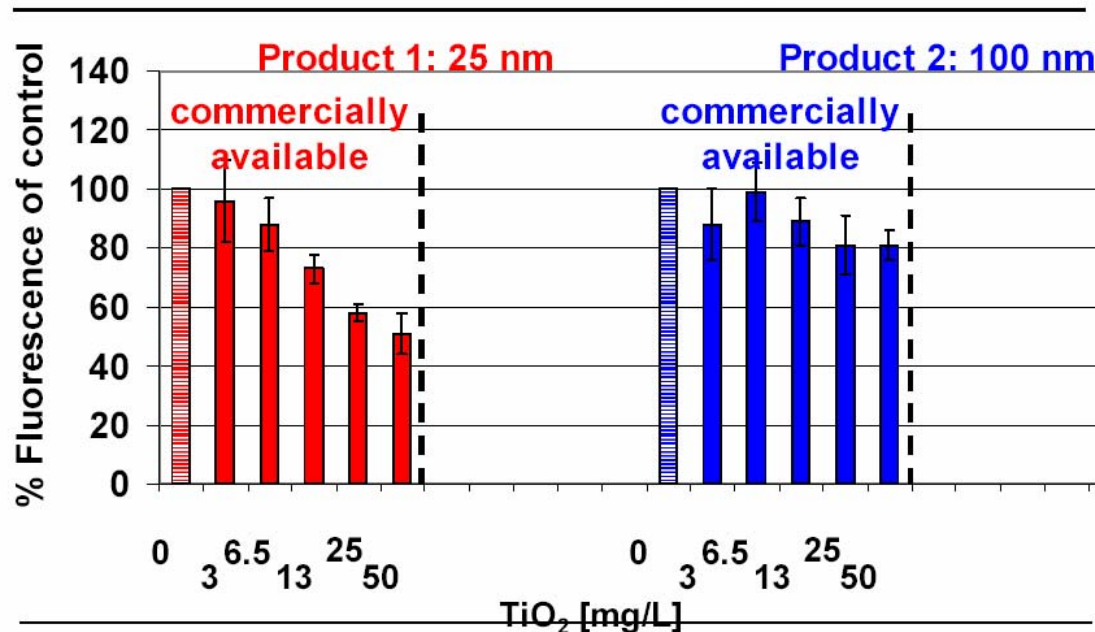
• Lovern and Klaper (2006) ETC 25: 1132-1137



Uptake in *Daphnia*, Stone 2006

Titanium Dioxide (25 nm vs 100 nm) inhibits algal growth (OECD 201)

Results: Growth inhibition test with algae (I)

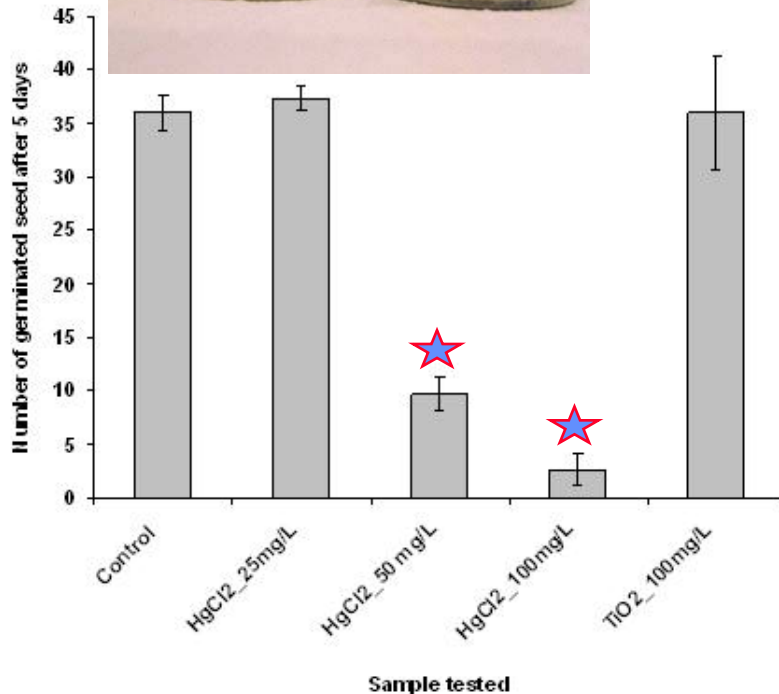


- Size-dependent inhibition of algal growth by TiO₂

Effect of TiO_2 on plants using terrestrial toxicity tests (unpublished data)

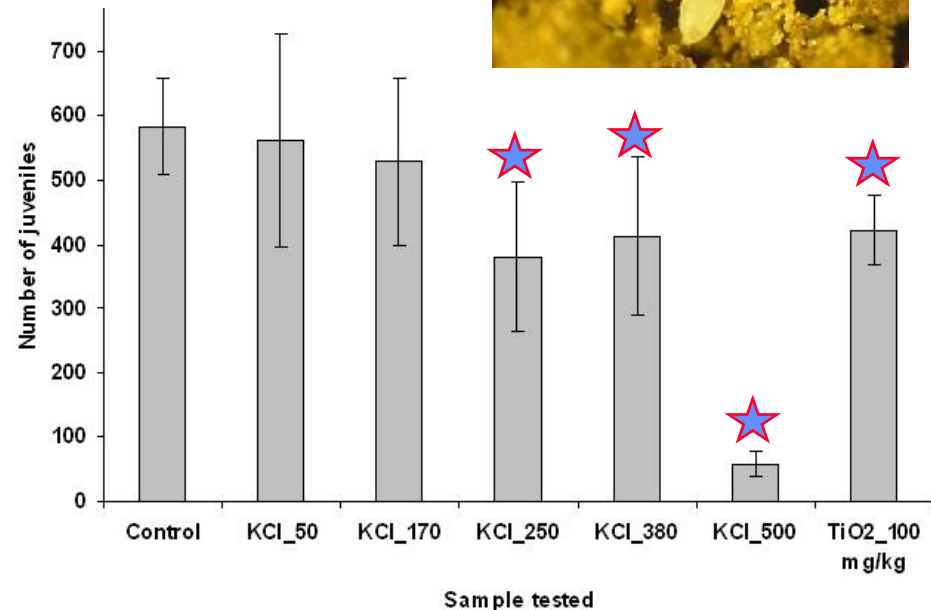


Plant seed germination (ASTM 1999, USEPA 2000)



- No apparent phytotoxicity by TiO_2

Folsomia candida - Survival and Reproduction test (ISO 11267:1998; Env Canada, 2005)



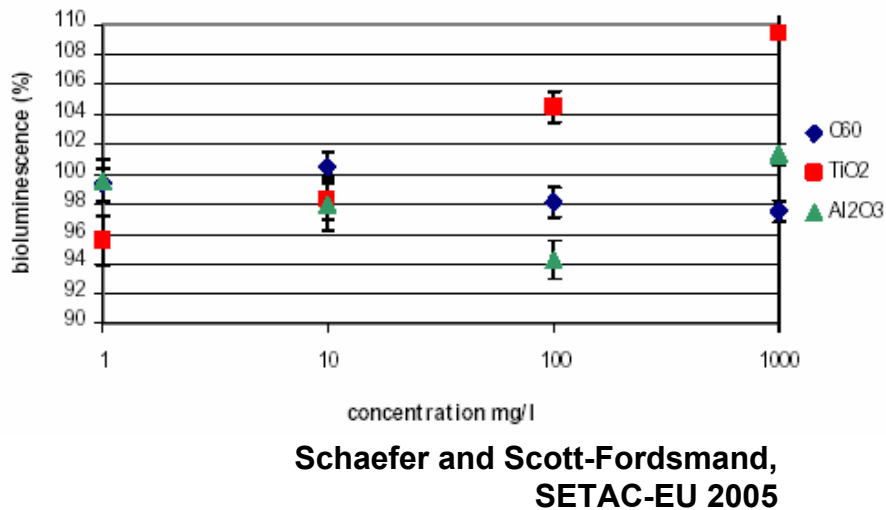
- Slight decrease in reproduction by TiO_2

NRC · CMRC

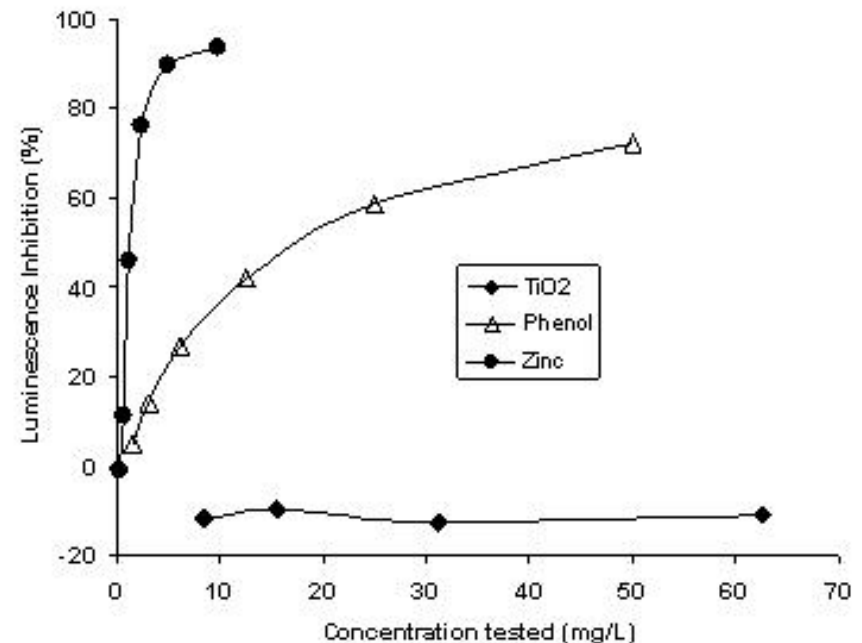
Effect of TiO_2 on Microtox test (*Vibrio fischeri*) (ISO 11348)

Fig. 1

Vibrio fischeri

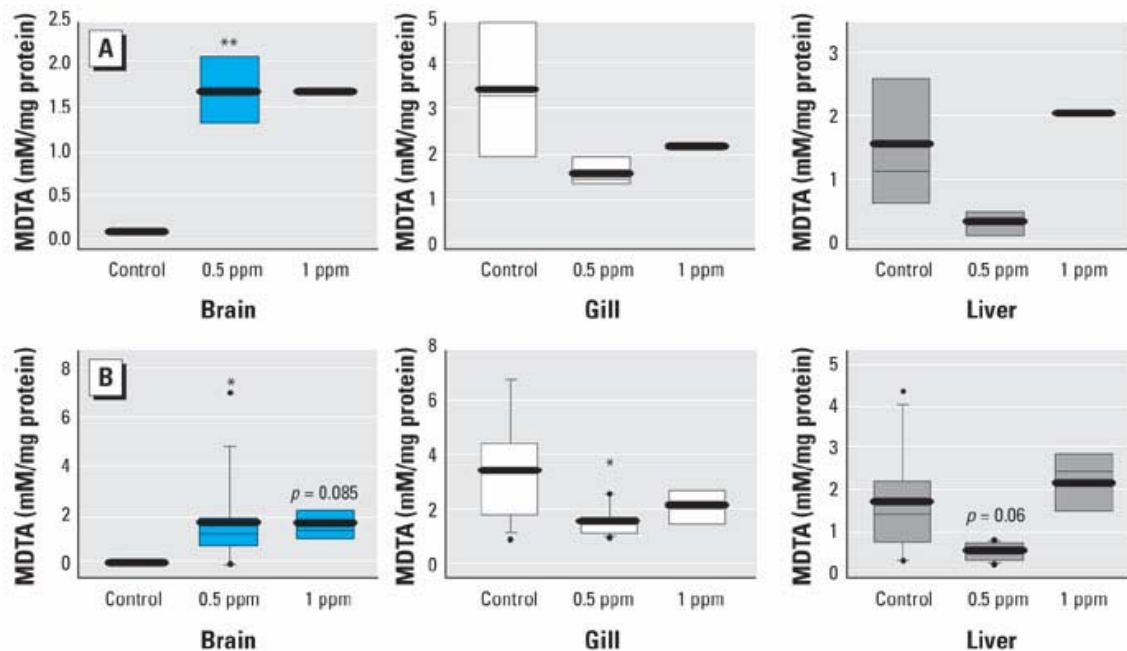


- NPs dispersed in water using sonication (15 min)
- No significant effects by TiO_2 (<100 nm) on *V. Fischeri*



- TiO_2 - sonicated 15 min
- Size to be confirmed
- No effect of TiO_2 on Microtox test

Effect of nC60 on Largemouth Bass



- *Micropterus salmoides*
- exposed to 0.5 ppm or 1 ppm of uncoated nC₆₀ (4 nm) for 48 hr.
- THF – vehicle (Rotovap); filtered nC₆₀

nC₆₀ exposure caused:

- slightly ↓ BW
- ↑ lipid peroxidation in brains at 0.5 ppm
- no clear DRC
- slight ↓ GSH in gills
- ↑ water clarity, possibly due to bactericidal activity.

Figure 1. Lipid peroxidation of brain, gill, and liver of largemouth bass after 48 hr of exposure to 0.5 or 1 ppm nC₆₀. MDTA, 1,1,3,3-tetraethoxy propane. (A) Aquarium averages. (B) Data using individual fish; in the brain, 0.5 ppm nC₆₀ caused a significant 17-fold increase in lipid peroxidation, whereas in gill and liver there is a trend for reduction of lipid peroxidation at 0.5 ppm nC₆₀. Heavy black bands represent the means; thinner lines indicate medians; boxes represent 25th and 75th percentiles; error bars indicate minimum and maximum; and circles represent outliers.

* $p < 0.05$. ** $p < 0.01$.

Bioaccumulation of NPs in fish: Environmental co-contaminants - Mixtures

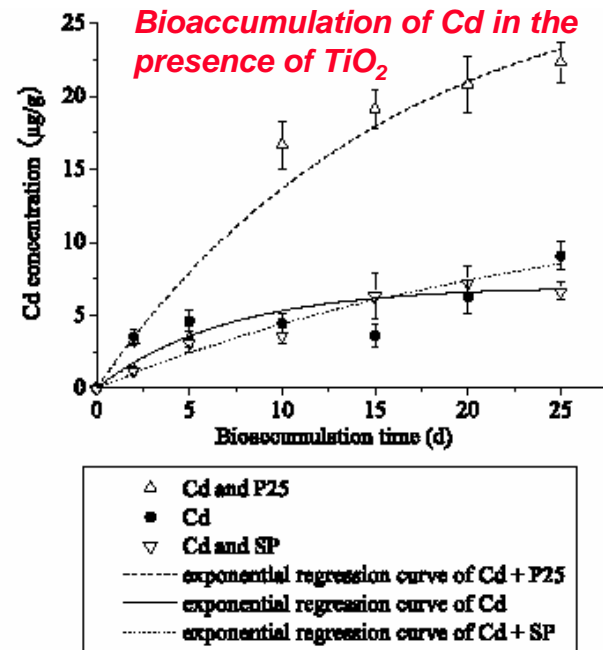
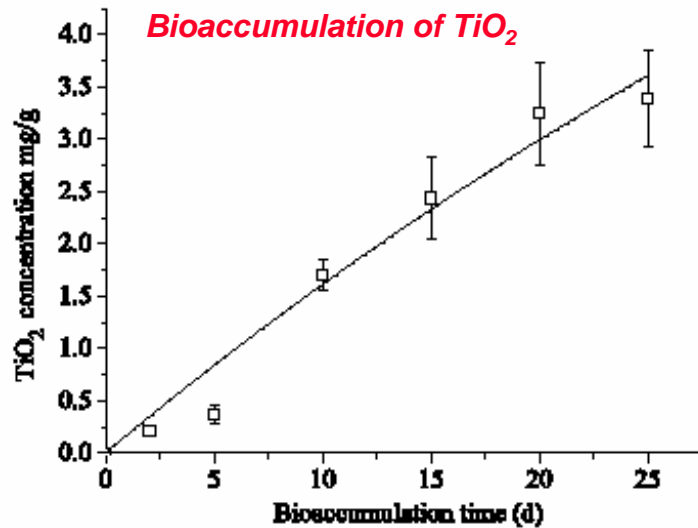


Fig. 4. The accumulation of TiO₂ nanoparticles in carp.

Zhang et al.(2007) *Chemosphere* 67: 160-166

BCFs for Cd and TiO₂ in different parts and whole body of carp at the 20th day

	Exposed media	Skin and scale	Muscle	Gills	Viscera	Whole body
Cd	Cd	4.11	0.31	33.6	213	64.4
	Cd and SP	5.41	1.72	58.3	364	88.9
	Cd and TiO ₂	11.1	3.49	152	1679	606
TiO ₂ ^a	Cd and TiO ₂	17.0	9.00	74.0	1065	325

^a The initial TiO₂ concentration 10 mg/l was used to calculate BCF for TiO₂.

What is the metric for bioaccumulation? Form?

Nano(eco)toxicological Risk Assessment: Issues and Priorities

Ecotoxicological Hazard Assessment:

- Novel Toxicity? Ecological receptors?
- Ecotoxicological effects (acute, chronic)?
- Measurement endpoints? Dose?
- Related to which physico-chemical properties?
- Physico-chemical form after bioaccumulation?

- Standardized ecotoxicity test protocols?
- Reference nanotoxicants?
- Adequate?
- Dose-metric?

Exposure Assessment:

- Environmental stability (persistence)?
- Rapid Mobility?
- Chemical and physical form in the environment?
- Exposure dose-metric? (Tissue residues?)

- Environmental Dose-metric for NPs?
- Reference nano-materials?
- Adequate?

Closing remarks

In summary,

- *Environmental health hazards related to NP exposure in air, water, and soil will challenge traditional ecotoxicological approaches.*

We should consider using.....

- *Multi-disciplinary approach (toxicologists, material scientists, and engineers)*
- *Integrated and holistic approach (LCIA) that links physico-chemical characteristics of NPs, effects in biological hierarchy, and environmental relevance.*

Thank you

- BRI-Applied Ecotoxicology Group
- Collaborators at the Biotechnology Research Institute (BRI)

